



PONTIFICIA UNIVERSIDAD CATÓLICA DE CHILE
Doctorado en Neurociencias

Tesis Doctoral

Dysfunction in striatal dopaminergic and oxytocinergic system on impulsive choice behavior

Tesis presentada a la Pontificia Universidad Católica de Chile como parte de los requisitos para optar al grado de Doctor en Neurociencias

Por

MACARENA GIPSY MORENO FERNÁNDEZ

Director de Tesis: José A. Fuentealba
Co-Director de Tesis: Georgina M. Renard

Presidente de Comisión: Katia Gysling
Evaluador Interno: Alvaro Vergés
Evaluador Externo: José L. Valdés

03 Mayo 2021



PONTIFICIA UNIVERSIDAD CATÓLICA DE CHILE
Doctorado en Neurociencias

El Comité de Tesis, constituido por los Profesores abajo firmantes, aprueba la Defensa Pública de la Tesis
Doctoral titulada:

**DYSFUNCTION IN STRIATAL DOPAMINERGIC AND OXYTOCINERGIC SYSTEMS ON
IMPULSIVE CHOICE BEHAVIOR**

Aprobación Defensa:

MACARENA GIPSY MORENO FERNÁNDEZ

Calificándose el trabajo realizado, el manuscrito sometido y la defensa oral, con nota

..... (.....)

Pontificia Universidad Católica de Chile

Dr. Mauricio Cuello
Director de Investigación y Doctorado
Escuela de Medicina
Pontificia Universidad Católica de Chile

Dr. Luis Ibáñez A.
Decano
Facultad de Medicina
Pontificia Universidad Católica de Chile

Dra. Claudia Sáez
Sub-Directora
Dirección de Investigación y Doctorado
Escuela de Medicina
Pontificia Universidad Católica de Chile

Dr. José Fuentealba
Director de Tesis
Facultad de Química y de Farmacia
Pontificia Universidad Católica de Chile

Dra. Georgina M. Renard
Co-Directora de Tesis
Facultad de Medicina
Universidad de Santiago

Dra. Katia Gysling
Profesor Evaluador Interno
Facultad de Ciencias Biológicas
Pontificia Universidad Católica de Chile

Dr. Francisco Aboitiz D.
Jefe Programa Doctorado en Neurociencias
Centro Interdisciplinario de Neurociencias
Facultad de Medicina
Pontificia Universidad Católica de Chile

Dr. José L. Valdés
Profesor Evaluador Externo
Facultad de Medicina
Universidad de Chile

Dr. Alvaro Vergés
Profesor Evaluador Interno
Facultad de Psicología

RESUMEN (Español e Inglés):

Una medida clave de la elección impulsiva implica la preferencia por un refuerzo más pequeño pero entregado de inmediato frente a recompensa más grande asociada con un retraso en su entrega. Este tipo de conducta está presente en varios trastornos neuropsiquiátricos, que incluyen: trastornos de la personalidad, trastorno por déficit atencional e hiperactividad y adicción. En los últimos años se ha propuesto que todas estas patologías comparten una alteración en la transmisión dopaminérgica mesencefálica, pero los mecanismos fisiológicos que acompañan la expresión de la elección impulsiva no están completamente aclarados. Además, hallazgos recientes sugieren que oxitocina (OXT), puede modular varios comportamientos relacionados con el sistema dopaminérgico y que existe una modulación bidireccional entre estos sistemas. El presente estudio tuvo como objetivo investigar: (1) La dinámica dopaminérgica basal en el estriado dorsolateral (SDL) presente en las diferencias individuales de las conductas de elección impulsiva. (2) El rol del sistema oxitocinérgico sobre la transmisión dopaminérgica nigro-estriatal y su modulación en las conductas de elección impulsiva.

Los resultados mostraron que: (1) La liberación de dopamina (DA) en el SDL es mayor en animales altamente impulsivos que en animales menos impulsivos, a pesar de que los niveles extracelulares de DA y la actividad del transportador de DA (DAT) no varían entre los grupos. (2) La activación bilateral de OXT-R en sustancia nigra pars compacta (SNpc), induce una disminución en la conducta impulsiva, principalmente en animales clasificados previamente como altamente impulsivos. Este hallazgo muestra que la liberación de DA en el SDL está aumentada en las ratas clasificadas como altamente impulsivas, lo que sugiere que la vía nigro-estriatal hiperactiva contribuye a la elección impulsiva. Por otro lado, la activación del OXT-R en la SNpc influye en la toma de decisiones relacionadas con el retraso, mediante cambios sutiles en la transmisión dopaminérgica estriatal. Estos resultados proponen una nueva línea de investigación sobre los mecanismos neurobiológicos que subyacen a la posible interacción entre los sistemas de DA y OXT, y su rol en conductas de elección impulsiva presentes en patologías asociadas a una alteración de estos dos sistemas en la vía nigro-estriatal.

A key measure of impulsive choice involves a preference for smaller but immediately delivered reinforcements versus larger rewards associated with delayed delivery. This behavior is present in several neuropsychiatric disorders, including personality disorders, attention deficit disorder, hyperactivity, and addiction. In recent years it has been proposed that all of these pathologies share an alteration in mesencephalic dopaminergic transmission, but the physiological mechanisms that accompany the expression of impulsive choice are not fully clarified. Moreover, recent findings suggest that oxytocin (OXT) can modulate several behaviors related to the dopaminergic system and a bidirectional modulation between them. The present study aimed to investigate: (1) The basal dopaminergic dynamics in the dorsolateral striatum (DLS) present in individual differences of impulsive choice behaviors. (2) The role of the oxytocinergic system on nigro-striatal dopaminergic transmission and its modulation in impulsive choice behaviors.

The results showed that: (1) Dopamine (DA) release in DLS is higher in highly impulsive animals (HI) than in low impulsive animals (LI). However, DA extracellular (DA_{ext}) levels and transporter activity (DAT) do not vary between groups. (2) The bilateral activation of OXT-R in substantia nigra pars compacta (SNpc) induces a decrease in impulsive behavior, in animals previously classified as HI. Moreover, the OXT-R activation in SNpc induces a modest increase in DA_{ext} in DLS. This finding shows that DA release is augmented in the DLS of rats classified as HI, suggesting that hyper-activity nigro-striatal pathway contributes to impulsive choice. On the other hand, the activation of the OXT-R in the SNpc influences delay-related decision making by subtle changes in the striatal dopamine transmission. These results propose a new line of research on the neurobiological mechanisms underlying the possible interaction between the OXT and DA systems and their role in impulsive choice behaviors present in pathologies associated with altering these two systems in the nigrostriatal pathway.

Dedicatoria

La presente tesis está dedicada a todas las personas que de una u otra forma estuvieron y son parte de mi crecimiento como persona y como científica.

Especialmente quiero dedicar este trabajo y todos los avances en mi carrera científica a mi Padre, que por circunstancias de la vida no alcanzaste a verme crecer y avanzar en mi carrera, pero de igual forma siempre has estado presente y desde el amor que dejaste en mi he podido sacar fuerzas para siempre continuar con una sonrisa en mi rostro y no perder la alegría de hacer lo que me gusta... te amo infinito y muchísimas gracias.

A mi Madre, por siempre estar, por siempre tener esa palabra de aliento o ese regaloneo necesario para continuar.

A mi Guelli, lamentablemente no alcanzaste a verme terminar, pero tengo tu dulce mirada en mi corazón que reconforta el alma.

A mi Teche, fuiste parte no solo de este proceso, sino de muchos otros en mi vida y sé que desde donde estás debes estar orgulloso de tu pequeña.

A mi familia (hermana, sobrina, mi puntito, tíos/as, primos/as y amigos/as), por el cariño, apoyo y fuerzas que siempre me entregaron, los quiero infinito.

Y finalizo con mi compañero de aventuras y novio Camilo, por estar en las buenas y malas, por el cariño, apoyo y paciencia infinita sobre todo durante el último tiempo, amor infinito hacia ti.

***“You have to act as if it were possible to
radically transform the world. And you have
to do it all the time.”***

Angela Davis

Acknowledgements

Firstly, I want to thank my advisors José Fuentealba, Ph.D and Georgina Renard, Ph.D, for the patience and all the advices and kind critics in the numerous meeting in all these years. I am sure that this thesis would not be the same without your help.

I would like to thank Dr. Francisco Aboitiz, Ph.D and Claudia Sáez, Ph.D, for all de support during the last time.

I want to thank Stan.B Floresco Ph.D, and his team-lab, for allowing me to do my doctoral internship in his laboratory, it was a great experience, of much learning.

I would also like to thank my thesis committee: Katia Gysling, Ph.D; Dr Alvaro Verges, Ph.D and José Luis Valdés, Ph.D. For all their support and willingness to always clarify my doubts or discuss my results, it is an-honor to have them as commission of my doctoral thesis.

I would like to thank my fellow lab-mates: Enzo, Rafa, Victor, Consuelo, for the support, discussions and knowledge learned.

In addition, I want to thank the lab-women power + Max: Geo, Maca, Vale, Fran, Jenn, Dani and Clau, for the interesting nerd or non-nerd conversations, laughter and anecdotes on the USACH-hill.

I want to thank my friends: Catherine Perez, Maca Gárate, Melany Garrido, Carlos Saldías, Christian Lamas, Juan Ahumada and others. Thanks for the support, laughs and good times with you.

I thank my colleagues from the school of psychology UBO: Vivi, Karla, Pablo, Irany, Leslie, Marcelo, Javi and Lautaro. Thanks for the support, for believing in me, I am lucky to be part of the team.

Finally, I thank to fellowship ANID-PCHA #21150508, FONDECYT project 1141088, for the financial support in this research.

Table of contents

1	INTRODUCTION	9
1.1	IMPULSIVITY	9
1.2	NEUROANATOMICAL CIRCUITS.....	10
1.3	MONOAMINE NEUROTRANSMITTER SYSTEMS.....	12
1.4	DOPAMINE SYTEM	12
1.5	ROLE OF DOPAMINE AND OXYTOCIN ON NEUROPSYCHIATRIC DISORDERS	14
1.6	OXYTOCINERGIC SYSTEM	¡ERROR! MARCADOR NO DEFINIDO.
1.7	INTERACTION BETWEEN OXYTOCINERGIC AND DOPAMINERGIC SYSTEM.....	16
2	HYPOTHESIS	19
2.1	CHAPTER 1:	19
2.2	CHAPTER 2:	19
3	OBJECTIVES	20
3.1	CHAPTER 1:	20
3.2	CHAPTER 2:	21
4	CHAPTER 1: HIGH IMPULSIVE CHOICE IS ACCOMPANIED BY AN INCREASE IN DOPAMINE RELEASE IN RAT DORSOLATERAL STRIATUM	22
4.1	ABSTRACT	24
4.2	INTRODUCTION.....	25

4.3	EXPERIMENTAL PROCEDURES	29
4.3.1	<i>Animals</i>	29
4.3.2	<i>Apparatus</i>	30
4.3.3	<i>Procedures</i>	31
4.3.4	<i>Surgery and no-net-Flux Microdialysis</i>	33
4.3.5	<i>Histology</i>	34
4.3.6	<i>Analysis of dialysate dopamine</i>	34
4.3.7	<i>Data Analysis</i>	35
4.3.8	<i>No-Net-Flux analysis</i>	36
4.3.9	<i>Statistical Analyses</i>	37
4.4	RESULTS	38
4.4.1	<i>Individual differences in basal level of impulsive choice</i>	38
4.4.2	<i>Animals classified as high impulsive in DDT showed higher DA C_{ext} in DLS</i> 38	
4.4.3	<i>Correlation between impulsive choice levels and DA dynamic in the DLS</i> 39	
4.5	DISCUSSION	40
4.6	ACKNOWLEDGMENTS	45
4.7	REFERENCES	46
5	CHAPTER 2: EFFECT OF OXT-R ACTIVATION ON THE NIGROSTRIATAL PATHWAY DURING DELAY DISCOUNTING TASK, A STUDY OF IMPULSIVE CHOICE BEHAVIORS	64
5.1	ABSTRACT	66
5.2	INTRODUCTION	68

5.3	METHODOLOGY.....	72
5.3.1	<i>Animals</i>	72
5.3.2	<i>Behavioral Testing</i>	73
5.3.3	<i>Delay discounting analysis</i>	76
5.3.4	<i>Conventional Microdialysis</i>	78
5.3.5	<i>Histology</i>	79
5.3.6	<i>Data Analysis</i>	80
5.4	RESULTS.....	81
5.4.1	EFFECT OF SNPC ADMINISTRATION OF WAY267464 ON IMPULSIVE CHOICE DURING DELAY DISCOUNTING TASK	81
5.4.2	<i>The effect of bilateral OXT-R activation in SNpc depends on basal impulsivity</i>	82
5.4.3	<i>Effect of non-peptide specific OXT-R agonist (WAY267464) administration in SNpc on DAext levels in DLS</i>	83
5.5	DISCUSSION	84
5.6	ACKNOWLEDGMENTS	89
5.7	REFERENCES	90
1.	FIGURES.....	104
6	GENERAL DISCUSSION	112
7	GENERAL CONCLUSIONS	119
8	SUPPLEMENTARY INFORMATION.....	121
8.1	PROPOSED MODEL.....	121

9	REFERENCES.....	122
----------	------------------------	------------

Table of Figure

CHAPTER 1: HIGH IMPULSIVE CHOICE IS ACCOMPANIED BY AN INCREASE IN DOPAMINE RELEASE IN RAT DORSOLATERAL STRIATUM

Figure 1: Delay discounting task design.....	59
Figure 2: Anatomical placements of the microdialysis probe in DLS.....	60
Figure 3: Individual differences in basal level of impulsive choice.....	61
Figure 4: Animals classified as high impulsive in DDT showed higher DA release in DLS.....	62
Figure 5: Correlation between impulsive choice level measured by AUC and basal dynamic of DA in DLS.....	63

CHAPTER 2: EFFECT OF OXT-R ACTIVATION ON THE NIGROSTRIATAL PATHWAY DURING DELAY DISCOUNTING TASK, A STUDY OF IMPULSIVE CHOICE BAHAVIORS

Figure 1: Experimental design of micro-infusion of WAY267 in SNpc and histology scheme.....	104
Figure 2: Effect of bilateral OXT-R activation in SNpc during DDT.....	105
Figure 3: Differential effect of bilateral OXT-R activation in SNpc by microinjection of WAY267464 (3 ug/ul) and saline solution	106

Figure 4: Representative scheme of probe locations during the microdialysis protocol.....	108
Figure 5: The effect of perfusing the SNpc with a high concentration of Potassium (70mM) on DA_{ext} in the nigrostriatal pathway.....	109
Figure 6: Effect of OXT-R activation in SNpc on DA_{ext} levels in DLS and SNpc	110
Figure Supplementary 1: DA basal dynamic activity present in impulsive choice behavior and the effect of OXT-R activation in SNpc during DDT.....	120

List of abbreviations

aCSF: Artificial cerebrospinal fluid

ADHD: Attention deficit hyperactive disorder

CNS: Central Nervous System

D2-R: Dopamine receptor, type 2

D1-R: Dopamine receptor, type 1

DA: Dopamine

DAG: Diacylglycerol

DAT: Dopamine transporter

DLS: Dorsal lateral Striatum

HPLC: High-performance liquid chromatography

IP3: Inositol Triphosphate

LY368,899: Potent, non-peptide oxytocin receptor antagonist

NAcb: Nucleus Accumbens

NAcbC: Nucleus Accumbens Core

nM: Nanomolar

OFC: Orbitofrontal Cortex

OXT: Oxytocin

OXT-R: Oxytocin receptor

PCA: Ácido perclórico

PFA: Paraformaldehyde

PFC: Prefrontal Cortex

PKC: Protein kinase C

PLC: Phospholipase C

PND: Postnatal day

PVN: Paraventricular Nucleus

Sec: Seconds

SN: Substantia nigra

SNpc: Substantia nigra pars compacta

SON: Supraoptic Nucleus

VGCCs: Voltage-gated calcium channel

VMAT: Vesicular monoamine transporter

VTa: Ventral tegmental area

WAY267464: Potent, non-peptide oxytocin receptor agonist

μl: Microlitre

μM: Micromolar

1 Introduction

1.1 Impulsivity

Impulsivity is a multifactorial construct that can be adaptive or maladaptive depending on the consequences resulting from it (Eysenck and Eysenck, 1977; Amorim Neto and True, 2011; Strickland and Johnson, 2020). Maladaptive impulsivity, in particular, has been classically described as the predisposition to inappropriate, badly planned decisions with negative consequences. This behavior is subdivided into two broad categories: impulsive choice and impulsive action (Winstanley et al., 2010). Impulsive action refers to a failure in inhibiting an inappropriate response to a stimulus (Schachar et al., 2007; Eagle et al., 2008). On the other hand, impulsive choice is characterized by impulsive decision-making caused by poor consideration of future behavioral consequences (e.g., preferences in choosing a small reward delivered immediately over larger rewards delivered with a delay) (Evenden and Ryan, 1996; Cardinal et al., 2001; Dalley et al., 2011; Hamilton et al., 2015). It is proposed that both types of impulsive behaviors are characterized by high individual variability, which is determined by environmental, social and physiological factors (Arce & Santisteban, 2006; Hamilton et al., 2015; C. Liu et al., 2016). The focus of our research is on impulsive choice, as it has been observed in several neuropsychiatric disorders, including personality disorders (Perry & Körner, 2011), mood disorders (Lombardo et al., 2012), attention deficit hyperactivity disorder (ADHD) (Avila et al., 2004) and addiction (Ersche et al., 2012), pointing to its possible role in the pathogenesis of these neuropathologies (Renda et al., 2014; Rung et al., 2018).

1.2 Neuroanatomical circuits

Research in humans and animals have associated impulsive choice with functional integration of the orbitofrontal cortex (OFC) and the nucleus accumbens core (NAcbC) (Dinn et al., 2001; Winstanley et al., 2004), proposing that changes in the subcortical-cortical connections are related to this behavior. Additionally, it can be interpreted that changes in patterns of functionality linked to a decrease in dopaminergic system activity contribute to the induction of high levels of impulsive choice (Brooks & Berns, 2013; Kravitz et al., 2015; Sellitto et al., 2011). Moreover, studies using functional magnetic resonance imaging (fMRI) showed that dysregulation in functional connectivity between cortical and putamen regions was associated with high impulsivity levels in subjects (Carmona et al., 2009; Wilbertz et al., 2012). This change in functionality has been corroborated in studies using the Delay Discounting Task (DDT), where the subject or animal must choose between a small reinforcer delivered immediately or a larger reinforcer delivered after a delay. These studies showed that the selection of large reinforcement is associated with an increase in putamen activity (Wittmann et al., 2007).

Interestingly, animals studies have suggested that prefrontal cortex (PFC) and NAcb contribute differentially to decision making, giving further insight into the dynamic interactions between the prefrontal and striatal systems (St. Onge et al., 2012). The subcortical circuit provides a visceral and intuitive control toward options that have larger rewards, in comparison with the PFC, that appears to have a supervisory role, monitoring the frequency of rewarded actions over time. This could be associated with fluctuations in tonic dopamine (DA) level, facilitating the implementation of decision policies (St. Onge

et al., 2012; St Onge et al., 2012b). Following this, newer studies have started to reveal that the dorsolateral striatum (DLS) or the putamen in humans, conventionally known for its participation in motor functions (Kobayashi et al., 2013), has also been associated with cognitive processes (Balleine et al., 2007; Hariri et al., 2006) hypothesizing that there exists a connection between drug-seeking habits and impulsive behaviors, and that they have a common biological substrate which is changes in the striatal DA levels (Besson et al., 2013; Jentsch & Taylor, 1999). Using DDT, lesions, and pharmacology techniques in animal models, it has been proposed that high level of impulsivity is associated with a decrease in DA level in the DLS. For example, Tedford., et al (2015) showed that the bilateral retrograde lesion with 6-hydroxydopamine (6-OHDA) in dopaminergic terminals in the DLS induces an increase in impulsivity choice during DDT, which is in concordance with studies with humans that showed that DA in DLS participates and is necessary for decision-making (Hong et al., 2013; McHugh et al., 2013). This dopaminergic hypofunction could be associated with an increase in dopamine transporter (DAT) activity, which is mainly mediated by presynaptic mechanisms related with D₂-R presynaptic activation, playing a key role in the regulation of DA neuronal activity and the control of synthesis, release, and uptake of DA (Baarendse & Vanderschuren, 2012). In contrast, the report of Magnard et al., 2018 showed that anterograde bilateral lesion with 6-OHDA in substantia nigra pars compacta (SNpc), does not induce changes in the impulsive choice behavior during DDT performance. This is interesting because both studies focus on the role of DLS and dopaminergic system in impulsive choice. However, to date, the changes in the endogenous DA dynamic into the nigrostriatal pathway contributing to impulsive choice behavior are unknown.

1.3 Monoamine neurotransmitter systems

Prior research has suggested that monoamine dysfunctional signaling, particularly in the dopaminergic and noradrenergic systems, contribute to the generation of impulsive behaviors (Humpston et al., 2013; Yates et al., 2014). Studies using radioligands with positron emission tomography (PET) in humans have identified a decrease in the availability of D₂ and/or D₃ receptors in striatal regions in impulsive subjects (Ghahremani et al., 2012; B. Lee et al., 2009). Moreover, studies in animal models using *in situ* hybridization have shown significant differences in mRNA expression for DA receptors in striatal regions (Simon et al., 2013). Finally, it has been proposed that high levels of impulsivity during DDT (linked to salience and experience to a known reinforcement) are associated with decreased levels of DA, probably mediated by presynaptic mechanisms. It should be noted that these mechanisms have not been well determined (see, for instance: Costa et al., 2014; Eubig et al., 2014; Smith et al., 2016; Tedford et al., 2015).

1.4 Dopamine system

The DA system has been classically linked to reward behavior, pleasure, euphoria, motor control, and impulsivity. DA is synthesized in two nuclei, the ventral tegmental area (VTA) and SNpc (C. R. Lee & Tepper, 2009; Roeper et al., 2013). The system consists of dopaminergic neurons, receptors expressed both at a pre- and a postsynaptic level and a membrane transporter, which contribute to the control and the balance of dopaminergic activity in several regions of the central nervous system (CNS) (Missale et al., 1998). DA

receptors share a high degree of homology. These are G protein-coupled metabotropic receptors, with seven transmembrane domains, but have different pharmacological properties: receptors belonging to the D₁ family are formed by D₁ and D₅ receptors and activate a family of proteins G_s linked to the activation of intracellular cascades mediated by adenylate cyclase. They contribute to the intracellular production of the second messenger cyclic adenosine monophosphate (cAMP) (Hernández-López et al., 1997; Missale et al., 1998; Surmeier et al., 2007). Additionally, these receptors are expressed exclusively in postsynaptic neurons; for example in medium spiny neurons (MSN) in the DLS (Centonze et al., 2003). On the other hand, D₂ family receptors including D₂, D₃, and D₄, are coupled to G_i protein, which is characterized by inhibiting adenylate cyclase (Fazio et al., 2011; Sokoloff et al., 2006). The role of D₂ and D₃ receptors are more complex than D₁ receptors, as they are expressed both at a pre- and a postsynaptic level, and are associated to the control of DA release mediated by activation of the presynaptic receptors (Barton et al., 1991; Missale et al., 1998). Another protein that helps regulating the levels of this neurotransmitter is the dopamine transporter (DAT), which is a membrane protein that is expressed pre-synaptically. Its function is to control the intensity of DA-mediated signaling by recapturing the neurotransmitter released by presynaptic neurons (Zhang et al., 2009). DAT belongs to a family of transporters which depends on the activity of the Na⁺/Cl⁻ pump (Kanner, 1994; Zhang et al., 2009). There is evidence to postulate a functional interaction between the activation of presynaptic D₂ receptors and increased expression of DAT in the cell membranes, contributing to improved clearance of DA in the synaptic space (Bolan et al., 2007).

1.5 Oxytocinergic system

OXT is a neuropeptide which is synthesized in a group of neurons located in the paraventricular (PVN) and supraoptic (SON) nucleus of the hypothalamus. Axons of these neurons project and move the OXT to the anterior pituitary, where it is finally released into the bloodstream as a hormone which functions in both central and peripheral systems (Loup et al., 1991; Zink & Meyer-Lindenberg, 2012). OXT projections have been described to reach extra-hypothalamic regions, like the prefrontal cortex, VTA, NAc, hippocampus, amygdala, and SN, the latter being one of our regions of interest (Dogterom et al., 1978; Mai, Berger, & Sofroniew, 1993). This neuropeptide is classically known for its actions in peripheral targets, for example, its role in uterine contraction and milk ejection during childbirth. In turn, at the central level, OXT has a role both as a neurotransmitter and/or as a neuromodulator, acting mainly through dendritic release (Leng & Ludwig, 2008; Neumann et al., 2012) regulating cognitive processes and complex social behaviors (Insel, 2010). Moreover, there is a growing number of studies showing that signaling pathways mediated by OXT are altered in neuropsychiatric disorders associated with several social deficits (Aspé-Sánchez et al., 2015; Meyer-Lindenberg et al., 2011).

There are two types of oxytocin receptors (OXT-R) located both in the central and the peripheral systems. They are a family of metabotropic receptors, which have seven transmembrane domains that acts through protein-coupled type G_q or G_i (Ebstein et al., 2012; Wilkie et al., 1991). The OXT-R is expressed in different areas of the CNS. For example, the OXT-R coupled to protein G_q is expressed in the SN, VTA, NAc, anterior

cingulate cortex, central and basolateral amygdala, medial preoptic area, anterior and ventromedial hypothalamus, among others (Boccia et al., 2013; Rault et al., 2013; Skuse, Gallagher, & Frith, 2009) while the OXT-R type G_i is abundantly expressed in the spinal cord, the optic chiasm, frontal and occipital cortices, caudate and putamen (Milligan, 1993). When OXT binds to the OXT-R type G_q , it initiates an intracellular pathway mediated by phospholipase C (PLC), generating inositol triphosphate (IP3) and 1,2 diacylglycerol (DAG). IP3 mobilizes calcium from intracellular stores and DAG activates protein kinase C (PKC) which contributes to phosphorylation protein. The activation of these receptors results in a depolarization induced by decrease inward rectifying K^+ (IRK) currents (Berrada et al., 2000; Gravati et al., 2010; Thibonnier & Schork, 1995). On the other hand, the OXT-R G_i inhibits adenylate cyclase activity, in turn decreasing the concentration of cAMP, which activates phosphatidylinositol-4,5-bisphosphate 3-kinase (PI3K) and increases IRK currents (Garibay et al., 1991; Gravati et al., 2010).

1.6 Role of dopamine and oxytocin on neuropsychiatric disorders

New research is proposing the use of neuropeptides such as oxytocin (OXT), as a neuromodulator of DA transmission. Studies also support its potential role as a therapeutic target for neuropsychiatric diseases associated with diminished social behavior (Baskerville & Douglas, 2010; Love, 2014; McGregor & Bowen, 2012; Tops et al., 2014), and as a treatment for neuropsychiatric disorders, such as depression, autism, addiction and others (McGregor & Bowen, 2012; Meyer-Lindenberg et al., 2011; Peters et al., 2013).

Most studies on OXT and its neuromodulatory role have involved models of addiction, in which it has been shown that increased oxytocinergic tone (the peripheral or central application), decreases drug seeking and attenuates anxiety levels (Bowen et al., 2011; McGregor & Bowen, 2012). Most studies on OXT and its neuromodulatory role have involved models of addiction. It has been shown that increased oxytocinergic tone (the peripheral or central application) decreases drug seeking and attenuates anxiety levels (Bowen et al., 2011; McGregor & Bowen, 2012). On the other hand, recent studies showed that the intranasal OXT administration moderated impulsive behavior in subjects with a single nucleotide polymorphism (SNP) rs2254298 genotype (Bozorgmehr et al., 2020). In line with these, the intracerebroventricular administration of OXT reduces risky decisions in animal models (Tapp et al., 2020). Nevertheless, the neurobiological mechanism that underlies the role of the OXT in impulsive behavior not fully understood.

1.7 Interaction between oxytocinergic and dopaminergic systems

It is known that there is a bidirectional modulation between dopaminergic and oxytocinergic systems (Moos & Richard, 1982; Shahrok et al., 2010). It has also been shown that DA is capable of modulating the oxytocinergic pathway (Moos & Richard, 1982). For example, at a neurophysiological level, it has been found that the effect of intraventricular injections of DA induced an increase in the electrical activity of oxytocinergic neurons in PVN (Moos and Richard 1982). It has also been shown that these hypothalamic neurons express both DA and OXT receptors and form heterodimers in

hypothalamic and striatal regions (Baskerville & Douglas, 2008; Romero-Fernandez, et al., 2013). Finally, it has been proposed that both dopaminergic and oxytocinergic pathways converge in the PFC, striatum and tegmental areas (Succu et al., 2007; Chang et al., 2014). Additionally, a dysfunction between dopaminergic and oxytocinergic transmission is present in a variety of neuropsychiatric disorders, such as schizophrenia, autism, ADHD and addiction (Baskerville & Douglas, 2010; Park et al., 2010; Rosenfeld, Lieberman, & Jarskog, 2011) which are disorders associated with impulsive behavior.

In recent years, there has been an increasing interest in the use of OXT as a new treatment for many neuropsychiatric disorders (McGregor & Bowen, 2012; Meyer-Lindenberg et al., 2011; Peters et al., 2013), but its role in impulsive choice (a feature present in several of these disorders) is not fully understood. There is evidence that the oxytocinergic system is able to activate mesocorticolimbic and nigrostriatal pathways (Y. Liu & Wang, 2003; Young & Wang, 2004, Xiao et al, 2017). Similarly, it has been showed that the central and peripheral OXT administration attenuates withdrawal effects induced by morphine, ethanol, and cocaine, preventing the generation of tolerance and stereotypical behaviors caused by repeated use of these drugs in animals models of addiction (Bentzley, Jhou, & Aston-Jones, 2014; Kovács et al., 1998; Peters et al., 2013). Clinical studies using fMRI techniques have shown that the administration of OXT induces an increase in the activity of VTA (Schott et al., 2008; Chang et al., 2014). Additionally, it has been shown in preclinical studies, that OXT perfusion in the VTA induces an increase in DA levels in the NAcB (Shahrokh et al., 2010). The mechanisms underlying modulation offered by OXT, and its role in the regulation of rewarding properties of drugs of abuse are not clear yet. Clinical experiments using single-photon emission computed tomography (SPECT)

and genotyping methods have indicated that there exists a correlation between OXT-R gene polymorphism (rs53576) and striatal DAT availability (Chang et al., 2014). Using SPECT, Love et al. 2012 showed that humans with anxiety phenotypes have a variation in the OXT-R gene associated with the presence of various polymorphisms, for example, the rs53576, that could be influencing the striatal DA activity, increasing the evidence of the interactions between oxytocinergic and dopaminergic systems.

Concluding, this research is presented in two chapters:

The first chapter aims to evaluate the striatal dopaminergic dynamic in impulsive choice in animals using the DDT paradigm. We particularly want to test the contribution of the DA basal dynamic in DLS, and how it is correlated with low or high impulsive behavior. For this, we used the quantitative neuro-chemical technique: no-net-flux microdialysis *in vivo* (see review Justice, 1993).

The second chapter aims to evaluate the role of the oxytocinergic system activation on the nigrostriatal dopaminergic signaling and its contribution to individual differences in impulsive choice behavior. For this, *in vivo* microdialysis was performed to study the effect of the perfusion of WAY267464 in SNpc on DA extracellular levels in DLS. In complementary studies, adult male rats were well-trained in the DDT. On separate test days, they received microinfusions of the OXT-R agonist (WAY267464) in the SNpc.

2 Hypothesis

General Hypothesis:

High impulsive choice level is associated with a decrease in the dopaminergic and oxytocinergic activity in the nigrostriatal pathway.

Specific Hypothesis:

2.1 Chapter 1:

Animals with high levels of impulsive choice have decreased levels of DA associated with increased DAT activity in DLS.

2.2 Chapter 2:

The activation of OXT-R in SNpc induces a decrease in impulsive choice behavior during DDT in high impulsive animals.

3 Objectives

General Objective:

Evaluate the contribution of dopaminergic and oxytocinergic systems in nigrostriatal pathway in the individual differences present in impulsive choice in rats.

Specific Objective:

3.1 Chapter 1:

1. To evaluate and classify the individual differences present in impulsive choice during delay discounting task.
2. To correlate the different impulsive choice levels in rats during DDT and the DA basal dynamic parameters (DA_{ext} , Ed , DA release) using No-Net-Flux microdialysis.

3.2 Chapter 2:

1. To evaluate the effect of bilateral administration of non-peptide specific OXT-R agonist (WAY267464) in SNpc on impulsive choice during DDT.
2. To determine the effect of administration of non-peptide specific OXT-R agonist (WAY267464) in SNpc on DA extracellular levels in DLS and SNpc.

4 Chapter 1: High impulsive choice is accompanied by an increase in dopamine release in rat dorsolateral striatum

Macarena Moreno¹, Víctor Azocar¹, Álvaro Vergés² and José Antonio

Fuentealba^{1*}.

¹ Department of Pharmacy and Interdisciplinary Center of Neuroscience, Faculty of Chemistry and Pharmacy, Pontificia Universidad Católica de Chile, Santiago, Chile.

² Escuela de Psicología, Pontificia Universidad Católica de Chile, Santiago, Chile.

*Corresponding Author:

José Antonio Fuentealba, Department of Pharmacy, Faculty of Chemistry and Pharmacy, Pontificia Universidad Católica de Chile, Santiago, Chile.

Email: jfuenta@uc.cl

Published at Behavioral Brain Research

DOI: <https://doi.org/10.1016/j.bbr.2021.113199>

Highlights

- Rats were classified as high impulsive and low impulsive according to the AUC.
- High impulsive rats showed an increased DA release in dorsolateral striatum.
- A negative correlation was observed between impulsive levels and DA release.

4.1 Abstract

Dopamine neurotransmission has been consistently associated with individual differences in impulsive choice. Clinical and preclinical evidence suggests that low striatal dopamine D₂ signaling predisposes to engage in impulsive behaviors. Although dopamine D₂ signaling controls dopamine (DA) extracellular levels, the relationship between striatal dopamine extracellular levels and impulsive choice remains poorly understood. Using quantitative microdialysis, we investigated whether extracellular DA levels in rat dorsolateral striatum (DLS) correlates with preference for an immediate small reward or for a delayed larger reward. Rats were tested in a delay-discounting task and classified as high impulsive (HI) or low impulsive (LI) according to the area under the discounting curve (AUC). No-net flux microdialysis experiments, assessing basal DA release, DA-uptake, and DA extracellular concentration (DA C_{ext}), were carried out in dorsolateral striatum (DLS) of urethane-anesthetized rats. Rats classified as HI showed a higher DA release compared with LI rats. Differences in DLS DA-uptake and DA C_{ext} were non-significant. Importantly, a significant negative correlation was observed between AUC and DA release, indicating that the lower the AUC, the higher the DLS DA release. This finding shows that DA release is augmented in the DLS of rats classified as HI, suggesting that a hyper-activated nigro-striatal pathway contributes to impulsive choice.

Keywords: Delay discounting task; impulsive choice; dorsolateral striatum; dopamine; No-net flux microdialysis.

Abbreviations: DDT, delay discounting task; DA C_{ext}, dopamine extracellular concentration; DLS, dorsolateral striatum; HI, high impulsive; LI, Low impulsive.

4.2 Introduction

Impulsivity is a construct characterized by unreflective decision making, premature behavior, and deficits in delaying gratification associated with an increased salience of the rewarding properties of the stimulus [1], [2]. High levels of impulsive behavior are key symptoms of attention-deficit hyperactivity disorder (ADHD), mania within bipolar disorder, problematic gambling, and substance use disorders [3]–[6]. Impulsivity has been proposed to comprise different dimensions, namely, impulsive action, and impulsive choice. Impulsive action is associated with a deficit in motor inhibition, whereas the impulsive choice, also known as impulsive decision-making, is associated with a deficit in delayed reward (for a review, see Dalley and Robbins [7] and Strickland & Johnson [8]).

Impulsive choice is a trait showing important individual differences, that is, characteristics that distinguish one organism from another and that are relatively stable over time and consistent across situations [9]. A common measure of impulsive choice is the delay discounting task (DDT), in which an excessive preference for a small immediate reward over a larger delayed reward, at the expense of more rewards earned per unit of time, is considered as an expression of potential maladaptive impulsive choice [10]–[12].

Clinical and preclinical evidence supports the notion that variability in dopamine (DA) neurotransmission function is a major contributor to individual differences observed in impulsive choice [13], [14]. For instance, genetic association studies provide evidence that the 10-variable number of tandem repeats (VNTR) polymorphism on the dopamine transporter gene (*DAT1*), related to hyperactive dopamine transporter (DAT) [15]–[18], is

believed to be a high risk allele associated to impulsive behaviors [19], [20]. Coincidentally, the DAT over-expression in rat nucleus accumbens (NAc) is accompanied by increased preference for a small reward in DDT [21]. These experiments suggested that an increase in DAT expression is associated with high impulsive choice.

Moreover, experiments using positron emission tomography (PET) showed that low dopamine D₂/D₃ receptor availability in the ventral tegmental area (VTA) and substantia nigra (SN) is associated with a highly impulsive phenotype [22]. Similarly, the knock-down of dopamine D₂ receptors in rat VTA induces preference for a smaller immediate reward indicating an increase in impulsive choice [23]. Decreased dopamine D₂/D₃ receptor availability in the dorsal striatum of methamphetamine users correlates with a high discounting rate of a delayed reward [24].

However, while it is known that an increased DAT activity and an attenuated nigro-striatal D₂ signaling could contribute to impulsive choice, the relationship between dorsal DA extracellular levels and impulsiveness needs further exploration. According to the inhibitory role of dopamine D₂ auto-receptor on DA release [25], an augmented amphetamine-stimulated DA release in human striatum is observed in impulsive subjects displaying low dopamine D₂/D₃ receptor availability [22]. Using functional magnetic resonance imaging (fMRI), Pine et al., [26] showed that an increased striatal activity induced by L-DOPA is observed in parallel with an enhanced preference for a smaller, immediate reward. These results suggest that an increased striatal DA release contributes to impulsive decision-making. However, the impulsivity associated with repeated exposure to psychostimulants has been associated with an attenuated striatal DA release

[27]. A blunted amphetamine-stimulated DA release in the dorsal striatum is associated with an impulsive choice of cocaine over money in cocaine users [28]. Insofar as psychostimulants users showed high levels of impulsivity, it has been suggested that a decreased striatal DA release observed in cocaine [29] and methamphetamine [30] abusers contribute to impulsive choice. In summary, low and high DA release could contribute to impulsive choice behavior, accordingly with the inverted-U shape pattern observed between DA signaling and cognitive function [31]–[33].

Nevertheless, preclinical approaches aimed to understand the mechanisms underlying the relationship between striatal DA extracellular levels and impulsive choice have only partially reproduced the imaging studies. In this context, evidence indicates that administration of the selective catecholamine uptake inhibitor 3,4-Methylenedioxypyrovalerone (MDPV) increase impulsive choice, accompanied by an increase in striatal DA turnover [34]. On the other hand, voltammetry experiments showed that rats classified as high impulsive in the DDT display a blunted ventral striatal DA release independent of the history of self-administered cocaine [35]. Similarly, a decrease in the electrically evoked tritiated DA release is observed in slices of NAc and mPFC of impulsive rats [36]. However, a recent amperometric study fails to show differences in ventral striatal DA release between rats classified as high and low impulsive in the DDT [37]. Moreover, studies assessing the role of the nigro-striatal dopaminergic pathway on impulsive choice fail to show conclusive results due to important methodological issues. Using intracranial self-stimulation as positive reinforcement in the delay discounting task, Tedford et al., [38] showed that lesion of the dopaminergic terminal in the dorsolateral striatum (DLS) by 6-hydroxydopamine (6-OHDA) is accompanied by an increase in

impulsive choice. However, using sucrose as a positive reinforcer, Magnard et al., [39] showed that (6-OHDA)-induced lesion of substantia nigra pars compacta (SNpc) is not accompanied by changes in impulsive choice. Because the dorsal striatal DA content was not measured in these studies, it remains unclear whether changes in DA concentration are associated with impulsive choice. The main goal of this research is to determine the relationship between delay discounting behavior and DA dynamics in rat DLS. Microdialysis no-net flux experiments were carried out to assess the contribution of DA release and DA-uptake on DLS DA extracellular concentration (DA C_{ext}) in rats classified as high or low impulsive in the DDT. Our results showed that an increased DLS DA release is positively correlated with impulsive levels, showing that high DA release is associated with higher levels of delay discounting

4.3 Experimental Procedures

4.3.1 Animals

Fifteen male Sprague Dawley rats, approximately postnatal day (PND) 25, were grown from the Animal Care Faculty of Biological Science, Pontificia Universidad Católica de Chile (Charles River; Wilmington, MA, USA; research resource identifiers: RGD_728193) under the strict supervision of a veterinarian. During behavioral test, rats were maintained in the Animal Care of the Department of Pharmacy, Pontificia Universidad Católica de Chile, using protocols and instructions approved by the veterinarian. Rats were housed in pairs in ventilated cages by an Air Handling Unit (Tecniplast, West Chester, PA) under controlled conditions including temperature ($22\pm1^{\circ}\text{C}$), and humidity ($50\pm10\%$). The animals were additionally maintained under a 12-hour light-dark cycle (lights on 07:00-19:00 hrs). Experiments were conducted during the light period. Rats were handled for seven days before starting the experiments. Six days after arrival (PND 31), food access was restricted, and rats were weighed daily to maintain body weights to 90% of the free-feed body weight. We allowed the increase in the restricted body weight per week to let the growth associated with age. The rats started with an approximate weight of 75 gr and ended up weighing approximately 420 gr. The target weights were adjusted according to normal growth curves obtained from the animal supplier. Water was available ad libitum. All procedures were in strict accordance with the guidelines published in the “NIH Guide for the Care and Use of Laboratory Animals” (8th ed) and principles presented in the “Guidelines for the Use of Animals in Neuroscience Research” by the Society for Neuroscience. Furthermore, all procedures were approved

by the ethics committee of the Pontificia Universidad Católica de Chile. Moreover, advice from a veterinarian was also obtained during the process.

4.3.2 Apparatus

Sessions were conducted in ventilated, sound-attenuating enclosures containing an operant conditioning chamber with an interior space of measures $31 \times 24 \times 21$ cm (Med Associates, Inc.; St Albans, VT). The front door and the rear panel were made from clear polycarbonate, and both ends were made of aluminum panels. The right panel was equipped with two response levers, horizontally aligned 11.5 cm apart. Above each lever, there was a translucent circle trans-illuminated with white light, 2.5 cm in diameter. A pellet receptacle was centrally located between the two levers. 45 mg of food chocolate in form (F0299 chocolate dustless precision pellets, Bio-serv; Flemington, NJ) was delivered to this receptacle from an external food dispenser. The panel on the opposite side of the chamber was equipped with a house-light centrally located near the top of the chamber. Data was collected using MED-PC IV software and a PC-compatible interface (Med Associates, Inc; St Albans, VT).

4.3.3 Procedures

Rats were tested in the behavioral procedures as described below. All procedures were adapted from Eubig P et al., [40] and Tanno et al., [41].

4.3.3.1 Initial lever press training

Approximately after PND 35, a shaping program was used to train rats to begin pressing the response levers. Both levers were extended at the onset of the session. As described in Eubig P et al., [40] the delivery was programmed according to a 3-min fixed-time schedule, in which a food pellet was dispensed every 3 min. Also, one pellet was delivered with every lever press. Sessions finished after either 60 min elapsed, or 100 reinforcements were delivered. The criterion for advancement to the next phase of training was 100 lever presses, performed for three consecutive sessions.

4.3.3.2 Fixed ratio training

The second training phase began between PND 39-40. During this phase, only one lever was extended, and the corresponding cue light was illuminated. After five lever presses, the extended lever was retracted, and the opposite lever was extended. Every response resulted in the delivery of a food pellet (i.e. a fixed ratio 1 contingency). After 100 reinforcements across three consecutive sessions, rats continued to the next training phase.

4.3.3.3 Delay discounting training and testing

The DDT training phase began approximately after PND 45. The phase consisted of 40 trials for each session. Two levers with different reinforcers were presented and were always accompanied by an illuminated cue light: the lever press of the larger reinforcer (LR) resulted in 3 pellets and the small reinforcer (SR) lever resulted in 1 pellet. There

was no delay in the delivery of either the small or large reinforcers in this phase. The session finished after 40 lever presses between large or small reinforcers. The criterion for advancement to the next phase of training required an 80 percent of preference for the larger reinforcer.

The DDT testing adapted from Tanno et al., [41], began in darkness with the levers retracted (blackout). After 10 minutes of blackout, the house-light turned on, and the session started. One session consisted of five blocks comprising 12 trials each. The duration of the trials was of 60 seconds. The first two trials of each block were forced-choice trials in which the LR lever and the SR lever were alternately introduced. Ten free-choice trials followed the force trials in each block. At the beginning of the free-choice trial, the lights over the levers were turned on, and the levers appeared. If a lever was pressed, both levers were retracted. If the rat pressed the SR lever, one pellet was immediately delivered, and both levers were retracted. If the LR lever was chosen, the stimulus light blinked during the delay and, following the delay, three pellets were delivered. Delays to the LR were increased from 0s in block 1 to 4s in block 2, 8s in block 3, 16s in block 4, and 32s in block 5. Failure to choose a response within 20 s resulted in the levers retracting and the trial was considered as an omission (see figure 1). The rats were trained five times per week for approximately ten weeks (50 sessions) in this task until stable performance was achieved. (See Data Analysis section for more details).

4.3.4 Surgery and no-net-Flux Microdialysis

Surgery and no-net-flux microdialysis were performed at approximately PND 115-120, two hours after the last DDT session. Rats were anesthetized with urethane (1.5 g/kg i.p) which remained constant throughout the experiment due to its extended half-life [42] and were placed in a stereotaxic apparatus. The skull of each rat was exposed, and a hole was drilled targeting the DLS. A concentric microdialysis probe (CMA-11 Microdialysis, Holliston, MA) was implanted in the DLS using the following coordinates +1.2 AP, +3.6 ML relative to bregma and -4.8 DV from the dura [43]. Body temperature was maintained by a thermostatically controlled electric heating pad. The microdialysis probe was perfused for 40 minutes to allow equilibrium with Krebs–Ringer phosphate (KRP) buffer at a rate of 2 μ L/min using a Harvard infusion pump (Harvard Apparatus, Holliston, MA). The composition of the KRP was 120 mM NaCl, 2.4 mM KCl, 1.2 mM CaCl₂, 0.9 mM NaH₂PO₄, 1.4 mM Na₂HPO₄, and 0.2 mM of ascorbic acid (pH 7.4). The probe was randomly perfused with four different concentrations of DA: 0, 5, 20, and 40 nM in a KRP to determine extracellular DA concentration (DA C_{ext}), DA release, and extraction fraction (Ed), which is an indirect measure of DA-uptake [44], [45]. Three consecutive samples were collected every 5 minutes for each concentration of DA after a stabilization period of 30 minutes. Perfusion samples were collected in 2 μ L of perchloric acid (0.2 N) and maintained at 4°C, until analysis (see, section 2.4.2).

4.3.5 Histology

After microdialysis experiments, rats were decapitated under deep anesthesia (urethane 1.5 g/kg i.p.), and brains were extracted and cleaned with a saline solution (NaCl 0.9%). Brains were stored in 4% paraformaldehyde (PFA). At least two days before slicing, brains were cryoprotected using a 30% sucrose solution. For probe implantation, brains were frozen and coronally sliced in sections of 50 μ m. Slices were stained with cresyl violet, and the probe placement was observed in a light microscope using the atlas of Paxinos & Watson for rats [41]. Only data coming from correct probe placements were considered for further analysis (see figure 2).

4.3.6 Analysis of dialysate dopamine

Ten ml of the dialysate was injected using a Rheodyne injector valve into an HPLC system (BASi America, West Lafayette, IN). The mobile phase contained 100 mM NaH_2PO_4 , 1 mM EDTA, 1 mM octane-1-sulfonic acid sodium salt, and 3% acetonitrile (pH 3.0), and was pumped at a flow rate of 700 μ L/min. The potential of the amperometric detector was set at 650 mV. Under these experimental conditions, the retention time for DA was 6 min.

4.3.7 Data Analysis

4.3.7.1 Delay discounting analysis

4.3.7.1.1 The logarithmic area under the curve (AUC_{log})

Delay-discounting curves were obtained by plotting the proportion of LR choices (the number of LR choices divided by the total number of successful trials) of each block versus each delay (sec) (see figure 3A). The area under the delay-discounting curve (AUC) was used as a measure of impulsive choice [46], [47]. The AUC was calculated using a log-transformation of delays (AUC_{log}) to avoid the disproportionate contribution at long delays in the delay-discounting curves [48]. Specifically, each delay was transformed to the logarithm of the delay divided by the bigger logged delay. To avoid the problem of calculating the logarithm of zero, a small amount (value=1) was added in each delay before the transformation. We use GraphPad Prism to calculate AUC_{log} (version 5, GraphPad Software, Inc.; La Jolla, CA). Higher AUC_{log} values indicate lower discounting by delay (i.e., preference for larger rewards, indicating lower impulsivity), while lower AUC_{log} values correspond to higher discounting by delay (i.e., preference for small and immediate rewards, indicating higher impulsivity) [46].

4.3.7.1.2 Behavioral Stability and Classification

To assess behavioral stability, we compared the AUC_{log} of five consecutive sessions (week 1) with the five following sessions (week 2). An animal was considered trained when the difference between weeks was not statistically significant. For each animal, a stable AUC_{log} value was calculated using the average of the last five sessions (Figure 3B). Rats were classified as low impulsive (LI) and high impulsive (HI) based on the median split of AUC_{log} . The minimum sample size was calculated based on a priori power analysis

[49]. The input parameters for behavioral experiments were the following: two-tails, $\alpha = 0.05$, power = 0.95, and ratio $N1/N2 = 1$. The effect size was based on preliminary DDT results, suggesting a minimum sample size of $n = 7/\text{group}$ for the experiment considering a Cohen's d effect size of 2, using G*Power statistical analysis software Version 3.1.9.4 [50].

4.3.7.1.3 Omission and latencies

Latency was defined as the time since the levers were extended until the rat pressed one of them during free-choice trials in each block. An omission was considered as a failure to choose a response within 20 s after lever appearance. For each animal, we recorded the latency and the number of omissions in the 5 daily blocks in free-choice trials. Final latency and omission values were calculated averaging total latency and omissions of the last five sessions.

4.3.8 No-Net-Flux analysis

The amount of DA gained or lost from the probe during the no-net flux microdialysis ($C_{in} - C_{out}$) was calculated for each animal at each perfusion concentration (C_{in} : 0, 5, 10, 20, 40 nM DA). The net change in DA was plotted against C_{in} ($C_{in} - C_{out}$ vs. C_{in}) and subjected to linear regression (figure 4A). The DA C_{ext} concentration and E_d were assessed using linear regression. The point where no DA was gained or lost ($C_{in} - C_{out} = 0$) represents an estimate of DA C_{ext} . The E_d is the slope of the linear regression and has been shown to provide an indirect measure of the DA-uptake. Basal dialysate DA was calculated for each animal as

the average of the three basal samples ($C_{in}=0$). An unpaired t-test was used to assess the effect of the impulsivity level on basal dialysate DA levels, DA C_{ext} , and Ed [45], [51].

4.3.9 Statistical Analyses

All statistical analyses were carried out using Prism 5.0 GraphPad Software. Each group of data was analyzed with the nonlinear regression ROUT method ($Q=1\%$) [52], the outlier detected was removed from the statistical analysis and graphs (total outliers: 1). The resulting data were analyzed using an unpaired t-test when it was appropriate (Figure 4; all data are reported as mean \pm SEM). Correlation analyses were performed using the Spearman correlation coefficient for non-parametric data (Figure 5).

4.4 Results

4.4.1 Individual differences in basal level of impulsive choice

Figure 3A shows the delay-discounting curve for all rats. According to an AUC_{log} median split, rats were classified as LI ($n=7$) and HI ($n=7$) (figure 3B LI: 0.81 ± 0.02 ; vs HI: 0.54 ± 0.05 ; $p = 0.0002$, according to unpaired t-test). Non-significant differences between HI and LI rats were observed in latency (LI: 1.74 ± 0.22 sec vs HI: 1.81 ± 0.01 sec, $p = 0.785$ according to unpaired t-test) and omissions (LI: 1.09 ± 0.64 vs HI: 1.81 ± 0.10 , $p = 0.322$ according to unpaired t-test).

4.4.2 Animals classified as high impulsive in DDT showed higher DA C_{ext} in DLS

To compare the DLS DA dynamics between HI and LI rats, no-net-flux microdialysis experiments were carried out after DDT. Rats classified as HI showed a significantly higher DA release in DLS compared with LI rats (figure 4B LI: 0.43 ± 0.11 nM vs HI: 0.89 ± 0.14 nM, $p = 0.022$ according to unpaired t-test). The extraction fraction (Ed) showed no significant differences between HI and LI rats (figure 4C LI: 0.52 ± 0.05 vs HI: 0.46 ± 0.04 , $p = 0.331$, according to unpaired t-test). Also, no significant differences were observed in DA extracellular concentration (C_{ext}) in DLS between rats classified as HI compared with LI rats (figure 4D LI: 1.13 ± 0.23 nM vs HI: 1.61 ± 0.28 nM; $p = 0.215$ according to unpaired t-test). Additionally, using a third classification criterion the differences in the DLS DA release in HI rats compared with LI rats are maintained (LI: 0.34 ± 0.11 , $n=5$ vs HI: 0.88 ± 0.20 , $n=5$; $p = 0.042$).

4.4.3 Correlation between impulsive choice levels and DA dynamic in the DLS

In order to evaluate if there is an association between basal levels of impulsive choice and DA dynamics in the DLS, a correlation between AUC and the different neurochemical parameters associated with DA dynamics was carried out. As illustrated in figure 5A a significant negative correlation was observed between AUC and DLS DA release ($r = -0.69$, $p = 0.008$; $n=14$). The AUC and Ed in DLS showed non-significant correlation (figure 5B $r = 0.26$, $p = 0.374$; $n=14$). A non-significant correlation was also observed between AUC and DLS DA C_{ext} (figure 5C $r = -0.51$; $p = 0.064$; $n=14$).

4.5 Discussion

It has been consistently shown that variability in DA neurotransmission is a mechanism underpinning individual differences observed in impulsive choice, also named delay-related decision making [21], [23], [53]. Single unit recording experiments showed that DA neurons in the SNpc modify their activity depending on delay and reward magnitude [54], implying that DA neurotransmission in the dorsal striatum could contribute to information processing during delay-related decision making [55]. In the current study, we examined the relationship between impulsive choice and dopaminergic dynamics in the rat DLS. Rats were classified as high and low impulsive using a median split of AUC in discounting curves. Given that, this classification criterion could be considered problematic because some rats may have AUCs near the average score, we also analyzed our data using selected extremes values after a third classification. In this analysis, the differences in DLS DA extracellular levels are maintained between rats classified as HI (lower third) and LI (upper third). Thus, our findings seem to be robust to different classification criteria, in the absence of a consensual procedure of classification in the delay discounting literature. A higher DLS DA release was observed in HI rats compared with LI rats. Also, a significant negative correlation was observed between AUC and DA release, indicating that lower AUC is associated with higher DLS DA release. Notwithstanding of limitations of our study, these results suggest that an increased DLS DA signaling contribute to delay-related decision-making.

In line with our results, clinical evidence shows that basal striatal DA concentration appears to be relevant to predict both vulnerabilities to engage in impulsive behaviors as

well as to the response to pharmacological treatment. The impulsiveness observed in ADHD has been correlated with both an increase [26], [56] and a decrease in striatal DA signaling [57]. Consequently, the therapeutic effectiveness of psychostimulants for ADHD treatment has been associated with a decrease [56] and an increase in striatal DA release [58]. This apparent discrepancy in results related to dopaminergic signaling and impulsivity could be attributed to individual variability in striatal basal DA concentration [11], [31]. It has been proposed that the relation between basal DA concentration and neural reward processing during decision-making shows an inverted-U shape pattern: excessively low or high DA concentration could predispose to impulsive behavior [31], [32]. Then, an increase in striatal DA release induced by psychostimulant drugs would benefit subjects with low DA concentration but could worsen impulsivity in subjects with elevated DA concentration [27], [32]. Then, given that our study is limited by the relatively small sample size, the results could be showing the descending limb of the curve, i.e., the impulsive choice associated with an increase in DLS dopamine neurotransmission. The ascendent limb of the curve, necessary to observe the inverted-U curve, that could be appear after a pharmacological blockade of DA neurotransmission was not tested in this study [33].

The increased DA release observed in rats classified as HI could be explained by presynaptic and postsynaptic mechanisms. It has been consistently shown that an attenuated D₂ receptor activity contributes to impulsive choice. Critically, it has proposed that individual differences in D₂ receptor mRNA expression predicts different levels of impulsive choice [59]. Low D₂ receptor mRNA expression in the NAc is observed in male rats showing higher levels of delay discounting [59]. Interestingly, a low D₂ receptor

mRNA expression in the mPFC has been observed in male and female rats classified as high impulsive in the DDT [59], [60]. In the same line, a polymorphism in the D₂ receptor gene (C957T) showing a decreased striatal binding of D₂ receptor has been associated with a higher preference for the immediate reinforcement in the DDT during stress [61]. Furthermore, clinical and preclinical research has shown that prolonged exposure to methamphetamine, amphetamine, or cocaine reduced D₂ receptor expression in the dorsal striatum, leading to high levels of impulsive choice [24], [62]–[64]. Similarly, the specific down expression of D₂ receptors in rat dopaminergic mesencephalic neurons is accompanied by an increase in impulsive choice [23]. Then, it is possible to suggest that the higher DA release observed in HI rats is due to a loss of the inhibitory tonic control of D₂ receptors on DA extracellular concentration [25]. Together with the D₂ receptor, DAT activity also controls striatal DA release, keeping DA C_{ext} levels stabilized [65]–[67]. In addition to its important role in DA re-uptake [68], it was recently shown that DAT also promotes DA release in the dorsal striatum, contributing to local short-term plasticity mechanisms of DA release [69]. Importantly, it has been shown that an increase in striatal DAT activity underlies impulsivity in ADHD [70], and consequently, DAT blockade has been proposed as a therapeutic mechanism for the treatment of impulsivity in clinical [58], [71] and preclinical [72]–[74] studies. In line with these findings, striatal over-expression of DAT in rat ventral striatum increases impulsive choice [21]. However, attenuation of DAT expression and repeated exposure to DAT antagonist MDVP also induced an increase in impulsive choice [21], [34]. This inconsistency may be due to DAT activity complex effects on DA release [69], so the consequences of modifying the DAT expression on striatal DA C_{ext} are difficult to predict. In this context, our results showed

no significant differences in striatal DA Ed between HI and LI rats, suggesting that changes in DAT activity in DLS do not contribute to individual differences in delay-related decision making. Because the Ed is an indirect measure of DA-uptake [44], [75], this result therefore should be interpreted with caution, and other approaches [76] should be considered to better understand the role of dorsal striatum DAT to impulsive choice.

Regarding postsynaptic mechanisms, the increased DA release observed in the DLS of HI rats could be associated with changes in neuronal activity in the nigro-striatal pathway. Supporting this idea, the persistent impulsivity observed after repeated administration of MDPV is accompanied by an increase in striatal DOPAC/DA ratio, suggesting an increased nigro-striatal pathway neuronal activity [34], [77]. The neuronal activity of mesencephalic DA neurons controls striatal DA C_{ext} [78], [79]. An increase in DLS DA release is observed concomitantly with the increase in the firing rate and burst activity of DA neurons in SNpc [79]. Then, it is possible to suggest that higher neuronal activity in the dopaminergic neurons of SNpc supports the increased DA release in DLS observed in rats classified as HI, compared with LI rats. In fact, we recently showed that the increase in DLS DA release observed after adolescent exposure to cannabinoids is accompanied by an increase in population activity in SNpc [80]. It has been established that GABA and glutamate inputs control the excitability of mesencephalic dopaminergic neurons. While the number of spontaneously active DA neurons is regulated by GABA afferences, the burst pattern of DA neurons depends on glutamate afferences [78]. Our results suggest that an activated nigro-striatal pathway contributes to high delay discounting probably associated with an increase in population activity or bursting activity of DA neurons in

the SNpc [67]. Whether variability in GABA and glutamate levels in SNpc contribute to individual differences in delay-discounting behavior is unknown.

In summary, our results showed that DA neurotransmission in DLS contributes to individual differences in impulsive choice. The increased DLS DA release observed in HI rats could maintain activated postsynaptic DA receptor in dorsal striatum responsible for increasing delay aversion [81]. Further research is necessary to elucidate the role of nigral DA neurons during impulsive choice and the mechanisms contributing to the activated nigro-striatal pathway associated with a high discounting rate of a delayed reward.

4.6 Acknowledgments

This work was supported by FONDECYT, Fondo Nacional de Desarrollo Científico y Tecnológico, Chile. ID (1141088; J.A.F.); ANID-PCHA (21150508; M.M); ANID-PCHA (21171 609; V.A).

The authors have no conflict of interest.

4.7 References

- [1] J. D. Jentsch and J. R. Taylor, “Impulsivity resulting from frontostriatal dysfunction in drug abuse: implications for the control of behavior by reward-related stimuli,” *Psychopharmacology (Berl)*., vol. 146, no. 4, pp. 373–390, Oct. 1999, doi: 10.1007/PL00005483.
- [2] G. S. Berlin and E. Hollander, “Compulsivity, impulsivity, and the DSM-5 process,” *CNS Spectrums*, vol. 19, no. 1. CNS Spectr, pp. 62–68, Feb-2014, doi: 10.1017/S1092852913000722.
- [3] K. R. Hamilton *et al.*, “Choice impulsivity: Definitions, measurement issues, and clinical implications,” *Personal. Disord. Theory, Res. Treat.*, vol. 6, no. 2, pp. 182–198, Apr. 2015, doi: 10.1037/per0000099.
- [4] B. Setlow, I. A. Mendez, M. R. Mitchell, and N. W. Simon, “Effects of chronic administration of drugs of abuse on impulsive choice (delay discounting) in animal models,” *Behavioural Pharmacology*, vol. 20, no. 5–6. Behav Pharmacol, pp. 380–389, Sep-2009, doi: 10.1097/FBP.0b013e3283305eb4.
- [5] J. M. Berg, R. D. Latzman, N. G. Bliwise, and S. O. Lilienfeld, “Parsing the heterogeneity of impulsivity: A meta-analytic review of the behavioral implications of the UPPS for psychopathology,” *Psychol. Assess.*, vol. 27, no. 4, pp. 1129–1146, Dec. 2015, doi: 10.1037/pas0000111.
- [6] J. Vassileva and P. J. Conrod, “Impulsivities and addictions: a multidimensional integrative framework informing assessment and interventions for substance use disorders,” *Philosophical transactions of the Royal Society of*

London. Series B, Biological sciences, vol. 374, no. 1766. NLM (Medline), p. 20180137, Feb-2019, doi: 10.1098/rstb.2018.0137.

[7] J. W. Dalley and T. W. Robbins, “Fractionating impulsivity: Neuropsychiatric implications,” *Nature Reviews Neuroscience*, vol. 18, no. 3. Nature Publishing Group, pp. 158–171, Mar-2017, doi: 10.1038/nrn.2017.8.

[8] J. C. Strickland and M. W. Johnson, “Rejecting Impulsivity as a Psychological Construct: A Theoretical, Empirical, and Sociocultural Argument,” *Psychol. Rev.*, 2020, doi: 10.1037/rev0000263.

[9] J. Peters and C. Büchel, “The neural mechanisms of inter-temporal decision-making: Understanding variability,” *Trends Cogn. Sci.*, vol. 15, no. 5, pp. 227–239, 2011, doi: 10.1016/j.tics.2011.03.002.

[10] A. T. Marshall, A. P. Smith, and K. Kirkpatrick, “Mechanisms of impulsive choice: I. Individual differences in interval timing and reward processing,” *J. Exp. Anal. Behav.*, vol. 102, no. 1, pp. 86–101, 2014, doi: 10.1002/jeab.88.

[11] J. D. Jentsch, J. R. Ashenurst, M. C. Cervantes, S. M. Groman, A. S. James, and Z. T. Pennington, “Dissecting impulsivity and its relationships to drug addictions,” *Ann. N. Y. Acad. Sci.*, vol. 1327, no. 1, pp. 1–26, 2014, doi: 10.1111/nyas.12388.

[12] J. E. Mazur and D. Coe, “Tests of transitivity in choice between fixed and variable reinforcer delays,” *J. Exp. Anal. Behav.*, vol. 47, no. 3, pp. 287–297, May 1987, doi: 10.1901/jeab.1987.47-287.

[13] M. M. van Gaalen, R. van Koten, A. N. M. Schoffeleer, and L. J. M. J. Vanderschuren, “Critical Involvement of Dopaminergic Neurotransmission in

Impulsive Decision Making,” *Biol. Psychiatry*, vol. 60, no. 1, pp. 66–73, 2006, doi: 10.1016/j.biopsych.2005.06.005.

[14] J. Joutsa, V. Voon, J. Johansson, S. Niemelä, J. Bergman, and V. Kaasinen, “Dopaminergic function and intertemporal choice,” *Transl. Psychiatry*, vol. 5, no. 1, p. e491, Jan. 2015, doi: 10.1038/tp.2014.133.

[15] J. M. Swanson *et al.*, “Dopamine genes and ADHD,” in *Neuroscience and Biobehavioral Reviews*, 2000, vol. 24, no. 1, pp. 21–25, doi: 10.1016/S0149-7634(99)00062-7.

[16] A. Heinz *et al.*, “Genotype influences in vivo dopamine transporter availability in human striatum,” *Neuropsychopharmacology*, vol. 22, no. 2, pp. 133–139, Feb. 2000, doi: 10.1016/S0893-133X(99)00099-8.

[17] A. Kirley *et al.*, “Dopaminergic system genes in ADHD: Toward a biological hypothesis,” *Neuropsychopharmacology*, vol. 27, no. 4, pp. 607–619, Oct. 2002, doi: 10.1016/S0893-133X(02)00315-9.

[18] S. H. VanNess, M. J. Owens, and C. D. Kilts, “The variable number of tandem repeats element in DAT1 regulates in vitro dopamine transporter density,” *BMC Genet.*, vol. 6, no. 1, p. 55, Nov. 2005, doi: 10.1186/1471-2156-6-55.

[19] I. D. Waldman *et al.*, “Association and linkage of the transporter gene and attention-deficit hyperactivity disorder in children: Heterogeneity owing to diagnostic subtype and severity,” *Am. J. Hum. Genet.*, vol. 63, no. 6, pp. 1767–1776, Dec. 1998, doi: 10.1086/302132.

[20] I. R. Gizer and I. D. Waldman, “Double Dissociation Between Lab Measures of Inattention and Impulsivity and the Dopamine Transporter Gene

(DAT1) and Dopamine D4 Receptor Gene (DRD4),” *J. Abnorm. Psychol.*, vol. 121, no. 4, pp. 1011–1023, Nov. 2012, doi: 10.1037/a0028225.

[21] W. Adriani, F. Boyer, L. Gioiosa, S. Macrì, J. L. Dreyer, and G. Laviola, “Increased impulsive behavior and risk proneness following lentivirus-mediated dopamine transporter over-expression in rats’ nucleus accumbens,” *Neuroscience*, 2009, doi: 10.1016/j.neuroscience.2008.11.042.

[22] J. W. Buckholtz *et al.*, “Dopaminergic network differences in human impulsivity,” *Science*, vol. 329, no. 5991. NIH Public Access, p. 532, Jul-2010, doi: 10.1126/science.1185778.

[23] K. A. Bernosky-Smith *et al.*, “Ventral tegmental area D2 receptor knockdown enhances choice impulsivity in a delay-discounting task in rats,” *Behav. Brain Res.*, vol. 341, no. September 2017, pp. 129–134, 2018, doi: 10.1016/j.bbr.2017.12.029.

[24] M. E. Ballard, A. C. Dean, M. A. Mandelkern, and E. D. London, “Striatal dopamine d2/d3 receptor availability is associated with executive function in healthy controls but not methamphetamine users,” *PLoS One*, vol. 10, no. 12, Dec. 2015, doi: 10.1371/journal.pone.0143510.

[25] A. Mayer, N. Limberger, and K. Starke, “Transmitter release patterns of noradrenergic, dopaminergic and cholinergic axons in rabbit brain slices during short pulse trains, and the operation of presynaptic autoreceptors,” *Naunyn-Schmiedeberg's Arch. Pharmacol.*, vol. 338, no. 6, pp. 632–643, 1988, doi: 10.1007/BF00165627.

[26] A. Pine, T. Shiner, B. Seymour, and R. J. Dolan, “Dopamine, time, and

impulsivity in humans,” *J. Neurosci.*, vol. 30, no. 26, pp. 8888–8896, Jun. 2010, doi: 10.1523/JNEUROSCI.6028-09.2010.

[27] P. Trifilieff and D. Martinez, “Imaging addiction: D2 receptors and dopamine signaling in the striatum as biomarkers for impulsivity,” *Neuropharmacology*, vol. 76, pp. 498–509, Jan. 2014, doi: 10.1016/J.NEUROPHARM.2013.06.031.

[28] D. Martinez *et al.*, “Amphetamine-induced dopamine release: Markedly blunted in cocaine dependence and predictive of the choice to self-administer cocaine,” *Am. J. Psychiatry*, vol. 164, no. 4, pp. 622–629, 2007, doi: 10.1176/ajp.2007.164.4.622.

[29] N. D. Volkow *et al.*, “Decreased striatal dopaminergic responsiveness in detoxified cocaine-dependent subjects,” *Nature*, vol. 386, no. 6627, pp. 830–833, Apr. 1997, doi: 10.1038/386830a0.

[30] G. J. Wang *et al.*, “Decreased dopamine activity predicts relapse in methamphetamine abusers,” *Mol. Psychiatry*, vol. 17, no. 9, pp. 918–925, Sep. 2012, doi: 10.1038/mp.2011.86.

[31] J. Yacubian *et al.*, “Gene-gene interaction associated with neural reward sensitivity,” *Proc. Natl. Acad. Sci. U. S. A.*, vol. 104, no. 19, pp. 8125–8130, May 2007, doi: 10.1073/pnas.0702029104.

[32] M. Leyton, “Conditioned and sensitized responses to stimulant drugs in humans,” *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, vol. 31, no. 8, Prog Neuropsychopharmacol Biol Psychiatry, pp. 1601–1613, Nov-2007, doi: 10.1016/j.pnpbp.2007.08.027.

- [33] J. Petzold *et al.*, “Presynaptic dopamine function measured with [18F]fluorodopa and L-DOPA effects on impulsive choice,” *Sci. Rep.*, vol. 9, no. 1, 2019, doi: 10.1038/s41598-019-54329-1.
- [34] W. S. Hyatt *et al.*, “Repeated administration of synthetic cathinone 3,4-methylenedioxypyrovalerone persistently increases impulsive choice in rats,” *Behav. Pharmacol.*, vol. 30, no. 7, pp. 555–565, Oct. 2019, doi: 10.1097/FBP.0000000000000492.
- [35] T. M. Moschak and R. M. Carelli, “Impulsive rats exhibit blunted dopamine release dynamics during a delay discounting task independent of cocaine history,” *eNeuro*, vol. 4, no. 2, 2017, doi: 10.1523/ENEURO.0119-17.2017.
- [36] L. Diergaarde *et al.*, “Impulsive Choice and Impulsive Action Predict Vulnerability to Distinct Stages of Nicotine Seeking in Rats,” *Biol. Psychiatry*, vol. 63, no. 3, pp. 301–308, 2008, doi: 10.1016/j.biopsych.2007.07.011.
- [37] T. G. Freels, D. B. K. Gabriel, D. B. Lester, and N. W. Simon, “Risky decision-making predicts dopamine release dynamics in nucleus accumbens shell,” *Neuropsychopharmacology*, vol. 45, no. 2, pp. 266–275, 2020, doi: 10.1038/s41386-019-0527-0.
- [38] S. E. Tedford, A. L. Persons, and T. C. Napier, “Dopaminergic lesions of the dorsolateral striatum in rats increase delay discounting in an impulsive choice task,” *PLoS One*, vol. 10, no. 4, pp. 1–17, 2015, doi: 10.1371/journal.pone.0122063.
- [39] R. Magnard *et al.*, “Nigrostriatal Dopaminergic Denervation Does Not Promote Impulsive Choice in the Rat: Implication for Impulse Control Disorders

in Parkinson's Disease," *Front. Behav. Neurosci.*, vol. 12, no. December, pp. 1–10, Dec. 2018, doi: 10.3389/fnbeh.2018.00312.

[40] P. A. Eubig, T. E. Noe, S. B. Floresco, J. J. Sable, and S. L. Schantz, "Sex differences in response to amphetamine in adult Long-Evans rats performing a delay-discounting task," *Pharmacol. Biochem. Behav.*, vol. 118, pp. 1–9, 2014, doi: 10.1016/j.pbb.2013.12.021.

[41] T. Tanno, D. R. Maguire, C. Henson, and C. P. France, "Effects of amphetamine and methylphenidate on delay discounting in rats: Interactions with order of delay presentation," *Psychopharmacology (Berl.)*, vol. 231, no. 1, pp. 85–95, Jan. 2014, doi: 10.1007/s00213-013-3209-3.

[42] M. Gumbleton and L. Z. Benet, "Drug metabolism and laboratory anesthetic protocols in the rat: Examination of antipyrine pharmacokinetics," *Pharm. Res.*, vol. 8, no. 4, pp. 544–546, 1991, doi: 10.1023/a:1015827917684.

[43] G. Paxinos and C. Watson, *The rat brain in stereotaxic coordinates*. Elsevier, 2009.

[44] A. D. Smith and J. B. Justice, "The effect of inhibition of synthesis, release, metabolism and uptake on the microdialysis extraction fraction of dopamine," *J. Neurosci. Methods*, vol. 54, no. 1, pp. 75–82, 1994, doi: 10.1016/0165-0270(94)90161-9.

[45] V. H. Azocar, G. Sepúlveda, C. Ruiz, C. Aguilera, M. E. Andrés, and J. A. Fuentealba, "The blocking of kappa-opioid receptor reverses the changes in dorsolateral striatum dopamine dynamics during the amphetamine sensitization," *J. Neurochem.*, vol. 148, no. 3, pp. 348–358, Feb. 2019, doi: 10.1111/jnc.14612.

- [46] J. Myerson, L. Green, and M. Warusawitharana, “Area under the curve as a measure of discounting,” *J. Exp. Anal. Behav.*, vol. 76, no. 2, pp. 235–43, Sep. 2001, doi: 10.1901/jeab.2001.76-235.
- [47] J. M. Slezak and K. G. Anderson, “Effects of variable training, signaled and unsignaled delays, and d-amphetamine on delay-discounting functions,” *Behav. Pharmacol.*, vol. 20, no. 5–6, pp. 424–436, 2009, doi: 10.1097/FBP.0b013e3283305ef9.
- [48] A. M. Borges, J. Kuang, H. Milhorn, and R. Yi, “An alternative approach to calculating Area-Under-the-Curve (AUC) in delay discounting research,” *J. Exp. Anal. Behav.*, vol. 106, no. 2, pp. 145–155, 2016, doi: 10.1002/jeab.219.
- [49] F. Faul, E. Erdfelder, A. G. Lang, and A. Buchner, “G*Power 3: A flexible statistical power analysis program for the social, behavioral, and biomedical sciences,” in *Behavior Research Methods*, 2007, vol. 39, no. 2, pp. 175–191, doi: 10.3758/BF03193146.
- [50] S. Mayr, E. Erdfelder, A. Buchner, and F. Faul, “A short tutorial of GPower,” *Tutor. Quant. Methods Psychol.*, vol. 3, no. 2, pp. 51–59, 2007, doi: 10.20982/tqmp.03.2.p051.
- [51] V. I. Chefer, A. Zapata, T. S. Shippenberg, and P. M. Bungay, “Quantitative no-net-flux microdialysis permits detection of increases and decreases in dopamine uptake in mouse nucleus accumbens,” *J. Neurosci. Methods*, vol. 155, no. 2, pp. 187–193, 2006, doi: 10.1016/j.jneumeth.2005.12.018.
- [52] H. J. Motulsky and R. E. Brown, “Detecting outliers when fitting data with nonlinear regression – a new method based on robust nonlinear regression and the

false discovery rate,” *BMC Bioinformatics*, vol. 7, no. 1, p. 123, Mar. 2006, doi: 10.1186/1471-2105-7-123.

[53] A. Hamidovic, A. Dlugos, A. Skol, A. A. Palmer, and H. de Wit, “Evaluation of Genetic Variability in the Dopamine Receptor D2 in Relation to Behavioral Inhibition and Impulsivity/Sensation Seeking: An Exploratory Study With d-Amphetamine in Healthy Participants,” *Exp. Clin. Psychopharmacol.*, vol. 17, no. 6, pp. 374–383, Dec. 2009, doi: 10.1037/a0017840.

[54] S. Kobayashi and W. Schultz, “Influence of Reward Delays on Responses of Dopamine Neurons,” *J. Neurosci.*, vol. 28, no. 31, pp. 7837–7846, Jul. 2008, doi: 10.1523/JNEUROSCI.1600-08.2008.

[55] B. W. Balleine, M. R. Delgado, and O. Hikosaka, “The Role of the Dorsal Striatum in Reward and Decision-Making,” *J. Neurosci.*, 2007, doi: 10.1523/JNEUROSCI.1554-07.2007.

[56] M. V. Solanto, “Dopamine dysfunction in AD/HD: Integrating clinical and basic neuroscience research,” in *Behavioural Brain Research*, 2002, vol. 130, no. 1–2, pp. 65–71, doi: 10.1016/S0166-4328(01)00431-4.

[57] N. D. Volkow *et al.*, “Motivation deficit in ADHD is associated with dysfunction of the dopamine reward pathway,” *Mol. Psychiatry*, vol. 16, no. 11, pp. 1147–1154, Nov. 2011, doi: 10.1038/mp.2010.97.

[58] N. D. Volkow *et al.*, “Methylphenidate-elicited dopamine increases in ventral striatum are associated with long-term symptom improvement in adults with attention deficit hyperactivity disorder,” *J. Neurosci.*, vol. 32, no. 3, pp. 841–849, Jan. 2012, doi: 10.1523/JNEUROSCI.4461-11.2012.

- [59] N. W. Simon, B. S. Beas, K. S. Montgomery, R. P. Haberman, J. L. Bizon, and B. Setlow, “Prefrontal cortical-striatal dopamine receptor mRNA expression predicts distinct forms of impulsivity,” *Eur. J. Neurosci.*, vol. 37, no. 11, pp. 1779–1788, 2013, doi: 10.1111/ejn.12191.
- [60] L. R. Hammerslag *et al.*, “Adolescent impulsivity as a sex-dependent and subtype-dependent predictor of impulsivity, alcohol drinking and dopamine D 2 receptor expression in adult rats,” *Addict. Biol.*, vol. 24, no. 2, pp. 193–205, 2019, doi: 10.1111/adb.12586.
- [61] M. J. White, B. R. Lawford, C. P. Morris, and R. M. D. Young, “Interaction between DRD2 C957T polymorphism and an acute psychosocial stressor on reward-related behavioral impulsivity,” *Behav. Genet.*, vol. 39, no. 3, pp. 285–295, May 2009, doi: 10.1007/s10519-008-9255-7.
- [62] A. H. Ashok, Y. Mizuno, N. D. Volkow, and O. D. Howes, “Association of stimulant use with dopaminergic alterations in users of cocaine, amphetamine, or methamphetamine a systematic review and meta-analysis,” *JAMA Psychiatry*, vol. 74, no. 5. American Medical Association, pp. 511–519, May-2017, doi: 10.1001/jamapsychiatry.2017.0135.
- [63] I. A. Mendez *et al.*, “Self-administered cocaine causes long-lasting increases in impulsive choice in a delay discounting task.,” *Behav. Neurosci.*, vol. 124, no. 4, pp. 470–477, 2010, doi: 10.1037/a0020458.
- [64] S. Puig, N. Marie, N. Benturquia, and F. Noble, “Influence of cocaine administration patterns on dopamine receptor regulation,” *Psychopharmacology (Berl)*., vol. 231, no. 16, pp. 3131–3137, 2014, doi: 10.1007/s00213-014-3488-3.

- [65] S. M. Meiergerd, T. A. Patterson, and J. O. Schenk, “D2 Receptors May Modulate the Function of the Striatal Transporter for Dopamine: Kinetic Evidence from Studies In Vitro and In Vivo,” *J. Neurochem.*, vol. 61, no. 2, pp. 764–767, 1993, doi: 10.1111/j.1471-4159.1993.tb02185.x.
- [66] C. P. Ford, “The role of D2-autoreceptors in regulating dopamine neuron activity and transmission,” *Neuroscience*, vol. 282, pp. 13–22, Dec. 2014, doi: 10.1016/J.NEUROSCIENCE.2014.01.025.
- [67] D. Sulzer, S. J. Cragg, and M. E. Rice, “Striatal dopamine neurotransmission: regulation of release and uptake.,” *Basal Ganglia*, vol. 6, no. 3, pp. 123–148, Aug. 2016, doi: 10.1016/j.baga.2016.02.001.
- [68] Y. Schmitz, M. Benoit-Marand, F. Gonon, and D. Sulzer, “Presynaptic regulation of dopaminergic neurotransmission,” *J. Neurochem*, vol. 87, pp. 273–289, 2003, doi: 10.1046/j.1471-4159.2003.02050.x.
- [69] M. D. Condon *et al.*, “Plasticity in striatal dopamine release is governed by release-independent depression and the dopamine transporter,” *Nat. Commun.*, vol. 10, no. 1, pp. 1–15, Dec. 2019, doi: 10.1038/s41467-019-12264-9.
- [70] K. H. Krause, S. H. Dresel, J. Krause, C. La Fougere, and M. Ackenheil, “The dopamine transporter and neuroimaging in attention deficit hyperactivity disorder,” in *Neuroscience and Biobehavioral Reviews*, 2003, vol. 27, no. 7, pp. 605–613, doi: 10.1016/j.neubiorev.2003.08.012.
- [71] C. J. Pietras, D. R. Cherek, S. D. Lane, O. V. Tcheremissine, and J. L. Steinberg, “Effects of methylphenidate on impulsive choice in adult humans,” *Psychopharmacology (Berl.)*, vol. 170, no. 4, pp. 390–398, Dec. 2003, doi:

10.1007/s00213-003-1547-2.

[72] A. Z. Rajala, R. L. Jenison, and L. C. Populin, “Decision making: Effects of methylphenidate on temporal discounting in nonhuman primates,” *J. Neurophysiol.*, vol. 114, no. 1, pp. 70–79, May 2015, doi: 10.1152/jn.00278.2015.

[73] J. R. Yates *et al.*, “Effects of d-amphetamine and MK-801 on impulsive choice: Modulation by schedule of reinforcement and delay length,” *Behav. Brain Res.*, vol. 376, Dec. 2019, doi: 10.1016/j.bbr.2019.112228.

[74] E. Martinez, B. Pasquereau, G. Drui, Y. Saga, É. Météreau, and L. Tremblay, “Ventral striatum supports Methylphenidate therapeutic effects on impulsive choices expressed in temporal discounting task,” *Sci. Rep.*, vol. 10, no. 1, Dec. 2020, doi: 10.1038/s41598-020-57595-6.

[75] J. B. Justice, “Quantitative microdialysis of neurotransmitters,” *Journal of Neuroscience Methods*. 1993, doi: 10.1016/0165-0270(93)90097-B.

[76] T. M. Moschak and R. M. Carelli, “Impulsive Rats Exhibit Blunted Dopamine Release Dynamics during a Delay Discounting Task Independent of Cocaine History,” *eneuro*, 2017, doi: 10.1523/ENEURO.0119-17.2017.

[77] P. V. Piazza, F. Rougé-Pont, J. M. Deminière, M. Kharoubi, M. Le Moal, and H. Simon, “Dopaminergic activity is reduced in the prefrontal cortex and increased in the nucleus accumbens of rats predisposed to develop amphetamine self-administration,” *Brain Res.*, vol. 567, no. 1, pp. 169–174, Dec. 1991, doi: 10.1016/0006-8993(91)91452-7.

[78] S. B. Floresco, A. R. West, B. Ash, H. Moorel, and A. A. Grace, “Afferent modulation of dopamine neuron firing differentially regulates tonic and phasic

dopamine transmission,” *Nat. Neurosci.*, vol. 6, no. 9, pp. 968–973, Sep. 2003, doi: 10.1038/nn1103.

[79] F. Panin, A. Cathala, P. V. Piazza, and U. Spampinato, “Coupled intracerebral microdialysis and electrophysiology for the assessment of dopamine neuron function in vivo,” *J. Pharmacol. Toxicol. Methods*, vol. 65, no. 2, pp. 83–92, Mar. 2012, doi: 10.1016/j.vascn.2012.01.003.

[80] E. J. Pérez-Valenzuela, M. E. Andres Coke, A. A. Grace, and J. A. Fuentealba Evans, “Adolescent exposure to WIN 55212-2 render the nigrostriatal dopaminergic pathway activated during adulthood,” *Int. J. Neuropsychopharmacol.*, Jul. 2020, doi: 10.1093/ijnp/pyaa053.

[81] E. Martinez, B. Pasquereau, Y. Saga, É. Météreau, and L. Tremblay, “The Anterior Caudate Nucleus Supports Impulsive Choices Triggered by Pramipexole,” *Mov. Disord.*, vol. 35, no. 2, pp. 296–305, Feb. 2019, doi: 10.1002/mds.27898.

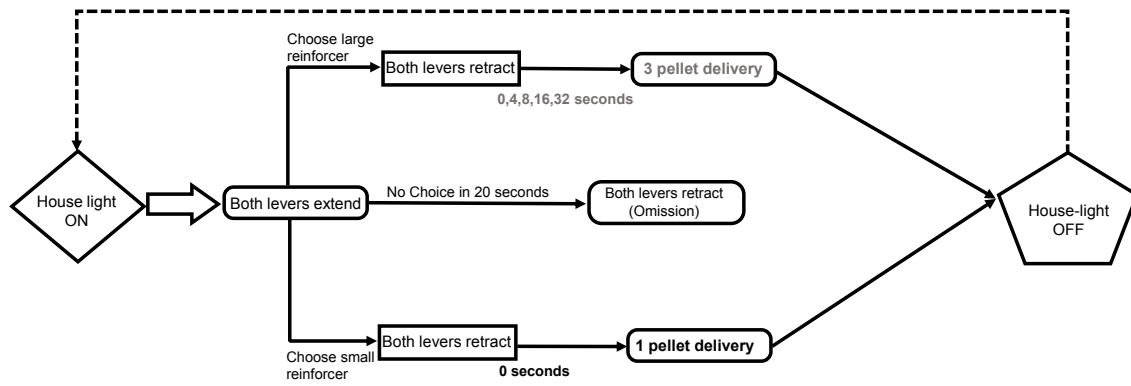


Figure 1: Delay discounting task design. Scheme of a single free-choice trial on the delay discounting task used in the present study.

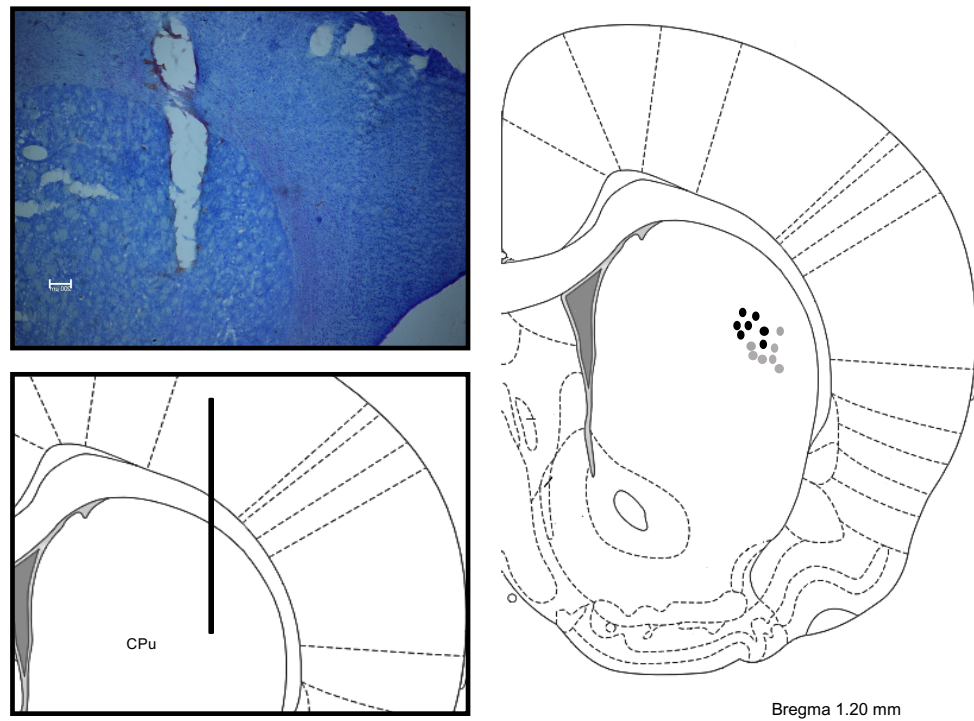


Figure 2: Anatomical placements of the microdialysis probe in DLS. **Left,** Photo of representative microdialysis probe placement in DLS, and map showing the microdialysis probe placement (black line) of the representative photo. **Right,** schematic, coronal section of the rat brain showing the placement of the microdialysis probe for LI (black dots) or HI (gray dots) rats in the No-net-Flux microdialysis experiment. Brain sections correspond to the atlas of Paxinos and Watson [41].

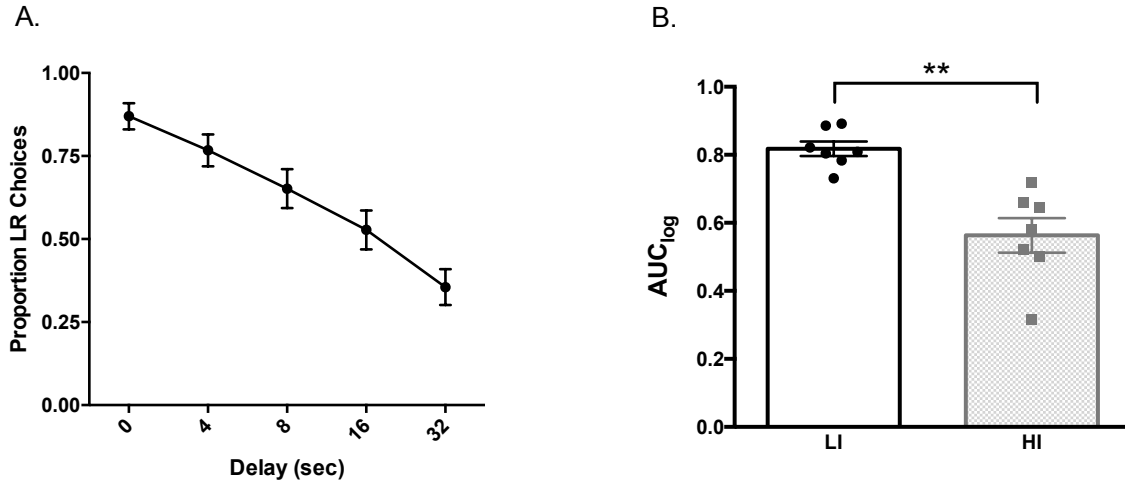


Figure 3: Individual differences in the basal level of impulsive choice. (A) Delayed-discounting curve for all animals (n=14) and (B) Rats classified as LI (n=7) show a higher AUC_{log} compared with rats classified as HI (n=7), ** represents $p < 0.05$ according to unpaired t-test.

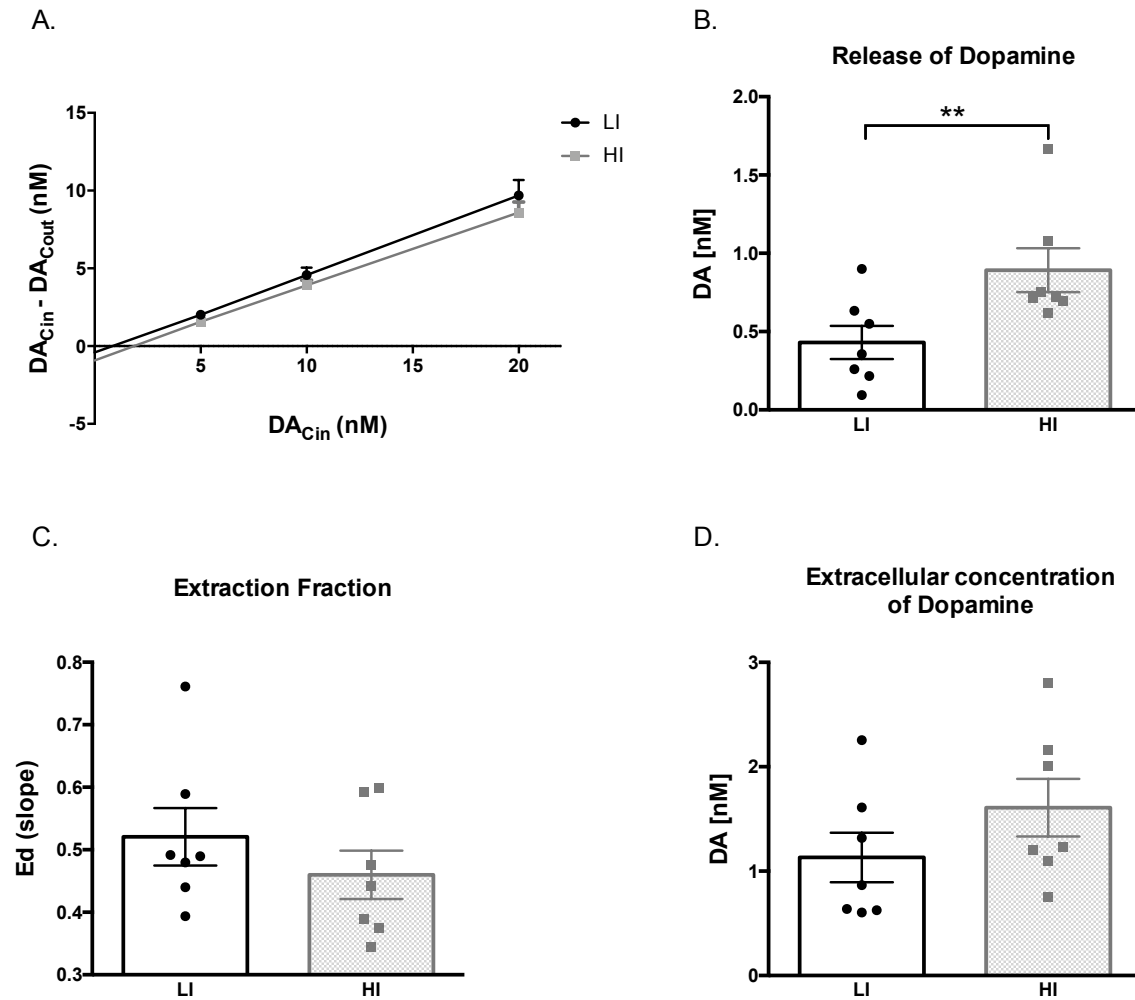


Figure 4: Animals classified as high impulsive in DDT showed higher DA release in DLS. In vivo No-net-flux microdialysis in anesthetized rats classified as HI (n= 7) and LI (n=7). **(A)** The plot of average gain or loss of DA ($C_{in}-C_{out}$) as a function of perfusate DA (C_{in}) and the linear regression of LI (black line) and HI (gray line) rats. The slope of the regression line represents the Ed. The point when DA is not perfused ($C_{in}= 0$) represents the DA release. When no DA is gained or lost from the perfusate ($C_{in}-C_{out}= 0$) represents an estimate of DA C_{ext} . **(B)** Bar graph of DA release, **(C)** Ed, and **(D)** DA C_{ext} . ** represents $p < 0.05$ according to an unpaired t-test.

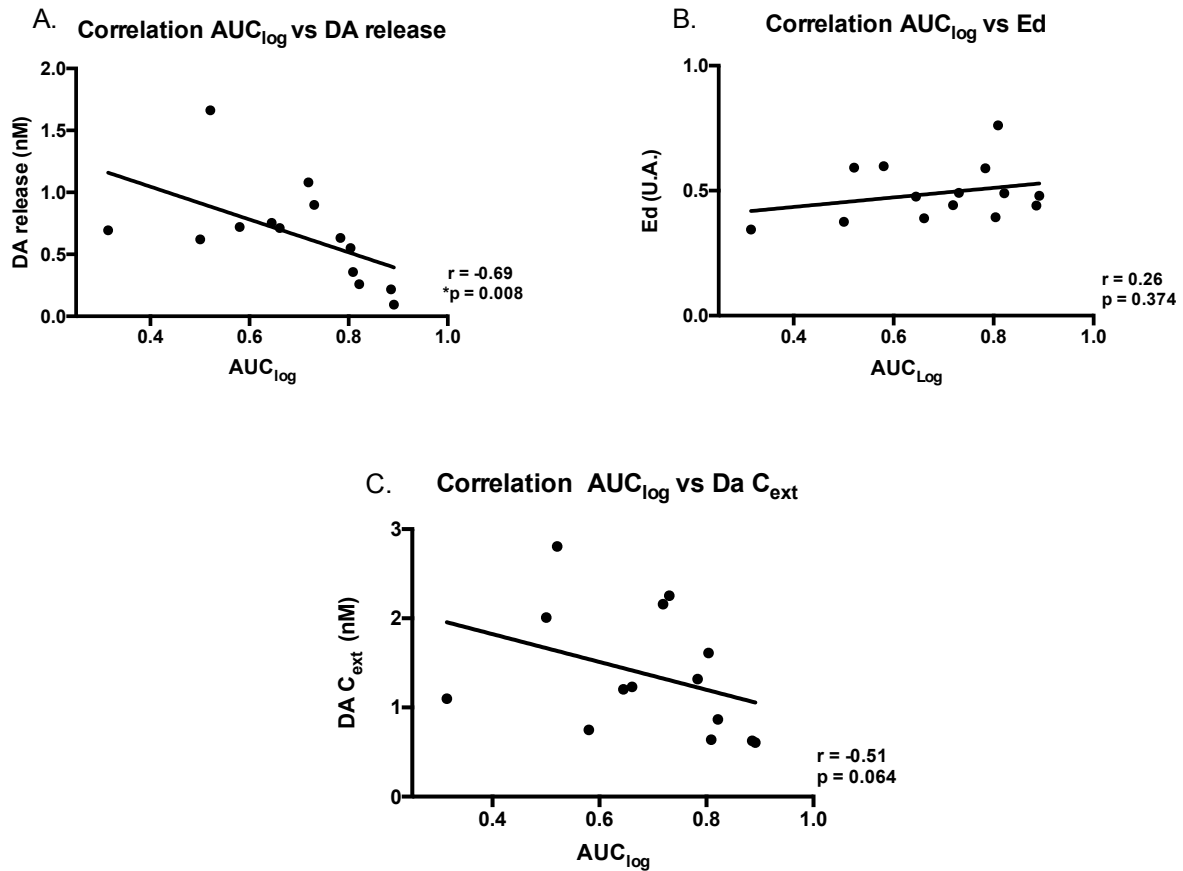


Figure 5: Correlation between impulsive choice level measured by AUC and basal dynamic of DA in DLS. (A) Correlation between AUC and DA release. A significant, negative correlation was observed ($r = -0.69$; $*p = 0.008$ according to Spearman correlation; $n = 14$). **(B)** Correlation between AUC and Ed. A non-significant correlation was observed ($r = 0.26$; $p = 0.374$ according to Spearman correlation; $n = 14$). **(C)** Correlation between AUC and DA C_{ext} . A non-significant correlation was observed ($r = -0.51$; $p = 0.064$ according to Spearman correlation; $n = 14$).

5 Chapter 2: Effect of OXT-R activation on the nigro-striatal pathway during delay discounting task, a study of impulsive choice behaviors

Macarena Moreno^{1,3}; Tania Araya¹; Stan B. Floresco²; Georgina. M Renard³ and José Antonio Fuentealba^{1*}.

Author Affiliations:

¹ Department of Pharmacy and Interdisciplinary Center of Neuroscience, Faculty of Chemistry and Pharmacy, Pontificia Universidad Católica de Chile, Santiago, Chile.

² Department of Psychology and Brain Research Center, University of British Columbia, Vancouver, British Columbia, V6T 1Z4 Canada.

³ Centro de Investigación Biomédica y Aplicada (CIBAP), Escuela de Medicina, Facultad de Ciencias Médicas, Universidad de Santiago de Chile, Santiago, Chile.

*Corresponding Author:

José Antonio Fuentealba, Department of Pharmacy, Faculty of Chemistry and Pharmacy, Pontificia Universidad Católica de Chile, Santiago, Chile.

Email: jfuentea@uc.cl

Manuscript in preparation

Highlights

- The bilateral activation of OXT-R in SNpc, induces a decrease in impulsive choice in HI animal during DDT.
- A slight but significant DA_{ext} increase in DLS was observed post OXT-R activation in SNpc.
- The activation of OXT-R in SNpc, induce a non-significant change in the DA_{ext} in SNpc.

5.1 Abstract

Impulsive choice has been used as a predictor of several neuropsychiatric disorders, including personality disorders, attention deficit hyperactivity disorder (ADHD), and addiction. All these pathologies are linked to a dysfunction in dopaminergic transmission. We recently showed that dorsal striatal dopamine (DA) signaling contributes to individual differences in impulsive choice. Because oxytocin (OXT) modulates the activity of mesencephalic dopaminergic neurons, we wonder whether activation of the oxytocin receptor (OXT-R) in the substantia nigra pars compacta (SNpc) also contributes to modulate impulsive choice. To fully understand how the OXT system modulates the DA release from mesencephalic neurons, we also studied the consequences of the activation of OXT-R in the SNpc on DA extracellular (DA_{ext}) level in the nigro-striatal pathway. Firstly, Adult male rats were well-trained on a delay discounting task, where they chose between a smaller (1 pellet) reward delivered immediately, or a larger 4-pellet reward delivered after a delay (0-45 s). On separate test days, they received the microinfusion of the non-peptide agonist WAY267464 (at a dose of $3\mu g/0.5\mu l$) in SNpc. These treatments decreased impulsive choice in high impulsive (HI) rats, displaying a greater preference for larger, delayed rewards. Secondly, using in vivo microdialysis revealed that the same agonist WAY267464 ($5\mu M$) perfusion in SNpc induces a slight increase in DA_{ext} level in the dorsolateral striatum (DLS) but does not induce a change in DA_{ext} level in SNpc. Together these findings suggest that activation of the OXT-R in the SNpc influences delay-related decision making by subtle changes in the striatal dopamine transmission.

Keywords: Delay discounting task; impulsive choice; dorsolateral striatum; dopamine; oxytocin.

Abbreviations: DDT, delay discounting task; DLS, dorsolateral striatum; HI, high impulsive; LI, Low impulsive; OXT-R, oxytocin receptor; SNpc, substantia nigra pars compacta.

5.2 Introduction

The dopaminergic and oxytocinergic systems alteration is present in a variety of neuropsychiatric disorders, such as schizophrenia, ADHD, and addiction [1–4], which are disorders that are related to a high level of impulsive behaviors [5–8]. We recently showed that DA transmission changes in DLS contribute to individual differences in impulsive choice; specifically, we find that DA release is augmented in the DLS of rats classified as HI, suggesting that a hyper-activated nigro-striatal pathway contributes to impulsive choice. This result is consistent with clinical and preclinical evidence that supports the notion that variability in DA neurotransmission function is a significant contributor to individual differences observed in impulsive choice [9,10]. Moreover, the clinical approach using neuroimaging techniques showed that low DA D_2/D_3 receptor availability in the ventral tegmental area (VTA) and substantia nigra (SN) is associated with a highly impulsive phenotype [11]. Preclinical evidence indicate that administration of the selective catecholamine uptake inhibitor 3,4-Methylenedioxypyrovalerone (MDPV) increase impulsive choice [12]. However, existing evidence proposes that low and high DA release could contribute to impulsive choice, accordingly to the inverted-U shape pattern observed between DA signaling and cognitive function [13–15]. Therefore, it is necessary to study the neurochemical and pharmacological performance at the impulsivity curve extremes. Approaches have been proposed that using psychostimulant drugs would decrease impulsive behavior in subjects with low DA concentration but could worsen impulsivity in subjects with elevated DA concentration [14,16]. Interestingly, in the DA and OXT interaction context, clinical studies showed a significant association between

basal plasma OXT and the Karolinska Scale of Personality (KSP) factor impulsiveness, suggesting positive correlation between impulsiveness and plasma OXT level male psychiatric outpatients [17]. On the other hand, resents studies showed that the intranasal OXT administration moderated impulsive behavior in subjects with a single nucleotide polymorphism (SNP) rs2254298 genotype [18]. Even so, the interaction of dopamine and OXT systems in impulsive behavior is still unknown.

OXT is a neuropeptide synthesized by neuron of hypothalamic paraventricular nucleus (PVN) and supraoptic nucleus (SON) and is transported along the axons projected into the neurohypophysis from where they are spread in the blood stream [19,20]. The principal role of OXT release into the blood is linked to parturition and lactation [21–23]. On the other hand, there are long-range axonal projection of OXT neurons in the hypothalamus to the extra-hypothalamic regions of the brain, such as the prefrontal cortex (PFC), nucleus accumbens (NAcb), lateral septum, amygdala, and SN [24–27]. In these extra-hypothalamic regions, OXT is capable of acting as a neurotransmitter and/or as a neuromodulator of cognitive processes and complex social behavior such as attachment, exploration, peer linking and social recognition [28,29], as well as in non-social behaviors, such as stress, anxiety, and aggressiveness [30]. It has been proven that the aforementioned prosocial behaviors can be also affected by a deep alteration in the dopaminergic pathways [31–33].

There is evidence of both the innervation and expression of OXT-R in different regions linked to the dopaminergic transmission, such as SN and VTA [26,34,35], which may be pointing to a regulatory role of OXT in different behaviors linked to the DA system

[35,36]. The OXT-R is a G-protein-coupled receptor (GPCRs), coupled to G_q , which active the phospholipase C (PLC) pathway to increase intracellular Ca^{2+} stores release [23]. It has also been shown that there exists a postsynaptic co-localization between the DA type D_2 -R with the OXT-R in brain regions that receive dopaminergic projections as NAcB and the DLS. These receptor crosstalk and/or heteromeric receptor complex (D_2 -R–OXT-R) could represent a molecular mechanism for OXT through positive allosteric regulation in receptor-receptor interaction, increases dopaminergic signaling, and contributes to the induce changes in social and emotional behavior [37,38].

It has also been shown that there exists a bidirectional modulation between dopaminergic and oxytocinergic systems [39–41]. Therefore, it has been established that both dopaminergic and oxytocinergic pathways converge in the PFC, the striatum, and the ventral tegmental areas [36,42,43]. In this context, the neurophysiological and behavioral approach has shown that DA intraventricular injections increase the frequency and amplitude of oxytocinergic neurons in PVN [39], in line with these, it has been shown that OXT release from PVN in VTA enhances the activity of a specific population of DA neurons that influence social interaction [36]. On the other hand, Xiao et al., [26] showed that OXT release in midbrain DA regions as VTA or SNpc exert a differential effect, where the OXT release in VTA induces an increase in the VTA-DA firing rate (FR). Moreover, the OXT release in the SNpc induces a heterogeneous effect, depending on the topographical location into SNpc, where the OXT release in lateral-SNpc induces a decrease in the SNpc-DA FR. These studies suggest that the action of OXT-VPN is essential for the reinforcing the component of social interaction via VTA DA neurons [36], and OXT-driven inhibition of DA neuron in SN can dampen non-social behavior,

where OXT infusion in SN decrease locomotor activity [35]. Nevertheless, the changes in the population neuronal activity in SNpc post OXT-R activation area associated with DA release changes in DLS, and their behavioral effect have not been studied.

Therefore, we directly investigated the role of OXT in the SNpc on impulsive choice behavior and on the modulation of nigro-striatal dopaminergic transmission. For behavioral studies, animals were well-trained on a delay discounting task (DDT), and on the test days, animals received bilateral microinfusions of the OXT-R agonist (WAY267464 dihydrochloride, at a dose of 3mg/0.5ml) in the SNpc. An in vivo microdialysis in adult male rats were performed to study the effect of the perfusion of WAY267464 in SNpc on DA extracellular (DA_{ext}) levels in the DLS and SNpc, respectively. Our result showed that bilateral activation of OXT-R in SNpc induces a decrease in impulsive choice in highly impulsive animals. Interestingly, the activation of OXT-R in SNpc induces a moderate increase in the DA_{ext} in SNpc, suggesting that activation of the OXT-R in the SNpc influences delay-related decision making by subtle changes in the striatal dopamine transmission.

5.3 Methodology

5.3.1 Animals

Adult male *Long-Evans* rats (Charles River Laboratories, St Constant Quebec, Canada) weighing between 275 to 300g at the start of the experiment were used in behavioral experiments. Rats were maintained with 14 g food per day and water *ad libitum*. All the animals were put in pairs in each of the houses, under a 12-hour light-dark cycle (lights were turned on at 8:30 AM) and controlled temperature ($21\pm 1^{\circ}\text{C}$).

Adult male *Sprague Dawley* rats (weighing between 300 and 400 g) were used in neurochemical experiments. They were housed in pairs in ventilated cages with an Air Handling Unit (Tecniplast, West Chester, PA) under controlled conditions of temperature ($22\pm 1^{\circ}\text{C}$), and humidity ($50\pm 10\%$), and were maintained under 12-hour light-dark cycle (lights on between 07:00 AM and 5:00 PM). Experiments were conducted during the light period. Rats were handled for seven days before starting the experiments. The food and water access were *ad libitum*.

All procedures were in strict accordance with the guidelines published in the 8th edition of the NIH Guide for the Care and Use of Laboratory Animals as well as principles presented in the *Guidelines for the Use of Animals in Neuroscience Research* by the Society for Neuroscience. Furthermore, all procedures were approved by the ethics committee of the Pontificia Universidad Católica de Chile as well as by the Canadian Council of Animal Care and the Animal Care Committee of the University of British Columbia.

5.3.2 Behavioral Testing

5.3.2.1 Apparatus

Sessions were conducted in ventilated, sound-attenuating enclosures containing an operant conditioning chamber with an interior space of measures 30.5×24×21 cm (Med-Associates, St. Albans, VT, USA). Each box has a house-light which is turned on at the beginning of the tests, two levers associated to another light in the chamber and a dispenser of food reinforcement (sucrose pellet, 45mg; Bioserv, Frenchtown, NJ, USA) between them. All experimental data was recorded by an IBM personal computer connected to the chamber via an interface using the Med-PC-IV software (Med-Associates, St. Albans, VT, USA).

5.3.2.2 Procedures

Rats were tested in the behavioral procedures as described below. All procedures were adapted from Eubig P et al., [44].

5.3.2.2.1 Initial lever press training

Approximately after PND 35, a shaping program was used to train rats to begin pressing the response levers. Both levers were extended at the onset of the session. As described in Eubig P et al., [44] the delivery was programmed according to a 3-min fixed-time schedule, in which a food pellet was dispensed every 3 min. Also, one pellet was delivered with every lever press. Sessions finished after either 60 min elapsed, or 100 reinforcements were delivered. Criterion for advancement to the next phase of training was 100 lever presses, performed for three consecutive sessions.

5.3.2.2.2 Fixed ratio training

The second training phase began between PND 39-40. During this phase, only one lever was extended, and the corresponding cue light was illuminated. After five lever presses, the extended lever was retracted, and the opposite lever was extended. Every response resulted in the delivery of a food pellet (i.e. a fixed ratio 1 contingency). After 100 reinforcements across three consecutive sessions, rats continued to the next training phase.

5.3.2.2.3 Delay discounting training and testing

The DDT training phase began approximately after PND 45. The phase consisted of 40 trials for each session. Two levers with different reinforcers were presented and were always accompanied by an illuminated cue light: the lever press of the larger reinforcer (LR) resulted in 4 pellets and the small reinforcer (SR) lever resulted in 1 pellet. There was no delay in the delivery of either the small or large reinforcers in this phase. The session finished after 40 lever presses between large or small reinforcers. Criterion for advancement to the next phase of training required an 80 percent of preference for the larger reinforcer.

The DDT testing began in darkness with the levers retracted (blackout). After 10 minutes of blackout, the house-light turned on and the session started. One session consisted of five blocks comprising 12 trials each. The duration of the trials was of 60 seconds. The first two trials of each block were forced-choice trials in which the LR lever and the SR lever were alternately introduced. Ten free-choice trials followed the force trials in each block. At the beginning of the free-choice trial, the lights over the levers were turned on, and the levers appeared. If a lever was pressed, both levers were retracted. If the rat pressed

the SR lever, one pellet was immediately delivered, and both levers were retracted. If the LR lever was chosen, the stimulus light blinked during the delay and, following the delay, three pellets were delivered. Delays to the LR were increased from 0s in block 1 to 15s in block 2, 30s in block 3, 45s in block 4. Failure to choose a response within 20s resulted in the lever retracting and the trial was considered as an omission. The rats were trained five times per week for approximately ten weeks (50 sessions) in this task until stable performance was achieved. They then were subjected to bilateral implantation of guide cannula in SNpc.

5.3.2.3 Bilateral microinfusion in the SNpc

Rats were given a subanesthetic dose of ketamine (50 mg/kg) and xylazine (4 mg/kg) and were maintained on isoflurane for the whole duration of the surgery. Bilateral implantation of guide cannulas (22G) was performed on the SNpc, and these guides were fixed to the skull with stainless steel screws and dental cement. The following stereotaxic coordinates were used: -5.6 AP from bregma, ± 2.1 ML and -8.1 DV, according to Paxinos and Watson [45]. Animals were allowed to recover for one week before resuming the behavioral test.

The microinfusions procedure, was adapted from Winstanley C.A [46]. Once the impulsive choice behavior was re-established post-surgery animals received a mock infusion prior to a training session to familiarize them with the procedure prior to the actual microinfusion test day (see, figure 1). Injectors were placed inside the guide cannula for 2 minutes, but no microinfusion was administrated. Rats were then placed in their home cage for 15 min until their training sessions started. The animals received their first

microinfusion 1 to 3 days after the mock injection. The animals trained to DDT received two bilateral microinfusions with either a saline or a non-peptide OXT-R agonist (WAY267464, 3 μ g/0.5 μ l), according to a counterbalanced design [25,47]. For all animals, the saline or drug was infused at a volume of 0.25 μ l and the microinfusion was made using a dual channel microinfusion pump (Harvard Apparatus, Holliston, MA, USA).

5.3.3 Delay discounting analysis

5.3.3.1 The logarithmic area under curve (AUC_{log})

Delay-discounting curves were obtained by plotting the proportion of LR choices (the number of LR choices divided by the total number of successful trials) of each block versus each delay (sec). The area under the delay-discounting curve (AUC) was used as a measure of impulsive choice [48,49]. The AUC was calculated using a log-transformation of delays (AUC_{log}) to avoid the disproportionate contribution at long delays in the delay-discounting curves [50]. Specifically, each delay was transformed to the logarithm of the delay divided by the bigger logged delay. To avoid the problem of calculating the logarithm of zero, a small amount (value=1) was added in each delay before the transformation. We use GraphPad Prism to calculate AUC_{log} (version 5, GraphPad Software, Inc.; La Jolla, CA). Higher AUC_{log} values indicate lower discounting by delay (i.e., preference for larger rewards, indicating lower impulsivity), while lower AUC_{log} values correspond to higher discounting by delay (i.e., preference for small and immediate rewards, indicating higher impulsivity) [48].

5.3.3.2 Behavioral Stability and Classification

To assess behavioral stability, we compared the AUC_{log} of five consecutive sessions (week 1) with the five following sessions (week 2). An animal was considered trained when the difference between weeks was not statistically significant. For each animal, a stable AUC_{log} value was calculated using the average of the last five sessions. Rats were classified as low impulsive (LI) and high impulsive (HI) based on the median split of AUC_{log} (see figure 3A and B). The minimum sample size was calculated based on a priori power analysis [51]. The input parameters for behavioral experiments were the following: two-tails, $\alpha = 0.05$, power = 0.80, and ratio $N1/N2 = 1$. The effect size was based on preliminary DDT results, suggesting a minimum sample size of $n = 4/\text{group}$ for the experiment considering a Cohen's d effect size of 2, using G*Power statistical analysis software Version 3.1.9.4 [52].

5.3.3.3 Omission and latencies

Latency was defined as the time since the levers were extended until the rat pressed one of them during free-choice trials in each block. An omission was considered as a failure to choose a response within 20s after lever appearance. For each animal, we recorded the latency and the number of omissions in the 4 daily blocks in free-choice trials. Final latency and omission values were calculated averaging total latency and omissions of the last five sessions.

5.3.4 Conventional Microdialysis

To study the effect of OXT-R activation in the nigro-striatal dopaminergic pathway induced by perfusion of a non-peptide OXT-R agonist (WAY267464), due to its chemical characteristics linked to the stability of the molecule and half-life. Altogether, it is an excellent candidate to cross the blood-brain barrier [53,54]. *In vivo* microdialysis in anesthetized animals was performed at approximately PND 60-75. Rats were deeply anesthetized with urethane (1.5 g/kg i.p.) and placed in a stereotaxic apparatus. Then, two microdialysis probes (CMA-11 Microdialysis, Holliston, MA, USA) were implanted into the brain: one in the DLS, using the following coordinates according to the atlas of Paxinos and Watson (2009): +1.4 AP, +3.2 ML, -4.8 DV; and the other in the SNpc, using the following coordinates: -5.0 AP, +2.1 ML y -8.1 DV. During the surgery, the body temperature of the animals was maintained and monitored.

5.3.4.1 Microdialysis Protocol

The microdialysis probe in the DLS was continuously perfused with a Krebs ringer phosphate solution (KRF; containing NaCl 120 mM; KCl 2.4 mM; CaCl₂ 1.2 mM; 0.9 mM NaH₂PO₄; Na₂HPO₄ 1.4 mM, pH = 7.4) at a flow rate of 2 µl/min using a Harvard infusion pump. The microdialysis probe in the SNpc was perfused with KRF during a stabilization period of 100 min, and three consecutive samples were collected every 5 min for determination of an average DA basal level in SNpc and DLS. After that, to corroborate the activation of the nigro-striatal pathway, two KRF potassium solutions (70 mM) were perfused in the SNpc for 5 minutes; only experiments where perfusion of K⁺ in SNpc induced an < 20% increase in DA_{ext} levels in the DLS were used. Meanwhile, the collection of samples also continued every 5 minutes in the SNpc and DLS. After that,

three different concentrations of KFR/WAY267464 (5 μ M, 20 μ M y 50 μ M) were perfused for 10 minutes [54]. Furthermore, the nonpeptide antagonist for OXT-R (LY369 100 mM) was also perfused (adapted doses from C.E king et al., [55]). Three consecutive samples were collected in each experimental condition. The collected perfusion samples were maintained on ice during the experiment and stored at -20°C until analysis.

5.3.4.2 Analysis of Dialysate Dopamine

Ten ml of the dialysate was injected using a Rheodyne injector valve into an HPLC system (BASi America, West Lafayette, IN). The mobile phase contained 100 mM NaH_2PO_4 , 1 mM EDTA, 1 mM octane-1-sulfonic acid sodium salt, and 3% acetonitrile (pH 3.0), and was pumped at a flow rate of 700 $\mu\text{L}/\text{min}$. The potential of the amperometric detector was set at 650 mV. Under these experimental conditions, the retention time for DA was 6 min. Dialysate samples were analyzed by comparing the peak area and elution time with reference standards (ChromNAV 2.0 Jasco Co, ltd, Tokyo, Japan).

5.3.5 Histology

After the behavioral test and microdialysis experiments, rats were decapitated under deep anesthesia (urethane 1.5 g/kg i.p.), and brains were extracted and cleaned with a saline solution (NaCl 0.9%). Brains were stored in 4% paraformaldehyde (PFA). At least two days before slicing, brains were cryoprotected using a 30% sucrose solution. For probe implantation, brains were frozen and coronally sliced in sections of 50 μm . Slices were stained with cresyl violet and the probe placement was observed in a light microscope

using the atlas of Paxinos & Watson for rats [41]. Only data coming from correct probe placements was considered for further analysis (see figure 1 and 4).

5.3.6 Data Analysis

5.3.6.1 Statistical Analyses

All statistical analyses were carried out using Prism 5.0 GraphPad Software. Each group of data was analyzed with the nonlinear regression ROUT method (Q= 1%) [56] to identify outliers and exclude them from the statistical analysis and graphs. No outliers were found in these studies. Resultant data were analyzed using unpaired t-test, one and two-way ANOVA with Bonferroni post-test when appropriate (all data are reported as mean \pm SEM).

5.4 Results

5.4.1 Effect of SNpc administration of WAY267464 on impulsive choice during delay discounting task

To assess the effect of OXT-R activation in DLS on impulsive choice during DDT, adult male *Long-Evans* rats were well-trained on a DDT. On separate test days, they received bilateral infusions of the OXT-R agonist (WAY267464) at a dose of $3\mu\text{g}/0.5\mu\text{l}$ in the SNpc (see experimental design in figure 1A). The bilateral infusion of WAY267464 in SNpc, which induces a decrease in impulsive choice, showed significant difference in the number of choices during the delays of 15, 30 and 45 seconds between saline and WAY267464 groups according to two-way ANOVA ($n=8$; Bonferroni test: delay 15: Saline: 0.42 ± 0.07 vs WAY267464: 0.62 ± 0.07 $p=0.03$; Delay 30: Saline: 0.25 ± 0.07 vs WAY267464: 0.47 ± 0.07 $p=0.01$; Delay 45: Saline: 0.19 ± 0.07 vs WAY267464: 0.38 ± 0.07 $p=0.04$, see figure 2A). The bilateral infusion of WAY267464 in the SNpc induced an increase in the area under the curve (AUC_{\log}) pointing to a higher preference to the larger reinforcement ($n=8$; t-test $p=0.02$, see figure 2B). The response latency and omissions in control (saline perfusion) and agonist OXT-R (WAY267464) experimental conditions showed non-significant differences between the two experimental conditions (latency: $n=8$ t-test; $p=0.35$, see figure 2C and omission: $n=8$; t-test; $p=0.41$; see figure 2D). Finally, this experiment showed that bilateral OXT-R activation in SNpc induced a decrease in impulsive choice behavior during DDT.

5.4.2 The effect of bilateral OXT-R activation in SNpc depends on basal impulsivity

An exploratory analysis was performed from the previous results, where we used the behavioral classification using AUC_{log} values during DDT as previously described (see methodology). From these, two groups were formed: LI (n =4) and HI (n =4) (see figure 3A and B). The effect of OXT-R activation is dependent on basal impulsivity levels, can be observed in animals previously classified as LI or HI (see Figures 3C and 3E). The bilateral OXT-R activation in LI animals showed a non-significant difference in the number of large reinforcement choices during delays of 15, 30, and 45 sec observed between saline and WAY267464 condition (according to Two-Way ANOVA; n= 4; Bonferroni test: delay 15: Saline: 0.65 ± 0.09 vs WAY267464: 0.72 ± 0.09 $p > 0.99$; Delay 30: Saline: 0.45 ± 0.09 vs WAY267464: 0.66 ± 0.09 $p = 0.16$; Delay 45: Saline: 0.39 ± 0.09 vs WAY267464: 0.51 ± 0.09 $p = 0.91$, see figure 3C) and non-significant different in the AUC_{log} value between saline and WAY267464 conditions (n= 4; t-test $p = 0.19$, see figure 3D). In HI animals, the bilateral OXT-R activation induced a decrease in impulsive behavior, with significant differences for 15 seconds delay (according to Two- Way ANOVA, Saline: 0.17 ± 0.11 vs WAY267464: 0.52 ± 0.11 $p = 0.05$, see figure 3E) and a non-significant difference in the AUC different in the AUC_{log} value between saline and WAY267464 conditions (n= 4; t-test $p = 0.05$, see figure 3F).

5.4.3 Effect of non-peptide specific OXT-R agonist (WAY267464) administration in SNpc on DA_{ext} levels in DLS

To fully understand how the OXT system modulates the DA release from mesencephalic neurons, we also studied the consequences of the activation of OXT-R in the SNpc on DA_{ext} level in the nigro-striatal pathway. We perfused a non-peptide agonist WAY267464 in the SNpc and collected samples from the SNpc and the DLS (see figure 4). First, to corroborate the activation of the nigro-striatal pathway, we perfused a depolarizing stimulus of K⁺ solution (70 mM) in SNpc and measured DA_{ext} levels in SNpc and DLS. The only experiments used were those where perfusion of K⁺ in SNpc induced an > 20% increase in DA_{ext} levels in the DLS (according to paired t-test: DLS: p= 0.004 and SNpc: p= 0.038; n=5, see figure 5A and 5B). To verify the oxytocinergic endogenous tone in SNpc, we used an OXT-R antagonist LY369 (100 mM). The perfusion of OXT-R antagonist in SNpc did not produce any change in DA_{ext} levels in the DLS and SNpc (according to paired t-test: DLS: p= 0.64; SNpc: p= 0.65, n=5, see figure 6A). Subsequently, to study the effect of OXT-R activation, we perfused different concentrations of WAY267464 (5, 20, and 50 µM). The experiment showed that the effect of OXT-R activation in SNpc is dose-dependent, and low doses (5 µM) generated a moderated but significant effect, inducing a subtle increase in DA_{ext} levels in the DLS (according to paired t-test; p= 0.05; n= 5, see figure 6 B) but a non-significant change in SNpc (according to paired t-test; p= 0.64 n=5). On the other hand, high doses (20-50 µM) did not have a significant effect in both DLS (according to paired t-test; 20 µM p= 0.08; 50 µM p= 0.15; n=5, see figure 6B) and SNpc (according to paired t-test; 20 µM p= 0.42; 50 µM p= 0.25; n=5, see figure 6C).

5.5 Discussion

Various approaches have suggested that hypothalamic oxytocinergic projection modulates midbrain DA regions [26,36], but the neurochemical DA dynamic and behavioral consequences of this potential interaction are not yet completely known. Here, we study the activation of OXT-R in the SNpc during DDT and its contribution to impulsive choice behavior. Moreover, we examine the effect of OXT-R activation in SNpc on DA_{ext} level in the nigro-striatal pathway.

Our behavioral experiment indicated that the bilateral activation of OXT-R by microinfusion of WAY267464 induces a decrease in impulsivity reflected in an increase in the number of choices for the LR. Due to the individual differences present in impulsive choice, this research showed that in animals classified as HI, the bilateral activation of OXT-R in SNpc decreases impulsive behaviors compared to the LI group. The difference between groups suggests that the OXT-R activation in SNpc could induce a differential effect in the nigro-striatal pathway, which could depend on the DA basal dynamic that underlies individual differences present in impulsive choice behavior. These results are in line with previous results (see chapter 1), where individual differences in impulsive choice level during DDT are accompanied by a differential change of DA in DLS. The individual differences in impulsive choice behavior could be due to changes in the expression and activity of D₂-R presynaptic in DLS. Several clinical and preclinical research has shown that prolonged exposure to methamphetamine, amphetamine, or cocaine reduced D₂-R expression in the midbrain region, such as the SN, VTA, and striatum, leading to high levels of impulsive choice [11,57,58]. It is then possible to propose that the differences

observed in HI animals are due to a loss of the inhibitory tonic control of D₂-R [59]. This basal neurophysiological condition could be modulated differentially by the activation of OXT-R in the nigro-striatal pathway. Interestingly, Xiao et al., [26] showed that the OXT-R activation in SNpc induces a heterogeneous set of responses, dependently on topographic location into SNpc; the dorsolateral OXT-R activation induces a decrease in the activity of dorsolateral SNpc DA neurons via inhibitory circuits, but in the medial region of SNpc did not find a significant change in medial SNpc DA neurons. These different topographical effects are generated by the high density for OXT-R in the dorsolateral region than the medial region of SNpc. Hence, we suggest that the mechanism associated with the bilateral activation of OXT-R could be linked to a decrease in the dopaminergic activity due to the activation of GABAergic neurons that express OXT-R. In this context, Xiao et al., [26] identified a high co-localization of OXT-R in cell Vgat⁺ (vesicular GABA transporter) in dorsolateral SNpc, compared to co-localization of the canonical marker of DA neurons, tyrosine hydroxylase (TH). In this sense, the increment in GABA signaling could be an indirect mechanism that provokes a hyperpolarization of membrane potential, causing a decrement in the DA FR in SNpc [26,60], producing a decrease of DA levels in the DLS [61,62]. This modulation could regulate the behavior by altering the tonic activity of DA striatal tone, helping to recover neurochemical homeostasis linked to impulsive behavior. Alternatively, we speculated that the decrease in impulsive choice level might be due to bilateral OXT-R activation in SNp dorsolateral, improving the somatodendritic D₂-R affinity for DA. It has been shown that an increase in the concentration of ligands for OXT-R induces an increase in the affinity of D₂-R for

DA [37,63–65], which induced changes in the dopaminergic activity during the presence of a salience stimulus, facilitating advantageous decision making [66,67].

In these behavioral experiments, a direct measure of locomotor activity and motivation was not collected. However, during DDT, the response latency and omissions were recorded, which could be used as indirect locomotor activity and motivation measures, respectively [68,69]. Therefore, we analyzed the changes in response latencies to assess whether the motor activity variations through the delay blocks were not related to the different levels of impulsive choice between experimental conditions. On the other hand, we analyzed the number of omissions to assess whether the variations in motivational behavior were not related to the experimental condition in the impulsive choice levels. Thus, these results indicate that the microinfusion of WAY267464 in SNpc only affect neurobiological mechanisms linked to decision making.

Exist evidence that OXT neuron from the PVN project to VTA and SNpc [26,70]. To identify if there is a tonic control of OXT, we used a selective non-peptide antagonist for the OXT-R receptor LY369 (100mM) in SNpc. This antagonist perfusion induced non-significant differences in the DA_{ext} level of the SNpc and the DLS, which suggests that no endogenous OXT tone exists in the SNpc. This finding is consistent with evidence that proposes that the dynamic of endogenous OXT release depends on the nature of the stimulus—for example, social interaction or parental care—promoting this OXT release[71,72]. On the other hand, our neurochemical approached shows that the activation of OXT-R by perfusion of WAY267464 in SNpc induces a non-significant difference in DA_{ext} level in SNpc. This result is in line with the heterogeneous set of

responses showed by Xiao et al., [26]. As presented above, the effect of oxytocinergic activity on SNpc is topographically dependent, reporting that the dorsolateral region of the SNpc is where the highest density and collocation of OXT-R is found in GABAergic neurons, and that the release of OXT or activation of OXT-R in dorsolateral region induces a decrease in the FR of the DA neurons of the SNpc. This finding could explain why we did not find a significant change in the somatodendritic release of DA in the SNpc, because the agonist's perfusion was done in more medial regions, making the activation of OXT-R difficult.

Interestingly, when analyzing the axonal DA release post-perfusion of WAY267464 (5 μ M), we were able to identify a modest increase in DA_{ext} levels. This subtle dose-dependent increase could be due to compensatory changes made to maintain homeostasis of the nigro-striatal pathway for its optimal functioning of the motor control, against the decrease of the FR of the DA neurons coming from the SNpc, and this compensatory mechanism is supported by numerous studies involving animal with varying striatal depletion [73–75]. In line with this evidences, it has been reported that the decrease of the FR, associated with the increase of the GABAergic activity in the SNpc, specifically by the activation of GABA_A receptors in interneurons, can induce an increase of the axonal DA release in the DLS [76]. The SNpc perfusion of higher concentrations of WAY267464 (20-50 μ M, respectively) did not produce any significant changes in the DA_{ext} levels in the DLS as SNpc. These results could be related to the activation of the different variants of OXT-R (G_i), as it has been shown that the concentration necessary for the activation of the OXT-G_q receptor is much lower than for the OXT-G_i receptor [77,78]. Therefore, the use of high concentrations of the agonist could induce the activation of G_i-type receptors,

associated with a decrease in the neuronal excitability, generates a decrease in the probability of neurotransmitter release [79,80] but the expression of this variant in SNpc has not yet been shown. Another mechanism that could be participating is the desensitization and internalization of OXT-R, a mechanism that prevents neuronal hyperstimulation [81]. The repeated acute administration of OXT or agonist of OXT-R induces the rapid and marked desensitization of neuronal response in different brain areas [82,83].

In summary, we reported previously that the individual differences in impulsive choice behavior are associated with a change in the basal DA dynamic in the nigro-striatal pathway. The present research shows that bilateral activation of OXT-R in SNpc generates a differential effect that depends on the basal impulsive choice level; specifically, the activation of OXT-R in SNpc affects highly impulsive animals and induces a decrease in impulsive choice behavior. Interestingly, the activation of OXT-R in SNpc induces a moderate increase in DA_{ext} levels in the DLS, it is necessary to note that the neurochemical experiments did not differentiate between HI or LI animals. Regarding the effect of OXT-R activation on reducing impulsive behavior, further research is necessary to elucidate the role of the oxytocinergic transmission in the nigro-striatal pathway on DA levels during impulsive choice and the mechanisms underlying the low discounting rate of delayed reward.

5.6 Acknowledgments

This work was supported by FONDECYT, Fondo Nacional de Desarrollo Científico y Tecnológico, Chile. ID (1141088; J.A.F.); ANID-PCHA (21150508; M.M).

The authors have no conflict of interest.

5.7 References

- [1] A.J. Rosenfeld, J.A. Lieberman, L.F. Jarskog, Oxytocin, dopamine, and the amygdala: a neurofunctional model of social cognitive deficits in schizophrenia., *Schizophr. Bull.* 37 (2011) 1077–87. <https://doi.org/10.1093/schbul/sbq015>.
- [2] T.A. Baskerville, A.J. Douglas, Dopamine and Oxytocin Interactions Underlying Behaviors: Potential Contributions to Behavioral Disorders, *CNS Neurosci. Ther.* 16 (2010) e92–e123. <https://doi.org/10.1111/j.1755-5949.2010.00154.x>.
- [3] Z. Sarnyai, G.L. Kovács, Oxytocin in learning and addiction: From early discoveries to the present, *Pharmacol. Biochem. Behav.* (2014). <https://doi.org/10.1016/j.pbb.2013.11.019>.
- [4] O. Levi-Shachar, H.Z. Gvirts, Y. Goldwin, Y. Bloch, S. Shamay-Tsoory, O. Zagoory-Sharon, R. Feldman, H. Maoz, The effect of methylphenidate on social cognition and oxytocin in children with attention deficit hyperactivity disorder, *Neuropsychopharmacology*. 45 (2020) 367–373. <https://doi.org/10.1038/s41386-019-0522-5>.
- [5] K.R. Hamilton, M.R. Mitchell, V.C. Wing, I.M. Balodis, W.K. Bickel, M. Fillmore, S.D. Lane, C.W. Lejuez, A.K. Littlefield, M. Luijten, C.W. Mathias, S.H. Mitchell, T.C. Napier, B. Reynolds, C.G. Schütz, B. Setlow, K.J. Sher, A.C. Swann, S.E. Tedford, M.J. White, C.A. Winstanley, R. Yi, M.N. Potenza, F.G. Moeller, Choice impulsivity: Definitions, measurement issues, and clinical implications., *Personal. Disord.* 6 (2015) 182–98. <https://doi.org/10.1037/per0000099>.

- [6] B. Setlow, I.A. Mendez, M.R. Mitchell, N.W. Simon, Effects of chronic administration of drugs of abuse on impulsive choice (delay discounting) in animal models, *Behav. Pharmacol.* 20 (2009) 380–389. <https://doi.org/10.1097/FBP.0b013e3283305eb4>.
- [7] J.M. Berg, R.D. Latzman, N.G. Bliwise, S.O. Lilienfeld, Parsing the heterogeneity of impulsivity: A meta-analytic review of the behavioral implications of the UPPS for psychopathology, *Psychol. Assess.* 27 (2015) 1129–1146. <https://doi.org/10.1037/pas0000111>.
- [8] J. Vassileva, P.J. Conrod, Impulsivities and addictions: a multidimensional integrative framework informing assessment and interventions for substance use disorders, *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 374 (2019) 20180137. <https://doi.org/10.1098/rstb.2018.0137>.
- [9] M.M. Van Gaalen, R. van Koten, A.N.M. Schoffelman, L.J.M.J. Vanderschuren, Critical Involvement of Dopaminergic Neurotransmission in Impulsive Decision Making, *Biol. Psychiatry.* 60 (2006) 66–73. <https://doi.org/10.1016/j.biopsych.2005.06.005>.
- [10] J. Joutsa, V. Voon, J. Johansson, S. Niemelä, J. Bergman, V. Kaasinen, Dopaminergic function and intertemporal choice, *Transl. Psychiatry.* 5 (2015) e491. <https://doi.org/10.1038/tp.2014.133>.
- [11] J.W. Buckholtz, M.T. Treadway, R.L. Cowan, N.D. Woodward, R. Li, M.S. Ansari, R.M. Baldwin, A.N. Schwartzman, E.S. Shelby, C.E. Smith, R.M. Kessler, D.H. Zald, Dopaminergic network differences in human impulsivity, *Science* (80-.). 329

- (2010) 532. <https://doi.org/10.1126/science.1185778>.
- [12] W.S. Hyatt, M.D. Berquist, N.M. Chitre, L.N. Russell, K.C. Rice, K.S. Murnane, W.E. Fantegrossi, Repeated administration of synthetic cathinone 3,4-methylenedioxypyrovalerone persistently increases impulsive choice in rats, *Behav. Pharmacol.* 30 (2019) 555–565. <https://doi.org/10.1097/FBP.0000000000000492>.
- [13] J. Yacubian, T. Sommer, K. Schroeder, J. Gläscher, R. Kalisch, B. Leuenberger, D.F. Braus, C. Büchel, Gene-gene interaction associated with neural reward sensitivity, *Proc. Natl. Acad. Sci. U. S. A.* 104 (2007) 8125–8130. <https://doi.org/10.1073/pnas.0702029104>.
- [14] M. Leyton, Conditioned and sensitized responses to stimulant drugs in humans, *Prog. Neuro-Psychopharmacology Biol. Psychiatry.* 31 (2007) 1601–1613. <https://doi.org/10.1016/j.pnpbp.2007.08.027>.
- [15] J. Petzold, Y. Lee, S. Poosch, L. Oehme, B. Beuthien-Baumann, E.D. London, T. Goschke, M.N. Smolka, Presynaptic dopamine function measured with [18F]fluorodopa and L-DOPA effects on impulsive choice, *Sci. Rep.* 9 (2019). <https://doi.org/10.1038/s41598-019-54329-1>.
- [16] P. Trifilieff, D. Martinez, Imaging addiction: D2 receptors and dopamine signaling in the striatum as biomarkers for impulsivity, *Neuropharmacology.* 76 (2014) 498–509. <https://doi.org/10.1016/J.NEUROPHARM.2013.06.031>.
- [17] M. Bendix, K. Uvnäs-Moberg, M. Petersson, P. Gustavsson, P. Svanborg, M.

- Åsberg, J. Jokinen, Plasma oxytocin and personality traits in psychiatric outpatients., *Psychoneuroendocrinology*. 57 (2015) 102–10. <https://doi.org/10.1016/j.psyneuen.2015.04.003>.
- [18] A. Bozorgmehr, R. Moayedi, B. Sadeghi, M. Ghadirivasfi, M.T. Joghataei, A. Shahbazi, A Novel Link between the Oxytocin Receptor Gene and Impulsivity, *Neuroscience*. 444 (2020) 196–208. <https://doi.org/10.1016/j.neuroscience.2020.07.033>.
- [19] M.V. Sofroniew, Morphology of Vasopressin and Oxytocin Neurones and Their Central and Vascular Projections, *Prog. Brain Res.* 60 (1983) 101–114. [https://doi.org/10.1016/S0079-6123\(08\)64378-2](https://doi.org/10.1016/S0079-6123(08)64378-2).
- [20] L.W. Swanson, P.E. Sawchenko, Hypothalamic Integration: Organization of the Paraventricular and Supraoptic Nuclei, *Annu. Rev. Neurosci.* 6 (1983) 269–324. <https://doi.org/10.1146/annurev.ne.06.030183.001413>.
- [21] M. Mitre, J. Minder, E.X. Morina, M. V. Chao, R.C. Froemke, Oxytocin modulation of neural circuits, in: *Curr. Top. Behav. Neurosci.*, Springer Verlag, 2018: pp. 31–53. https://doi.org/10.1007/7854_2017_7.
- [22] M.J. Freund-mercier, F. Moos, D.A. Poulain, P. Richard, F. Rodriguez, D.T. Theodosis, J. didier Vincent, Role of central oxytocin in the control of the milk ejection reflex, *Brain Res. Bull.* 20 (1988) 737–741. [https://doi.org/10.1016/0361-9230\(88\)90085-8](https://doi.org/10.1016/0361-9230(88)90085-8).
- [23] G. Gimpl, F. Fahrenholz, The oxytocin receptor system: structure, function, and

- regulation., *Physiol. Rev.* 81 (2001) 629–683.
- [24] H.S. Knobloch, A. Charlet, L.C. Hoffmann, M. Eliava, S. Khrulev, A.H. Cetin, P. Osten, M.K. Schwarz, P.H. Seeburg, R. Stoop, V. Grinevich, Evoked Axonal Oxytocin Release in the Central Amygdala Attenuates Fear Response, *Neuron*. 73 (2012) 553–566. <https://doi.org/10.1016/j.neuron.2011.11.030>.
- [25] N. Lahoud, M. Maroun, Oxytocinergic manipulations in corticolimbic circuit differentially affect fear acquisition and extinction., *Psychoneuroendocrinology*. 38 (2013) 2184–95. <https://doi.org/10.1016/j.psyneuen.2013.04.006>.
- [26] L. Xiao, M.F. Priest, J. Nasenbeny, T. Lu, Y. Kozorovitskiy, Biased Oxytocinergic Modulation of Midbrain Dopamine Systems, *Neuron*. 95 (2017) 368–384.e5. <https://doi.org/10.1016/j.neuron.2017.06.003>.
- [27] R. Menon, T. Grund, I. Zoicas, F. Althammer, D. Fiedler, V. Biermeier, O.J. Bosch, Y. Hiraoka, K. Nishimori, M. Eliava, V. Grinevich, I.D. Neumann, Oxytocin Signaling in the Lateral Septum Prevents Social Fear during Lactation, *Curr. Biol.* 28 (2018) 1066–1078.e6. <https://doi.org/10.1016/j.cub.2018.02.044>.
- [28] T.R. Insel, The Challenge of Translation in Social Neuroscience: A Review of Oxytocin, Vasopressin, and Affiliative Behavior, *Neuron*. 65 (2010) 768–779. <https://doi.org/10.1016/j.neuron.2010.03.005>.
- [29] A. Meyer-Lindenberg, G. Domes, P. Kirsch, M. Heinrichs, Oxytocin and vasopressin in the human brain: social neuropeptides for translational medicine, *Nat. Rev. Neurosci.* 12 (2011) 524–538. <https://doi.org/10.1038/nrn3044>.

- [30] H.-J. Lee, A.H. Macbeth, J. Pagani, W.S. Young, Oxytocin: The Great Facilitator of Life, *Prog. Neurobiol.* 88 (2009) 127–51. <https://doi.org/10.1016/j.pneurobio.2009.04.001>.
- [31] L. Strathearn, Maternal neglect: Oxytocin, dopamine and the neurobiology of attachment, *J. Neuroendocrinol.* 23 (2011) 1054–1065. <https://doi.org/10.1111/j.1365-2826.2011.02228.x>.
- [32] H.M. Jin, S. Shrestha Muna, T.R. Bagalkot, Y. Cui, B.K. Yadav, Y.C. Chung, The effects of social defeat on behavior and dopaminergic markers in mice, *Neuroscience.* 288 (2015) 167–177. <https://doi.org/10.1016/j.neuroscience.2014.12.043>.
- [33] G. Blázquez, A. Castañé, A. Saavedra, M. Masana, J. Alberch, E. Pérez-Navarro, Social Memory and Social Patterns Alterations in the Absence of STriatal-Enriched Protein Tyrosine Phosphatase, *Front. Behav. Neurosci.* 12 (2019) 317. <https://doi.org/10.3389/fnbeh.2018.00317>.
- [34] K.T. Beier, E.E. Steinberg, K.E. Deloach, S. Xie, K. Miyamichi, L. Schwarz, X.J. Gao, E.J. Kremer, R.C. Malenka, L. Luo, Circuit Architecture of VTA Dopamine Neurons Revealed by Systematic Input-Output Mapping, *Cell.* 162 (2015) 622–634. <https://doi.org/10.1016/j.cell.2015.07.015>.
- [35] L. Angioni, C. Cocco, G.-L. Ferri, A. Argiolas, M.R. Melis, F. Sanna, Involvement of nigral oxytocin in locomotor activity: A behavioral, immunohistochemical and lesion study in male rats, (2016). <https://doi.org/10.1016/j.yhbeh.2016.05.012>.

- [36] L.W. Hung, S. Neuner, J.S. Polepalli, K.T. Beier, M. Wright, J.J. Walsh, E.M. Lewis, L. Luo, K. Deisseroth, G. Dölen, R.C. Malenka, Gating of social reward by oxytocin in the ventral tegmental area, 2017. <https://doi.org/10.1126/science.aan4994>.
- [37] W. Romero-Fernandez, D.O. Borroto-Escuela, L.F. Agnati, K. Fuxe, Evidence for the existence of dopamine d2-oxytocin receptor heteromers in the ventral and dorsal striatum with facilitatory receptor-receptor interactions, *Mol. Psychiatry*. (2013). <https://doi.org/10.1038/mp.2012.103>.
- [38] K. Fuxe, D.O. Borroto-Escuela, W. Romero-Fernandez, F. Ciruela, P. Manger, G. Leo, Z. Díaz-Cabiale, L.F. Agnati, On the role of volume transmission and receptor-receptor interactions in social behaviour: Focus on central catecholamine and oxytocin neurons, *Brain Res.* 1476 (2012) 119–131. <https://doi.org/10.1016/j.brainres.2012.01.062>.
- [39] F. Moos, P. Richard, Excitatory effect of dopamine on oxytocin and vasopressin reflex releases in the rat, *Brain Res.* 241 (1982) 249–260. [https://doi.org/10.1016/0006-8993\(82\)91061-7](https://doi.org/10.1016/0006-8993(82)91061-7).
- [40] D.K. Shahrokh, T.-Y. Zhang, J. Diorio, A. Gratton, M.J. Meaney, Oxytocin-dopamine interactions mediate variations in maternal behavior in the rat., *Endocrinology*. 151 (2010) 2276–86. <https://doi.org/10.1210/en.2009-1271>.
- [41] T.M. Love, Oxytocin, motivation and the role of dopamine, *Pharmacol. Biochem. Behav.* 119 (2014) 49–60. <https://doi.org/10.1016/j.pbb.2013.06.011>.

- [42] P.Y. Liao, Y.M. Chiu, J.H. Yu, S.K. Chen, Mapping Central Projection of Oxytocin Neurons in Unmated Mice Using Cre and Alkaline Phosphatase Reporter, *Front. Neuroanat.* 14 (2020). <https://doi.org/10.3389/fnana.2020.559402>.
- [43] S.X. Luo, E.J. Huang, Dopaminergic neurons and brain reward pathways: From neurogenesis to circuit assembly, *Am. J. Pathol.* 186 (2016) 478–488. <https://doi.org/10.1016/j.ajpath.2015.09.023>.
- [44] P.A. Eubig, T.E. Noe, S.B. Floresco, J.J. Sable, S.L. Schantz, Sex differences in response to amphetamine in adult Long–Evans rats performing a delay-discounting task, *Pharmacol. Biochem. Behav.* 118 (2014) 1–9. <https://doi.org/10.1016/j.pbb.2013.12.021>.
- [45] G. Paxinos, C. Watson, *The rat brain in stereotaxic coordinates*, Elsevier, 2009.
- [46] C.A. Winstanley, Y. Chudasama, J.W. Dalley, D.E.H. Theobald, J.C. Glennon, T.W. Robbins, Intra-prefrontal 8-OH-DPAT and M100907 improve visuospatial attention and decrease impulsivity on the five-choice serial reaction time task in rats., *Psychopharmacology (Berl)*. 167 (2003) 304–14. <https://doi.org/10.1007/s00213-003-1398-x>.
- [47] D. Viviani, A. Charlet, E. van den Burg, C. Robinet, N. Hurni, M. Abatis, F. Magara, R. Stoop, Oxytocin Selectively Gates Fear Responses Through Distinct Outputs from the Central Amygdala, *Science (80-.)*. 333 (2011) 104–107. <https://doi.org/10.1126/science.1201043>.
- [48] J. Myerson, L. Green, M. Warusawitharana, Area under the curve as a measure of

- discounting., J. Exp. Anal. Behav. 76 (2001) 235–43.
<https://doi.org/10.1901/jeab.2001.76-235>.
- [49] J.M. Slezak, K.G. Anderson, Effects of variable training, signaled and unsignaled delays, and d-amphetamine on delay-discounting functions, *Behav. Pharmacol.* 20 (2009) 424–436. <https://doi.org/10.1097/FBP.0b013e3283305ef9>.
- [50] A.M. Borges, J. Kuang, H. Milhorn, R. Yi, An alternative approach to calculating Area-Under-the-Curve (AUC) in delay discounting research, *J. Exp. Anal. Behav.* 106 (2016) 145–155. <https://doi.org/10.1002/jeab.219>.
- [51] F. Faul, E. Erdfelder, A.G. Lang, A. Buchner, G*Power 3: A flexible statistical power analysis program for the social, behavioral, and biomedical sciences, in: *Behav. Res. Methods*, Psychonomic Society Inc., 2007: pp. 175–191. <https://doi.org/10.3758/BF03193146>.
- [52] S. Mayr, E. Erdfelder, A. Buchner, F. Faul, A short tutorial of GPower, *Tutor. Quant. Methods Psychol.* 3 (2007) 51–59. <https://doi.org/10.20982/tqmp.03.2.p051>.
- [53] C. Hicks, W. Jorgensen, C. Brown, J. Fardell, J. Koehbach, C.W. Gruber, M. Kassiou, G.E. Hunt, I.S. McGregor, The Nonpeptide Oxytocin Receptor Agonist WAY 267,464: Receptor-Binding Profile, Prosocial Effects and Distribution of c-Fos Expression in Adolescent Rats, *J. Neuroendocrinol.* 24 (2012) 1012–1029. <https://doi.org/10.1111/j.1365-2826.2012.02311.x>.
- [54] R.H. Ring, L.E. Schechter, S.K. Leonard, J.M. Dwyer, B.J. Platt, R. Graf, S.

- Grauer, C. Pulicicchio, L. Resnick, Z. Rahman, S.J. Sukoff Rizzo, B. Luo, C.E. Beyer, S.F. Logue, K.L. Marquis, Z.A. Hughes, S. Rosenzweig-Lipson, Receptor and behavioral pharmacology of WAY-267464, a non-peptide oxytocin receptor agonist., *Neuropharmacology*. 58 (2010) 69–77. <https://doi.org/10.1016/j.neuropharm.2009.07.016>.
- [55] C.E. King, W.C. Griffin, L.N. Luderman, M.M. Kates, J.F. McGinty, H.C. Becker, Oxytocin Reduces Ethanol Self-Administration in Mice., *Alcohol. Clin. Exp. Res.* 41 (2017) 955–964. <https://doi.org/10.1111/acer.13359>.
- [56] H.J. Motulsky, R.E. Brown, Detecting outliers when fitting data with nonlinear regression – a new method based on robust nonlinear regression and the false discovery rate, *BMC Bioinformatics*. 7 (2006) 123. <https://doi.org/10.1186/1471-2105-7-123>.
- [57] K.A. Bernosky-Smith, Y.Y. Qiu, M. Feja, Y.B. Lee, B. Loughlin, J.X. Li, C.E. Bass, Ventral tegmental area D2 receptor knockdown enhances choice impulsivity in a delay-discounting task in rats, *Behav. Brain Res.* 341 (2018) 129–134. <https://doi.org/10.1016/j.bbr.2017.12.029>.
- [58] M.E. Ballard, A.C. Dean, M.A. Mandelkern, E.D. London, Striatal dopamine d2/d3 receptor availability is associated with executive function in healthy controls but not methamphetamine users, *PLoS One*. 10 (2015). <https://doi.org/10.1371/journal.pone.0143510>.
- [59] A. Mayer, N. Limberger, K. Starke, Transmitter release patterns of noradrenergic, dopaminergic and cholinergic axons in rabbit brain slices during short pulse trains,

- and the operation of presynaptic autoreceptors, *Naunyn. Schmiedeberg's. Arch. Pharmacol.* 338 (1988) 632–643. <https://doi.org/10.1007/BF00165627>.
- [60] C.J. Lobb, C.J. Wilson, C.A. Paladini, A dynamic role for GABA receptors on the firing pattern of midbrain dopaminergic neurons, *J. Neurophysiol.* 104 (2010) 403–413. <https://doi.org/10.1152/jn.00204.2010>.
- [61] P.L. Wood, Actions of GABAergic agents on dopamine metabolism in the nigrostriatal pathway of the rat., *J. Pharmacol. Exp. Ther.* 222 (1982).
- [62] J.L. Waddington, GABAergic mechanisms in the substantia nigra, *Nature.* 283 (1980) 696–697. <https://doi.org/10.1038/283696a0>.
- [63] S. Stagkourakis, H. Kim, D.J. Lyons, C. Broberger, Dopamine Autoreceptor Regulation of a Hypothalamic Dopaminergic Network, *Cell Rep.* 15 (2016) 735–747. <https://doi.org/10.1016/j.celrep.2016.03.062>.
- [64] S.J. Cragg, S.A. Greenfield, Differential autoreceptor control of somatodendritic and axon terminal dopamine release in substantia nigra, ventral tegmental area, and striatum., *J. Neurosci.* 17 (1997) 5738–46.
- [65] M.P. de la Mora, D. Pérez-Carrera, M. Crespo-Ramírez, A. Tarakanov, K. Fuxe, D.O. Borroto-Escuela, Signaling in dopamine D2 receptor-oxytocin receptor heterocomplexes and its relevance for the anxiolytic effects of dopamine and oxytocin interactions in the amygdala of the rat, *Biochim. Biophys. Acta - Mol. Basis Dis.* 1862 (2016) 2075–2085. <https://doi.org/10.1016/j.bbadis.2016.07.004>.
- [66] N.D. Volkow, R.D. Baler, NOW vs LATER brain circuits: Implications for obesity

- and addiction, *Trends Neurosci.* 38 (2015) 345–352.
<https://doi.org/10.1016/j.tins.2015.04.002>.
- [67] R.M. Bilder, J. Volavka, H.M. Lachman, A.A. Grace, The catechol-O-methyltransferase polymorphism: Relations to the tonic-phasic dopamine hypothesis and neuropsychiatric phenotypes, *Neuropsychopharmacology*. 29 (2004) 1943–1961. <https://doi.org/10.1038/sj.npp.1300542>.
- [68] J.R. St. Onge, S.B. Floresco, Prefrontal Cortical Contribution to Risk-Based Decision Making, *Cereb. Cortex*. 20 (2010) 1816–1828.
<https://doi.org/10.1093/cercor/bhp250>.
- [69] J. Fam, F. Westbrook, E. Arabzadeh, Behavioral correlates of the decision process in a dynamic environment: post-choice latencies reflect relative value and choice evaluation., *Front. Behav. Neurosci.* 9 (2015) 261.
<https://doi.org/10.3389/fnbeh.2015.00261>.
- [70] M. Watabe-Uchida, L. Zhu, S.K. Ogawa, A. Vamanrao, N. Uchida, Whole-Brain Mapping of Direct Inputs to Midbrain Dopamine Neurons, *Neuron*. 74 (2012) 858–873. <https://doi.org/10.1016/j.neuron.2012.03.017>.
- [71] C.S. Carter, A.J. Grippio, H. Pournajafi-Nazarloo, M.G. Ruscio, S.W. Porges, Oxytocin, vasopressin and sociality, *Prog. Brain Res.* 170 (2008) 331–336.
[https://doi.org/10.1016/S0079-6123\(08\)00427-5](https://doi.org/10.1016/S0079-6123(08)00427-5).
- [72] E.B. Keverne, K.M. Kendrick, Oxytocin Facilitation of Maternal Behavior in Sheep, *Ann. N. Y. Acad. Sci.* 652 (1992) 83–101. <https://doi.org/10.1111/j.1749->

6632.1992.tb34348.x.

- [73] P.A. Garris, Q.D. Walker, R.M. Wightman, Dopamine release and uptake rates both decrease in the partially denervated striatum in proportion to the loss of dopamine terminals, *Brain Res.* 753 (1997) 225–234. [https://doi.org/10.1016/S0006-8993\(97\)00003-6](https://doi.org/10.1016/S0006-8993(97)00003-6).
- [74] B.P. Bergstrom, P.A. Garris, “Passive stabilization” of striatal extracellular dopamine across the lesion spectrum encompassing the presymptomatic phase of Parkinson’s disease: A voltammetric study in the 6-OHDA-lesioned rat, *J. Neurochem.* 87 (2003) 1224–1236. <https://doi.org/10.1046/j.1471-4159.2003.02104.x>.
- [75] X.A. Perez, N. Parameswaran, L.Z. Huang, K.T. O’Leary, M. Quik, Pre-synaptic dopaminergic compensation after moderate nigrostriatal damage in non-human primates, *J. Neurochem.* 105 (2008) 1861–1872. <https://doi.org/10.1111/j.1471-4159.2008.05268.x>.
- [76] N. Balon, B. Kriem, M. Weiss, J.C. Rostain, GABAA receptors in the pars compacta and GABAB receptors in the pars reticulata of rat substantia nigra modulate the striatal dopamine release, *Neurochem. Res.* 27 (2002) 373–379. <https://doi.org/10.1023/A:1015595729411>.
- [77] M. Busnelli, A. Saulière, M. Manning, M. Bouvier, C. Galés, B. Chini, Functional Selective Oxytocin-derived Agonists Discriminate between Individual G Protein Family Subtypes, *J. Biol. Chem.* 287 (2012) 3617–3629. <https://doi.org/10.1074/jbc.M111.277178>.

- [78] M. Busnelli, E. Bulgheroni, M. Manning, G. Kleinau, B. Chini, Selective and Potent Agonists and Antagonists for Investigating the Role of Mouse Oxytocin Receptors, *J. Pharmacol. Exp. Ther.* 346 (2013) 318–327. <https://doi.org/10.1124/jpet.113.202994>.
- [79] G. Milligan, E. Kostenis, Heterotrimeric G-proteins: a short history, *Br. J. Pharmacol.* 147 (2009) S46–S55. <https://doi.org/10.1038/sj.bjp.0706405>.
- [80] Y. Yudin, T. Rohacs, Inhibitory G_{i/o}-coupled receptors in somatosensory neurons: Potential therapeutic targets for novel analgesics, *Mol. Pain.* 14 (2018) 174480691876364. <https://doi.org/10.1177/1744806918763646>.
- [81] M.P. Smith, V.J. Ayad, S.J. Mundell, C.A. McArdle, E. Kelly, A. López Bernal, Internalization and desensitization of the oxytocin receptor is inhibited by dynamin and clathrin mutants in human embryonic kidney 293 cells, *Mol. Endocrinol.* 20 (2006) 379–388. <https://doi.org/10.1210/me.2005-0031>.
- [82] M.G. Terenzi, C.D. Ingram, Oxytocin-induced excitation of neurones in the rat central and medial amygdaloid nuclei, *Neuroscience.* 134 (2005) 345–354. <https://doi.org/10.1016/j.neuroscience.2005.04.004>.
- [83] M. Busnelli, B. Chini, Molecular basis of oxytocin receptor signalling in the brain: what we know and what we need to know, in: *Curr. Top. Behav. Neurosci.*, Springer Verlag, 2018: pp. 3–29. https://doi.org/10.1007/7854_2017_6.

1. Figures

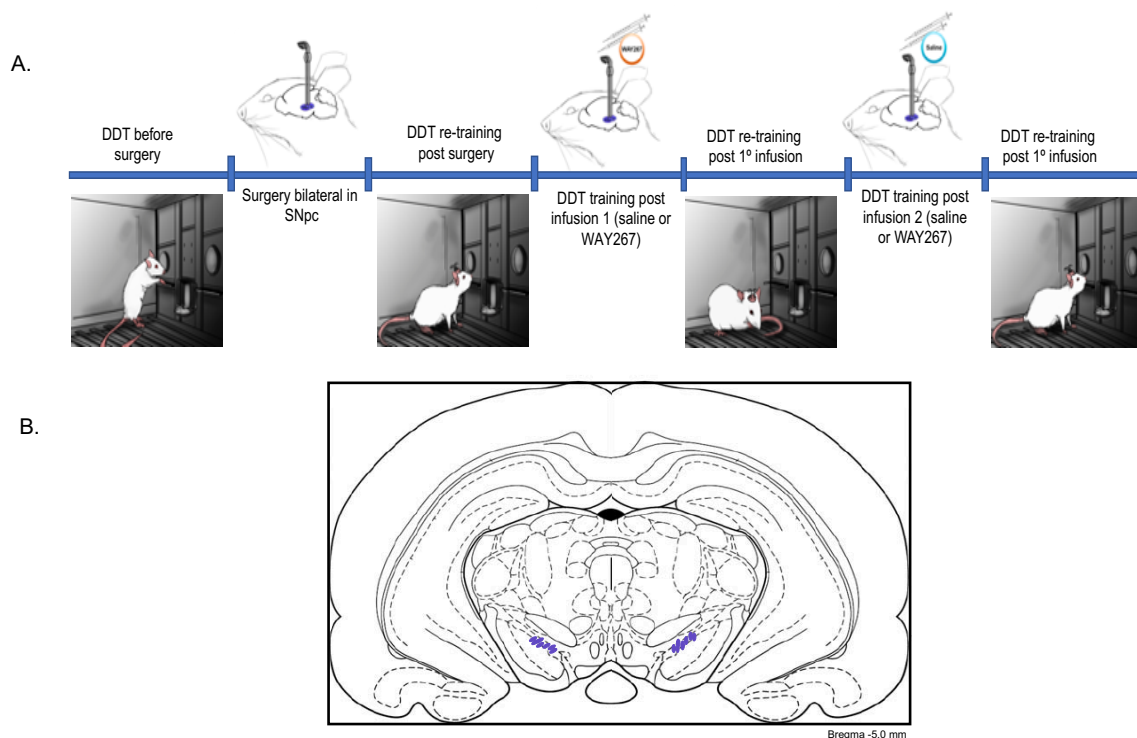


Figure 1: Experimental design of micro-infusion of WAY267 in SNpc and histology scheme. (A) Experimental design. (B) Histology. Schematic coronal sections of the rat brain showing the placements of the cannula for all rats in the experiment that evaluated the effect of OXT-R activation in SNpc on the behavior of impulsive choice. Rats received micro-infusion of OXT-R agonist (WAY267464) or vehicle (saline) into the SNpc. Brain sections correspond to the atlas of [44].

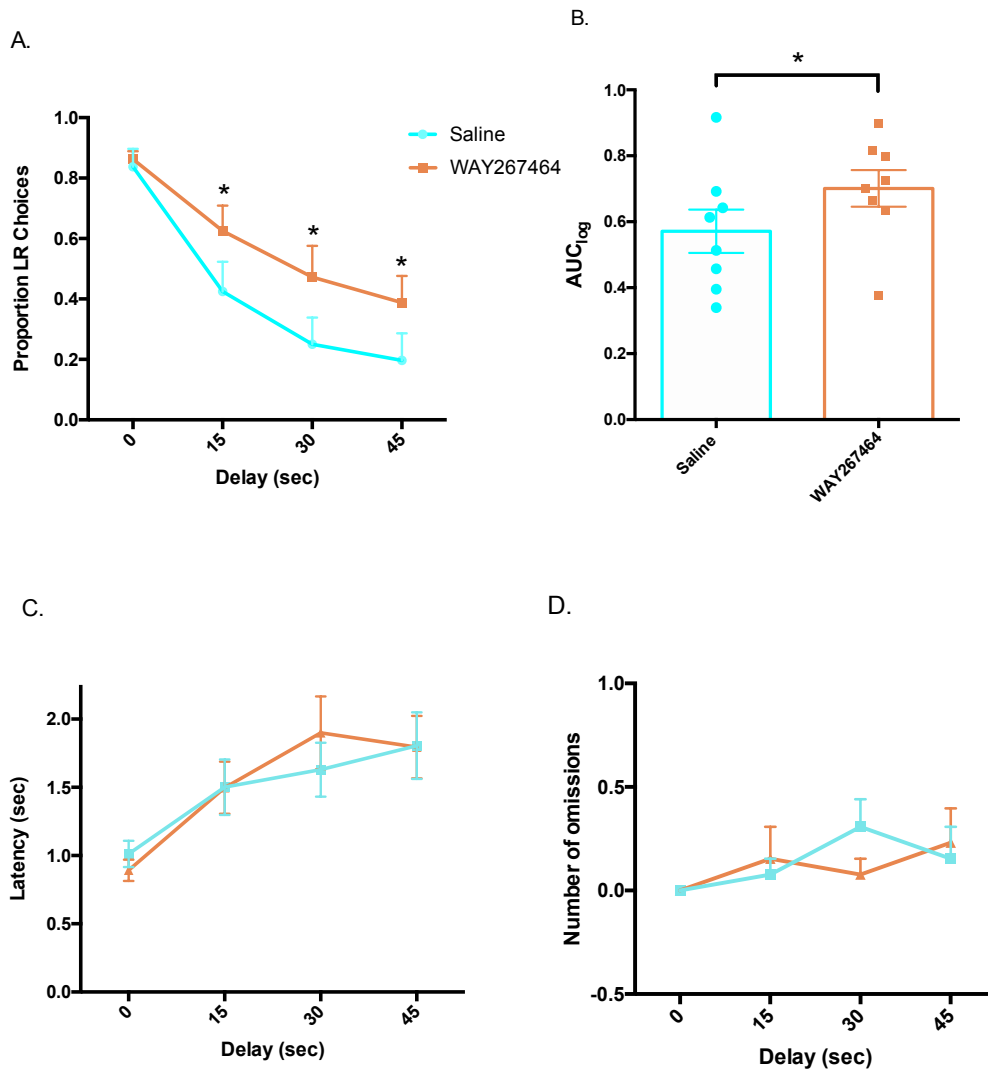


Figure 2: Effect of bilateral OXT-R activation in SNpc during DDT. (A) Delay-discounting curve of basal (saline microinfusion) condition and bilateral microinfusion of non-peptide agonists WAY267464 (3 μ g/0.5 μ l) in SNpc. Showed significant difference in the number of choices during the delays of 15, 30 and 45 seconds between saline and WAY267464 groups according to two-way ANOVA (n= 8; Bonferroni test: delay 15: Saline: 0.42 ± 0.07 vs WAY267464: 0.62 ± 0.07 p= 0.03; Delay 30: Saline: 0.25 ± 0.07 vs WAY267464: 0.47 ± 0.07 p= 0.01; Delay 45: Saline: 0.19 ± 0.07 vs WAY267464: 0.38 ± 0.07 p= 0.04). (B) The bilateral microinfusion of WAY267464 in the SNpc induced an increase in the area under the curve (AUC_{log}) pointing to a higher preference to the larger reinforcement (n= 8; t-test p= 0.02). (C) Graph of response latency during the DDT showed no differences between the two experimental conditions (n=8 t-test: p= 0.35). (D) Number of total omissions during free choice showed non-significant differences between the two experimental conditions (n=8; t-test; p= 0.41).

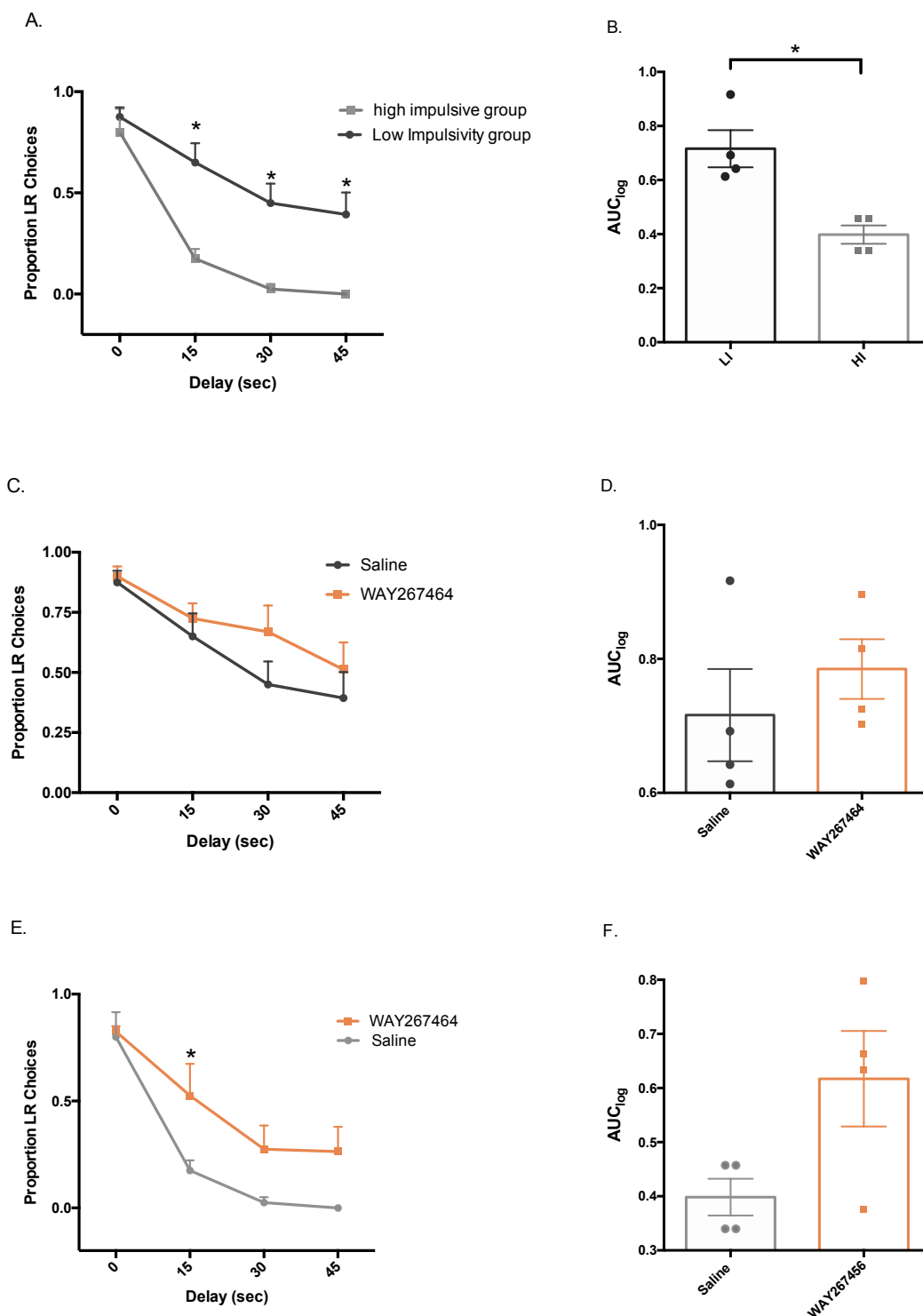


Figure 3: Differential effect of bilateral OXT-R activation in SNpc by microinjection of WAY267464 (3 ug/ul) and saline solution. (A) Delay discounting curve of HI and LI

animals as classified by the AUC_{log} criteria (LI=4; HI=4). Showed significant difference in the number of choices during the delays of 15, 30 and 45 seconds between HI and LI groups according to two-way ANOVA ($n= 4/\text{group}$; Bonferroni test: delay 15: LI: 0.65 ± 0.08 vs HI: 0.17 ± 0.08 $p= 0.001$; Delay 30: LI: 0.45 ± 0.08 vs HI: 0.02 ± 0.08 $p= 0.002$; Delay 45: LI: 0.39 ± 0.08 vs HI: 0.1 ± 0.08 $p= 0.003$). **(B)** HI animals showed an increase in the AUC_{log} pointing to a higher preference to the larger reinforcement compared to LI animals ($n= 4/\text{group}$; t-test $p= 0.01$). **(C)** Bilateral microinfusion of non-peptide agonists WAY267464 ($3\mu\text{g}/0.5\mu\text{l}$) in SNpc. For the LI group, there were non-significant differences in the number of large reinforcement choices during delays of 5, 30 and 45 seconds according to two-way ANOVA ($n= 4$; Bonferroni test: delay 15: Saline: 0.65 ± 0.09 vs WAY267464: 0.72 ± 0.09 $p> 0.99$; Delay 30: Saline: 0.45 ± 0.09 vs WAY267464: 0.66 ± 0.09 $p= 0.16$; Delay 45: Saline: 0.39 ± 0.09 vs WAY267464: 0.51 ± 0.09 $p= 0.91$). **(D)** The bilateral microinfusion of WAY267464 in the SNpc induced a non-significant difference in AUC_{log} pointing to a higher preference to the larger reinforcement ($n= 4$; t-test $p= 0.19$). **(E)** Bilateral microinfusion of non-peptide agonists WAY267464 ($3\mu\text{g}/0.5\mu\text{l}$) in SNpc. For the HI group, there were one significant difference in the number of large reinforcement choices during delay of 15sec and non-significant differences in 30 and 45 seconds according to two-way ANOVA ($n= 4$; Bonferroni test: delay 15: Saline: 0.17 ± 0.11 vs WAY267464: 0.52 ± 0.11 $p= 0.05$; Delay 30: Saline: 0.02 ± 0.11 vs WAY267464: 0.27 ± 0.11 $p= 0.21$; Delay 45: Saline: 0.0 ± 0.11 vs WAY267464: 0.26 ± 0.11 $p= 0.16$). **(F)** The bilateral microinfusion of WAY267464 in the SNpc induced a non-significant difference in AUC_{log} pointing to a higher preference to the larger reinforcement ($n= 4$; t-test $p= 0.05$).

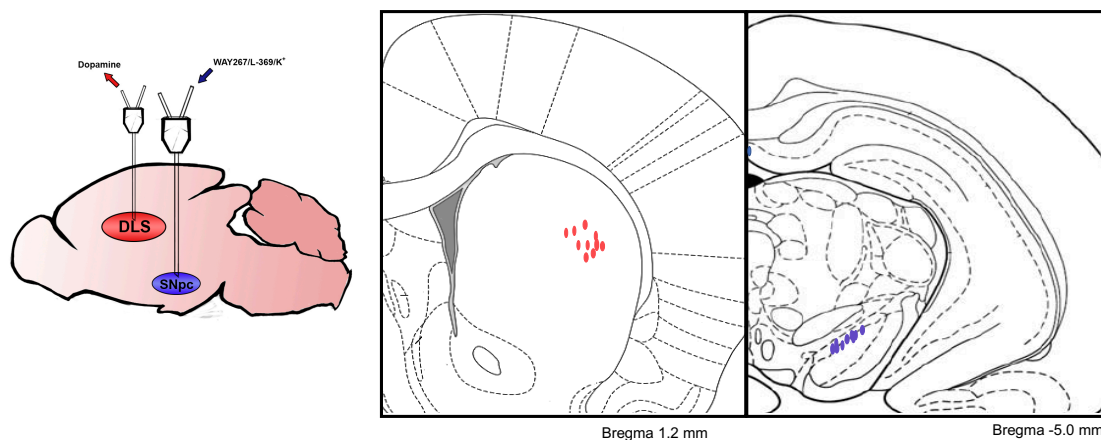


Figure 4: Representative scheme of probe locations during the microdialysis protocol. On the **Left**, the scheme shows two probes: one probe in SNpc in which different concentrations of WAY267464, K^+ solution and the OXT-R antagonist (LY369) were perfused. Another probe was inserted in the DLS to measure extracellular DA levels. On the **Right**: Histology. Schematic coronal sections of the rat brain showing the placements of the microdialysis cannula tips for all rats in the experiment analyzing the effect on DLS extracellular DA levels due to OXT-R activation in SNpc. Brain sections correspond to the atlas of Paxinos and Watson [45].

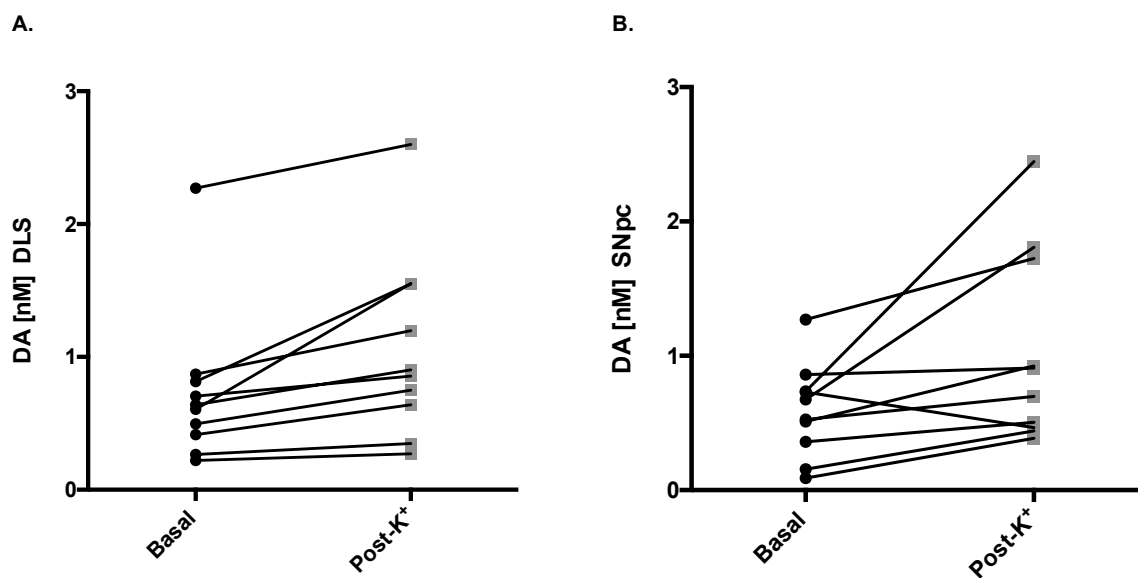


Figure 5: The effect of perfusing the SNpc with a high concentration of Potassium (70mM) on DA_{ext} in the nigro-striatal pathway. (A) and (B), Perfusion with depolarizing potassium solutions (70 mM) for 5 min in SNp, >20% of increase DA_{ext} level in the DLS and SNpc, respectively (according to paired t-test: DLS: $p=0.004$ and SNpc: $p=0.038$; $n=10$).

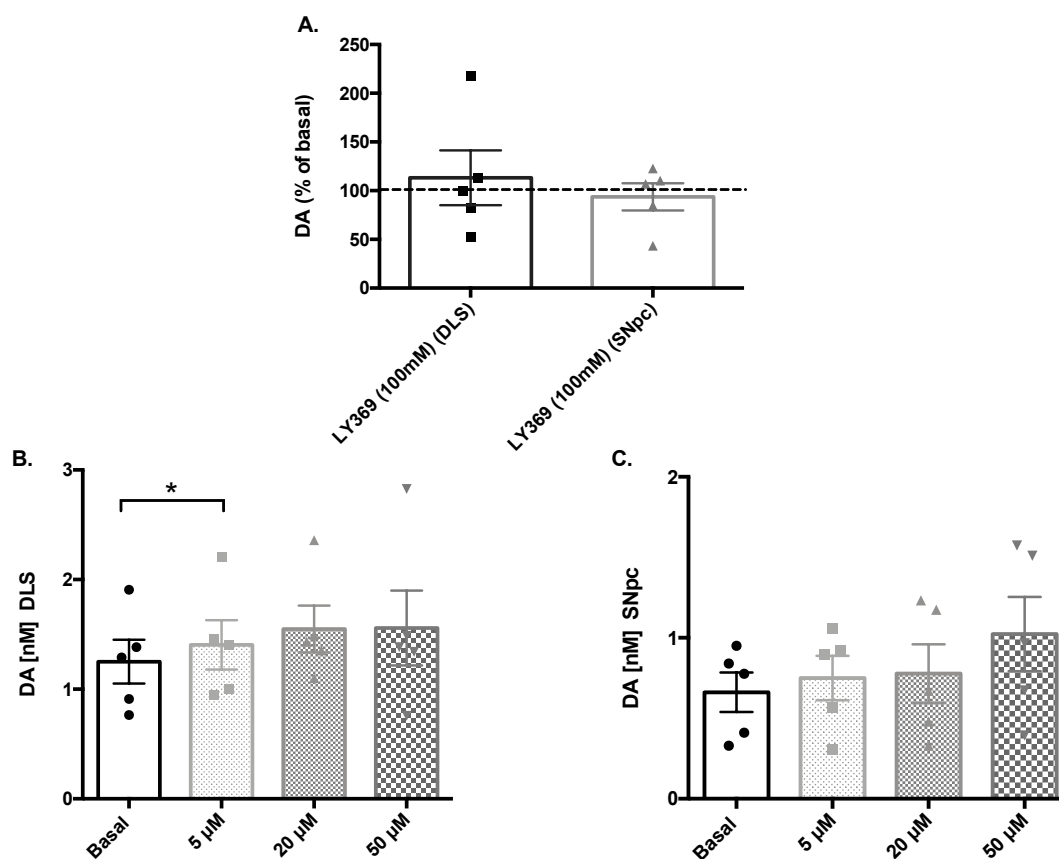


Figure 6: Effect of OXT-R activation in SNpc on DA_{ext} levels in DLS and SNpc. (A), showed that the perfusion of OXT-R antagonist LY369 (100mM) in SNpc, did not produce any change in DA_{ext} levels in DLS and SNpc (according to paired t-test: DLS, $p=0.64$; SNpc, $p=0.65$, $n=5$). (B) and (C) Perfusion of different concentrations of OXT-R agonist WAY267464 (5, 20, 50 mM) in the SNpc on DA_{ext} levels in DLS and SNpc, respectively. DA is expressed as nM concentration. (B) Results showed a moderated but significant increase in DLS DA_{ext} levels after infusion of WAY267464 (5 mM) (according to paired t-test; $p=0.05$; $n=5$). (C) results showed a non-significant change in DA_{ext} levels in SNpc after infusion of WAY267464 in the SNpc (according to paired t-test; 20 μM $p=0.42$; 50 μM $p=0.25$; $n=5$).

6 General Discussion

Evidence supports the notion that variability in dopamine (DA) neurotransmission function is a major contributor to individual differences observed in impulsive choice (Joutsa et al., 2015; Van Gaalen et al., 2006). Interestingly, various approaches have suggested that hypothalamic oxytocinergic projection modulates midbrain DA regions (Hung et al., 2017; Xiao et al., 2017), but the neurochemical DA dynamic and behavioral consequences of this potential interaction are not yet completely known. Here, we report that a higher DLS DA release was observed in HI rats than LI rats. Also, a significant negative correlation was observed between AUC and DA release. These results suggest that an increased DLS DA signaling contributes to delay-related decision-making. Subsequently, we showed that activation of the OXT-R in the SNpc induces a decreased impulsive choice in HI rats, displaying a greater preference for larger delayed rewards. Secondly, the OXT-R activation in SNpc induces a slight increase in DA_{ext} level in DLS but does not induce a change in DA_{ext} level in SNpc. This latter finding suggests that activation of the OXT-R in the SNpc influences delay-related decision making by subtle changes in the striatal dopamine transmission.

Our first general hypothesis was modified due to technical drawbacks that did not allow us to study one of the research's initial focuses associated with identifying oxytocinergic transmission variations. Specifically, studying changes in the expression of OXT-R in the SNpc present at impulsive choice behaviors; this could not be studied due to the low specificity of the existing antibodies for OXT-R used in the colorimetric immunohistochemistry (Yoshida et al., 2009). Therefore, labeling, identification, and quantification under these conditions made it impossible to obtain a reliable result. For

this reason, we reformulated the general hypothesis to fully evaluate it by the different experimental designs performed in this doctoral research. The new general hypothesis was formulated based on the finding by Tedford et al., (2015), where they associated high levels of impulsive choice with dopaminergic hypofunction. Moreover, evidence has showed that DA and OXT interaction (Hung et al., 2017; Xiao et al., 2017). Clinical studies showed a significant association between basal plasma OXT and the Karolinska Scale of Personality (KSP) factor impulsiveness, suggesting a positive correlation between impulsiveness and plasma OXT level in male psychiatric outpatients (Bendix et al., 2015).

The new general hypothesis was partially corroborated by the results obtained in chapters 1 and 2, showing that high DA release in DLS is associated with higher levels of impulsive choice and that increase in the oxytocinergic activity in the nigro-striatal pathway induce a decrease in the impulsive choice. In order to corroborate the specific hypothesis stated in chapter 1, our objective was to evaluate the relationship between delay discounting behavior and DA dynamics in the DLS in rats, but the hypothesis was not true; our results showed that a higher DLS DA release was observed in HI rats compared with LI rats. A significant negative correlation was observed between AUC and DA release, indicating that lower AUC is associated with higher DLS DA release. Notwithstanding the limitations of our study, these results suggest that an increased DLS DA signaling contributes to delay-related decision-making.

The increased DA release observed in rats classified as HI could be explained by a presynaptic mechanism associated with a loss of the inhibitory tonic control of D₂

receptors on DA extracellular concentration (Mayer et al., 1988); it has been proposed that individual differences in D₂ receptor mRNA expression predict different levels of impulsive choice (Simon et al., 2013). Furthermore, it could be explained by the postsynaptic mechanism associated with an increase in population activity or bursting activity of DA neurons in the SNpc that contribute to HI animals (Martinez et al., 2019). Our results showed that DA neurotransmission in DLS contributes to individual differences in impulsive choice. The increased DLS DA release observed in HI rats could maintain activated postsynaptic DA receptors in the dorsal striatum responsible for increasing delay aversion (Martinez et al., 2019).

On the other hand, the specific hypothesis studied in chapter 2 was fully corroborated. For this purpose, we evaluated the effect of the bilateral activation of OXT-R in SNpc in impulsive choice behavior. Our finding showed that bilateral microinfusion of WAY267464 induces a decrease in impulsivity. Due to the individual differences present in impulsive choice, this finding showed that in animals classified as HI, the bilateral activation of OXT-R in SNpc decreases impulsive behaviors compared to the LI group. Xiao et al. (2017), showed that the OXT-R activation in SNpc induces a heterogeneous set of responses, dependently on topographic location into SNpc; the dorsolateral OXT-R activation induces a decrease in the activity of dorsolateral SNpc DA neurons via inhibitory circuits, but in the medial region of SNpc did not find a significant change in medial SNpc DA neurons. In this sense, the mechanism underlying the effect exerted by OXT-R activation on impulsive behavior could be linked to an increment in GABA signaling, which could be an indirect mechanism that provokes a hyperpolarization of membrane potential, causing a decrement in the DA FR in SNpc (Lobb et al., 2010; Xiao

et al., 2017), producing a decrease of DA levels in the DLS (Waddington, 1980; Wood, 1982). This modulation could regulate the behavior by altering the tonic activity of DA striatal tone, helping to recover neurochemical homeostasis linked to decreased impulsive choice behavior. Another alternative mechanism could be associated with improving the somatodendritic D₂-R affinity for DA. It has been shown that an increase in the concentration of ligands for OXT-R induces an increase in the affinity of D₂-R for DA (Cragg & Greenfield, 1997; de la Mora et al., 2016; Romero-Fernandez et al., 2013; Stagkourakis et al., 2016), which induced changes in the dopaminergic activity during the presence of a salience stimulus, facilitating advantageous decision making (Bilder et al., 2015).

Various research has attributed an essential role to striatal regions involved in the inhibitory control of cognitive operations, in the positive reward choice (Ghahremani et al., 2012; Robinson et al., 2009), and in the habits and routines (Tricomi et al., 2009). In line with these, neuroimaging studies have proven that in response to a stimulus experimented as gratifying, it correlates with an increase of fast DA tone in ventral and dorsal striatal areas (Volkow et al., 2014). Two types of dopaminergic activity have been identified: low-frequency tonic firing (2–10 Hz), which determines basal extracellular DA level, and phasic firing patterns (≥ 20 Hz), which produce transient large-amplitude increases in DA level (Floresco et al., 2003; Grace & Bunney, 1984; Siciliano et al., 2015). The phasic activity of DA neurons has been observed during the expectation of rewards (Apicella et al., 1992; Schultz et al., 2015). At the same time, tonic DA modulates performance without altering learning (Beeler et al., 2010). Grace. A, (1991) proposed

that phasic and tonic DA signaling has a separate but complementary role in maintaining the homeostasis of circuits associated with the reward system.

Considering this evidence, we propose the following mechanism: (1) The animals classified as HI (showed more preferences for SR or “NOW” reinforcement) presented higher DA release in the DLS, possibly mediated by presynaptic mechanism (e.g., decrease in the D₂-R activity) or postsynaptic mechanism (e.g., increase in population activity or bursting activity of DA neurons in the SNpc) which could contribute to HI behavior (Ford, 2014; Marcott et al., 2014; Rice & Patel, 2015; Sulzer et al., 2016). The increase of DA release in DLS levels is associated with an increase in the tonic DA tone in HI groups, which could alter the phasic DA activity during the attention of salient stimulus, promoting the choice of SR or “NOW” reinforcement (see supplementary figure 1.B). It has been shown that the hyperactivity of the DA system may attenuate the firing of phasic burst (Stopper et al., 2014) and that diminished tonic DA tone promote an increase in burst firing of DA neurons would result in overstimulation via phasic dopamine release (Floresco et al., 2003). Therefore, it has been proposed that tonic DA release regulates the intensity of the phasic DA response. In this way, the DA tonic release would establish the background level of stimulation of dopamine receptors (Grace. A, 1991). (2) The animals classified as LI (showed more preferences for LR or “LATER” reinforcement) present normal DA release and normal tonic DA tone in the DLS than the HI group (see figure supplementary 1.C). Thus, the expression as an activity mediated by the presynaptic D₂-R receptors is intact, allowing homeostasis between the tonic and phasic DA activity, thus generating an optimum balance in the behaviors associated with the LR or “LATER” preference.

From the results finding in chapter 2, we suggest the following mechanism: (1) There exists a difference between tonic and phasic dopaminergic activity linked to decision making: our results suggest that the HI animals present an imbalance between the tonic-phasic DA tone, showing an alteration in the phasic tone in the presence of salience of stimulus inducing a disadvantages choice or preferences for “NOW” reinforcement (see supplementary figure 1.D). (2) The bilateral activation by the non-peptide agonists of OXT-R microinfusion in the SNpc induces a decrease in the levels of impulsive choice during DDT. As previously discussed, it has been proposed that the increase in DA levels due to OXT-R in SNpc could be facilitating the interaction between presynaptic D₂-R and DA. This mechanism, could balance the homeostasis in phasic striatal dopamine tone, facilitating de LR choice. On the other hand, in animals previously classified as LI, the bilateral activation of OXT-R in SNpc does not significantly alter their behavior. This mechanism could be explained because these LI animals balance the phasic and tonic DA tone, facilitating their flexibility when choosing between LR or SR during the behavioral test (Hikida et al., 2010; Yawata et al., 2012).

Even though theses result differs from previous results (see chapter 1), it was found that high impulsive choice level is associated to high DA basal level in DLS and the OXT-R activation in SNpc decreases impulsive in HI animals. However, low concentration of WAY267464 in SNpc induce a subtle DA increase in DLS. Our neurochemical study did not differentiate HI from LI rats. Future studies are needed to study OXT-R activation, as it may have opposite effects on the nigro-striatal pathway depending on neurochemical basal conditions. The OXT-R have different isomorphs that become activated depending on the concentration of its ligand. This means that they present a higher or lower affinity

for the ligand, depending on their concentration (Busnelli et al., 2012). When analyzing the doses that were used in the infusion and microdialysis, differences can be observed in the concentrations used. This is mainly due to methodologic differences linked to the use of the microdialysis probes, which have a membrane that allows fluid exchange but also interferes in the percentage of the real drug concentration perfused during these protocols. The microinfusion uses a guide cannula into which a microinjector is introduced which doesn't have any kind of membrane at its end, generating an expedited flow of the solution with the desired concentration ensured. Also, it is important to remember the topographical effect of OXT-R in dorsolateral SNpc showed by Xiao et al., (2017).

The present research opens new possibilities for investigation and determination of different mechanisms linked to the individual differences present in impulsive behaviors. It would be fascinating to measure the levels of DA in the DLS during DDT and verify the dynamic changes in basal DA levels during decision making. On the other hand, research on the interaction between the OXT and DA systems with neurochemical and behavioral correlations generates several open questions for future research: (1) What are the dopaminergic neurochemical dynamics during DDT in different impulsive choice behaviors? (2) What is the mechanism through which the activation of OXT-R in SNpc induces changes in DA levels in the DLS? (3) Is the behavioral effect of the bilateral activation of OXT-R in SNpc dose-dependent and mediated only by OXT-R? Further investigations are needed to answer these questions and have the possibility to contribute to our understanding of mechanism associated to dopamine nigro-striatal pathway during impulsive choice behaviors.

7 General Conclusions

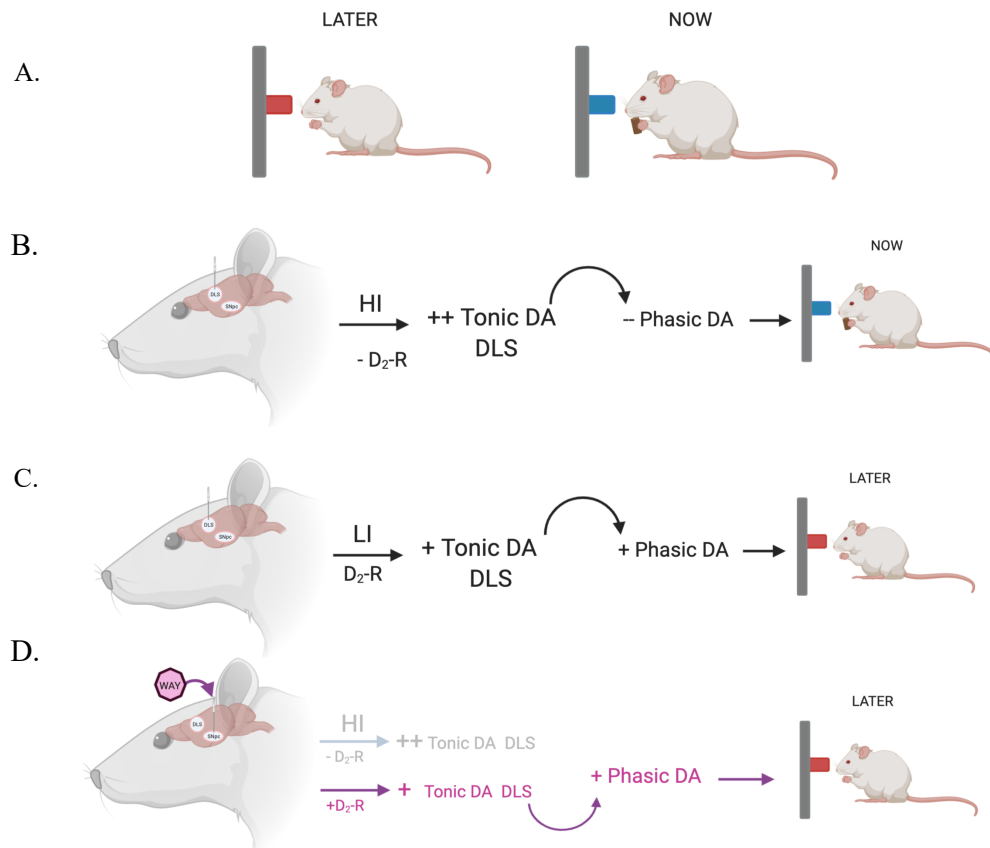
1. High levels of impulsive choice behavior are associated to an increase in the release DA in the DLS in anesthetized condition, possibly mediated by presynaptic mechanism.
2. Pharmacological and behavioral data show that bilateral activation of OXT-R in the SNpc by microinfusion of WAY267465 induces a decrease in impulsive choice in high impulsive animals.
3. Neurochemical data shows that the activation of OXT-R through the perfusion of WAY267464 (5 μ M) in anesthetized animals, induces a subtle increase in the DA levels in the DLS.
4. The perfusion of the antagonist OXT-R induced non-significant differences in the DA_{ext} level in the SNpc and DLS, which suggest that no endogenous OXT tone exists in the SNpc.

In summary, we reported that DA and OXT neurotransmission contributes to individual differences in impulsive choice. This finding suggests that HI animals have higher tonic levels of DA, so when a salient stimulus is presented, they release less phasic DA that induces a defective coding of a high-value reward. The activation of OXT-R in the SNpc stimulates the GABA signaling, this could be an indirect mechanism that was causing a decrement in the DA FR in SNpc, producing a decrease of DA levels in the DLS (Waddington, 1980; Wood, 1982). This modulation could regulate the behavior by

altering the tonic activity of DA striatal tone, helping to recover neurochemical homeostasis linked to impulsive behavior, facilitating the phasic activity against a salient stimulus, decreasing the impulsive behavior. Several questions remain to be answered. Future research should investigate the neurobiological mechanisms that underlie the dopaminergic and oxytocinergic systems modulation in the nigro-striatal pathway in impulsive choice behavior.

8 Supplementary information

8.1 Proposed Model



Supplementary figure 1: DA basal dynamic activity present in impulsive choice behavior and the effect of OXT-R activation in SNpc during DDT. (A) Different lever press in DDT, **Left**, lever press in red associated to large (LR) or “LATER” reinforcement. **Right**, lever press in blue associated to small (SR) or “NOW” reinforcement. (B) High impulsive rats have decreased presynaptic D₂-R activity and higher tonic DA levels in the DLS. Therefore, less phasic DA activity during a stimulus salience presentation, generating a wrong coding of the value reward (favoring SR or NOW choice). (C) Low impulsive animal has a normal presynaptic D₂-R activity and tonic DA level in DLS. Therefore, regular phasic DA activity during a stimulus salience presentation, generating a correct coding of the value reward (favoring LR or LATER choice). (D) Bilateral activation of OXT-R in the SNpc in HI animals, increase in presynaptic D₂-R affinity for DA and decrease of tonic DA activity, increased phasic DA activity during the presentation stimulus salience, decreasing impulsivity (favoring LATER behavior).

9 References

- Amorim Neto, R. D. C., & True, M. (2011). The development and treatment of impulsivity. *Psico*, 42(1), 134–141. <https://doi.org/10.1080/J006v03n02>
- Apicella, P., Scarnati, E., Ljungberg, T., & Schultz, W. (1992). Neuronal activity in monkey striatum related to the expectation of predictable environmental events. *Journal of Neurophysiology*, 68(3), 945–960. <https://doi.org/10.1152/jn.1992.68.3.945>
- Arce, E., & Santisteban, C. (2006). Impulsivity: a review. *Psicothema*, 18(2), 213–220.
- Aspé-Sánchez, M., Moreno, M., Rivera, M. I., Rossi, A., & Ewer, J. (2015). Oxytocin and Vasopressin Receptor Gene Polymorphisms: Role in Social and Psychiatric Traits. *Frontiers in Neuroscience*, 9, 510. <https://doi.org/10.3389/fnins.2015.00510>
- Avila, C., Cuenca, I., Félix, V., Parcet, M.-A., & Miranda, A. (2004). Measuring impulsivity in school-aged boys and examining its relationship with ADHD and ODD ratings. *Journal of Abnormal Child Psychology*, 32(3), 295–304.
- Baarendse, P. J. J., & Vanderschuren, L. J. M. J. (2012). Dissociable effects of monoamine reuptake inhibitors on distinct forms of impulsive behavior in rats. *Psychopharmacology*, 219(2), 313–326. <https://doi.org/10.1007/s00213-011-2576-x>
- Balleine, B. W., Delgado, M. R., & Hikosaka, O. (2007). The Role of the Dorsal Striatum in Reward and Decision-Making. *Journal of Neuroscience*, 27(31), 8161–8165. <https://doi.org/10.1523/JNEUROSCI.1554-07.2007>

- Barton, A. C., Kang, H. C., Rinaudo, M. S., Monsma, F. J., Stewart-Fram, R. M., Macinko, J. A., ... Sibley, D. R. (1991). Multiple fluorescent ligands for dopamine receptors. I. Pharmacological characterization and receptor selectivity. *Brain Research*, 547(2), 199–207. [https://doi.org/10.1016/0006-8993\(91\)90963-V](https://doi.org/10.1016/0006-8993(91)90963-V)
- Baskerville, T. A., & Douglas, A. J. (2008). Interactions between dopamine and oxytocin in the control of sexual behaviour. *Progress in Brain Research*, 170, 277–290. [https://doi.org/10.1016/S0079-6123\(08\)00423-8](https://doi.org/10.1016/S0079-6123(08)00423-8)
- Baskerville, T. A., & Douglas, A. J. (2010). Dopamine and Oxytocin Interactions Underlying Behaviors: Potential Contributions to Behavioral Disorders. *CNS Neuroscience & Therapeutics*, 16(3), e92–e123. <https://doi.org/10.1111/j.1755-5949.2010.00154.x>
- Beeler, J. A., Daw, N., Frazier, C. R. M., & Zhuang, X. (2010). Tonic dopamine modulates exploitation of reward learning. *Frontiers in Behavioral Neuroscience*, 4(NOV), 170. <https://doi.org/10.3389/fnbeh.2010.00170>
- Bendix, M., Uvnäs-Moberg, K., Petersson, M., Gustavsson, P., Svanborg, P., Åsberg, M., & Jokinen, J. (2015). Plasma oxytocin and personality traits in psychiatric outpatients. *Psychoneuroendocrinology*, 57, 102–110. <https://doi.org/10.1016/j.psyneuen.2015.04.003>
- Bentzley, B. S., Jhou, T. C., & Aston-Jones, G. (2014). Economic demand predicts addiction-like behavior and therapeutic efficacy of oxytocin in the rat. *Proceedings of the National Academy of Sciences*, 111(32), 11822–11827. <https://doi.org/10.1073/pnas.1406324111>

- Berrada, K., Plesnicher, C. L., Luo, X., & Thibonnier, M. (2000). Dynamic interaction of human vasopressin/oxytocin receptor subtypes with G protein-coupled receptor kinases and protein kinase C after agonist stimulation. *The Journal of Biological Chemistry*, 275(35), 27229–27237. <https://doi.org/10.1074/jbc.M002288200>
- Besson, M., Pelloux, Y., Dilleen, R., Theobald, D. E., Lyon, A., Belin-Rauscent, A., ... Belin, D. (2013). Cocaine Modulation of Frontostriatal Expression of Zif268, D2, and 5-HT2c Receptors in High and Low Impulsive Rats. *Neuropsychopharmacology*, 38(10), 1963–1973. <https://doi.org/10.1038/npp.2013.95>
- Bilder, R. M., Volavka, J., Lachman, H. M., & Grace, A. A. (2004, November). The catechol-O-methyltransferase polymorphism: Relations to the tonic-phasic dopamine hypothesis and neuropsychiatric phenotypes. *Neuropsychopharmacology*. Neuropsychopharmacology. <https://doi.org/10.1038/sj.npp.1300542>
- Boccia, M. L., Petrusz, P., Suzuki, K., Marson, L., & Pedersen, C. A. (2013). Immunohistochemical localization of oxytocin receptors in human brain. *Neuroscience*, 253, 155–164. <https://doi.org/10.1016/j.neuroscience.2013.08.048>
- Bolan, E. A., Kivell, B., Jaligam, V., Oz, M., Jayanthi, L. D., Han, Y., ... Shippenberg, T. S. (2007). D2 receptors regulate dopamine transporter function via an extracellular signal-regulated kinases 1 and 2-dependent and phosphoinositide 3 kinase-independent mechanism. *Molecular Pharmacology*, 71(5), 1222–1232. <https://doi.org/10.1124/mol.106.027763>
- Bowen, M. T., Carson, D. S., Spiro, A., Arnold, J. C., & McGregor, I. S. (2011). Adolescent Oxytocin Exposure Causes Persistent Reductions in Anxiety and Alcohol

- Consumption and Enhances Sociability in Rats. *PLoS ONE*, 6(11).
<https://doi.org/10.1371/journal.pone.0027237>
- Bozorgmehr, A., Moayedi, R., Sadeghi, B., Ghadirivasfi, M., Joghataei, M. T., & Shahbazi, A. (2020). A Novel Link between the Oxytocin Receptor Gene and Impulsivity. *Neuroscience*, 444, 196-208. <https://doi.org/10.1016/j.neuroscience.2020.07.033>.
- Brooks, A. M., & Berns, G. S. Aversive stimuli and loss in the mesocorticolimbic dopamine system, 17 *Trends in Cognitive Sciences* § (2013). Elsevier Current Trends.
- Busnelli, M., Saulière, A., Manning, M., Bouvier, M., Galés, C., & Chini, B. (2012). Functional Selective Oxytocin-derived Agonists Discriminate between Individual G Protein Family Subtypes. *Journal of Biological Chemistry*, 287(6), 3617–3629. <https://doi.org/10.1074/jbc.M111.277178>
- Cardinal, R. N., Pennicott, D. R., Sugathapala, C. L., Robbins, T. W., & Everitt, B. J. (2001). Impulsive Choice Induced in Rats by Lesions of the Nucleus Accumbens Core. *Science*, 292(5526), 2499–2501. <https://doi.org/10.1126/science.1060818>
- Carmona, S., Proal, E., Hoekzema, E. A., Gisbert, J.-D., Picado, M., Moreno, I., ... Vilarroya, O. (2009). Ventro-striatal reductions underpin symptoms of hyperactivity and impulsivity in attention-deficit/hyperactivity disorder. *Biological Psychiatry*, 66(10), 972–977. <https://doi.org/10.1016/j.biopsych.2009.05.013>
- Centonze, D., Grande, C., Usiello, A., Gubellini, P., Erbs, E., Martin, A. B., ... Calabresi,

- P. (2003). Receptor subtypes involved in the presynaptic and postsynaptic actions of dopamine on striatal interneurons. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 23(15), 6245–6254.
- Chang, W. H., Lee, I. H., Chen, K. C., Chi, M. H., Chiu, N.-T., Yao, W. J., ... Chen, P. S. (2014). Oxytocin receptor gene rs53576 polymorphism modulates oxytocin–dopamine interaction and neuroticism traits—A SPECT study. *Psychoneuroendocrinology*, 47, 212–220. <https://doi.org/10.1016/j.psyneuen.2014.05.020>
- Costa, V. D., Tran, V. L., Turchi, J., & Averbek, B. B. (2014). Dopamine modulates novelty seeking behavior during decision making. *Behavioral Neuroscience*, 128(5), 556–566. <https://doi.org/10.1037/a0037128>
- Cragg, S. J., & Greenfield, S. A. (1997). Differential autoreceptor control of somatodendritic and axon terminal dopamine release in substantia nigra, ventral tegmental area, and striatum. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 17(15), 5738–5746.
- Dalley, J. W., Everitt, B. J., & Robbins, T. W. (2011). Impulsivity, Compulsivity, and Top-Down Cognitive Control. *Neuron*, 69(4), 680–694. <https://doi.org/10.1016/j.neuron.2011.01.020>
- de la Mora, M. P., Pérez-Carrera, D., Crespo-Ramírez, M., Tarakanov, A., Fuxe, K., & Borroto-Escuela, D. O. (2016). Signaling in dopamine D2 receptor-oxytocin receptor heterocomplexes and its relevance for the anxiolytic effects of dopamine and oxytocin interactions in the amygdala of the rat. *Biochimica et Biophysica Acta*

- (*BBA*) - *Molecular Basis of Disease*, 1862(11), 2075–2085.
<https://doi.org/10.1016/j.bbadis.2016.07.004>
- Dinn, W. M., Robbins, N. C., & Harris, C. L. (2001). Adult attention-deficit/hyperactivity disorder: Neuropsychological correlates and clinical presentation. *Brain and Cognition*, 46(1–2), 114–121. [https://doi.org/10.1016/S0278-2626\(01\)80046-4](https://doi.org/10.1016/S0278-2626(01)80046-4)
- Dogterom, J., Snijdwint, F. G. M., & Buijs, R. M. (1978). The distribution of vasopressin and oxytocin in the rat brain. *Neuroscience Letters*, 9(4), 341–346.
[https://doi.org/10.1016/0304-3940\(78\)90206-9](https://doi.org/10.1016/0304-3940(78)90206-9)
- Eagle, D. M., Bari, A., & Robbins, T. W. (2008, August 10). The neuropsychopharmacology of action inhibition: Cross-species translation of the stop-signal and go/no-go tasks. *Psychopharmacology*. Springer.
<https://doi.org/10.1007/s00213-008-1127-6>
- Ebstein, R. P., Knafo, A., Mankuta, D., Chew, S. H., & Lai, P. S. (2012). The contributions of oxytocin and vasopressin pathway genes to human behavior. *Hormones and Behavior*, 61(3), 359–379. <https://doi.org/10.1016/j.yhbeh.2011.12.014>
- Ersche, K. D., Jones, P. S., Williams, G. B., Turton, A. J., Robbins, T. W., & Bullmore, E. T. (2012). Abnormal brain structure implicated in stimulant drug addiction. *Science (New York, N.Y.)*, 335(6068), 601–604.
<https://doi.org/10.1126/science.1214463>
- Eubig, P. A., Noe, T. E., Floresco, S. B., Sable, J. J., & Schantz, S. L. (2014). Sex differences in response to amphetamine in adult Long–Evans rats performing a

- delay-discounting task. *Pharmacology Biochemistry and Behavior*, 118, 1–9.
<https://doi.org/10.1016/j.pbb.2013.12.021>
- Evenden, J. L., & Ryan, C. N. (1996). The pharmacology of impulsive behaviour in rats: the effects of drugs on response choice with varying delays of reinforcement. *Psychopharmacology*, 128(2), 161–170.
- Eysenck, S. B. G., & Eysenck, H. J. (1977). The place of impulsiveness in a dimensional system of personality description. *British Journal of Social and Clinical Psychology*, 16(1), 57–68. <https://doi.org/10.1111/j.2044-8260.1977.tb01003.x>
- Fazio, L., Blasi, G., Taurisano, P., Papazacharias, A., Romano, R., Gelao, B., ... Bertolino, A. (2011). D2 receptor genotype and striatal dopamine signaling predict motor cortical activity and behavior in humans. *NeuroImage*, 54(4), 2915–2921.
<https://doi.org/10.1016/j.neuroimage.2010.11.034>
- Floresco, S. B., West, A. R., Ash, B., Moorel, H., & Grace, A. A. (2003). Afferent modulation of dopamine neuron firing differentially regulates tonic and phasic dopamine transmission. *Nature Neuroscience*, 6(9), 968–973.
<https://doi.org/10.1038/nn1103>
- Ford, C. P. (2014). The role of D2-autoreceptors in regulating dopamine neuron activity and transmission. *Neuroscience*. <https://doi.org/10.1016/j.neuroscience.2014.01.025>
- Garibay, J. L., Kozasa, T., Itoh, H., Tsukamoto, T., Matsuoka, M., & Kaziro, Y. (1991). Analysis by mRNA levels of the expression of six G protein alpha-subunit genes in mammalian cells and tissues. *Biochimica et Biophysica Acta*, 1094(2), 193–199.

[https://doi.org/10.1016/0167-4889\(91\)90008-1](https://doi.org/10.1016/0167-4889(91)90008-1)

Ghahremani, D. G., Lee, B., Robertson, C. L., Tabibnia, G., Morgan, A. T., De Shetler, N., ... London, E. D. (2012). Striatal dopamine D₂/D₃ receptors mediate response inhibition and related activity in frontostriatal neural circuitry in humans. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 32(21), 7316–7324. <https://doi.org/10.1523/JNEUROSCI.4284-11.2012>

Grace, A. A. (1991). Phasic versus tonic dopamine release and the modulation of dopamine system responsivity: A hypothesis for the etiology of schizophrenia. *Neuroscience*, 41(1), 1–24. [https://doi.org/10.1016/0306-4522\(91\)90196-U](https://doi.org/10.1016/0306-4522(91)90196-U)

Grace, A. A., & Bunney, B. S. (1984). The control of firing pattern in nigral dopamine neurons: Burst firing. *Journal of Neuroscience*, 4(11), 2877–2890. <https://doi.org/10.1523/jneurosci.04-11-02877.1984>

Gravati, M., Busnelli, M., Bulgheroni, E., Reversi, A., Spaiardi, P., Parenti, M., ... Chini, B. (2010). Dual modulation of inward rectifier potassium currents in olfactory neuronal cells by promiscuous G protein coupling of the oxytocin receptor. *Journal of Neurochemistry*, 114(5), 1424–1435. <https://doi.org/10.1111/j.1471-4159.2010.06861.x>

Hamilton, K. R., Mitchell, M. R., Wing, V. C., Balodis, I. M., Bickel, W. K., Fillmore, M., ... Moeller, F. G. (2015). Choice impulsivity: Definitions, measurement issues, and clinical implications. *Personality Disorders*, 6(2), 182–198. <https://doi.org/10.1037/per0000099>

- Hariri, A. R., Brown, S. M., Williamson, D. E., Flory, J. D., de Wit, H., & Manuck, S. B. (2006). Preference for Immediate over Delayed Rewards Is Associated with Magnitude of Ventral Striatal Activity. *Journal of Neuroscience*, 26(51), 13213–13217. <https://doi.org/10.1523/JNEUROSCI.3446-06.2006>
- Hernández-López, S., Bargas, J., Surmeier, D. J., Reyes, A., & Gálarraga, E. (1997). D1 receptor activation enhances evoked discharge in neostriatal medium spiny neurons by modulating an L-type Ca^{2+} conductance. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 17(9), 3334–3342.
- Hikida, T., Kimura, K., Wada, N., Funabiki, K., & Nakanishi Shigetada, S. (2010). Distinct Roles of Synaptic Transmission in Direct and Indirect Striatal Pathways to Reward and Aversive Behavior. *Neuron*, 66(6), 896–907. <https://doi.org/10.1016/j.neuron.2010.05.011>
- Hong, S.-B., Zalesky, A., Cocchi, L., Fornito, A., Choi, E.-J., Kim, H.-H., ... Yi, S.-H. (2013). Decreased Functional Brain Connectivity in Adolescents with Internet Addiction. *PLoS ONE*, 8(2), e57831. <https://doi.org/10.1371/journal.pone.0057831>
- Humpston, C. S., Wood, C. M., & Robinson, E. S. (2013). Investigating the roles of different monoamine transmitters and impulse control using the 5-choice serial reaction time task. *Journal of Psychopharmacology*, 27(2), 213–221. <https://doi.org/10.1177/0269881112466182>
- Hung, L. W., Neuner, S., Polepalli, J. S., Beier, K. T., Wright, M., Walsh, J. J., ... Malenka, R. C. (2017). Gating of social reward by oxytocin in the ventral tegmental area. *Science* (Vol. 357). <https://doi.org/10.1126/science.aan4994>

- Insel, T. R. (2010). The challenge of translation in social neuroscience: a review of oxytocin, vasopressin, and affiliative behavior. *Neuron*, 65(6), 768–779. <https://doi.org/10.1016/j.neuron.2010.03.005>
- Jentsch, J. D., & Taylor, J. R. (1999). Impulsivity resulting from frontostriatal dysfunction in drug abuse: implications for the control of behavior by reward-related stimuli. *Psychopharmacology*, 146(4), 373–390. <https://doi.org/10.1007/PL00005483>
- Joutsa, J., Voon, V., Johansson, J., Niemelä, S., Bergman, J., & Kaasinen, V. (2015). Dopaminergic function and intertemporal choice. *Translational Psychiatry*, 5(1), e491. <https://doi.org/10.1038/tp.2014.133>
- Justice, J. B. Quantitative microdialysis of neurotransmitters, 48 *Journal of Neuroscience Methods* § (1993). [https://doi.org/10.1016/0165-0270\(93\)90097-B](https://doi.org/10.1016/0165-0270(93)90097-B)
- Kanner, B. I. (1994). Sodium-coupled neurotransmitter transport: structure, function and regulation. *The Journal of Experimental Biology*, 196(1), 237–249.
- Kobayashi, K., Fukabori, R., & Nishizawa, K. (2013). Neural circuit mechanism for learning dependent on dopamine transmission: roles of striatal direct and indirect pathways in sensory discrimination. *Advances in Pharmacology (San Diego, Calif.)*, 68, 143–153. <https://doi.org/10.1016/B978-0-12-411512-5.00007-5>
- Kovács, G. L., Sarnyai, Z., & Szabó, G. (1998). Oxytocin and addiction: a review. *Psychoneuroendocrinology*, 23(8), 945–962.
- Kravitz, A. V, Tomasi, D., LeBlanc, K. H., Baler, R., Volkow, N. D., Bonci, A., & Ferré, S. (2015). Cortico-striatal circuits: Novel therapeutic targets for substance use

- disorders. *Brain Research*, 1628, 186–198.
<https://doi.org/10.1016/j.brainres.2015.03.048>
- Lee, B., London, E. D., Poldrack, R. A., Farahi, J., Nacca, A., Monterosso, J. R., ... Mandelkern, M. A. (2009). Striatal dopamine d2/d3 receptor availability is reduced in methamphetamine dependence and is linked to impulsivity. *The Journal of Neuroscience : The Official Journal of the Society for Neuroscience*, 29(47), 14734–14740. <https://doi.org/10.1523/JNEUROSCI.3765-09.2009>
- Lee, C. R., & Tepper, J. M. (2009). Basal ganglia control of substantia nigra dopaminergic neurons. *Journal of Neural Transmission. Supplementum*, (73), 71–90.
- Leng, G., & Ludwig, M. (2008). Neurotransmitters and peptides: whispered secrets and public announcements. *The Journal of Physiology*, 586(23), 5625–5632.
<https://doi.org/10.1113/jphysiol.2008.159103>
- Liu, C., Wang, J., Zhan, B., & Cheng, G. (2016). Neuronal activity and the expression of hypothalamic oxytocin and vasopressin in social versus cocaine conditioning. *Behavioural Brain Research*, 310, 84–92. <https://doi.org/10.1016/j.bbr.2016.05.010>
- Liu, Y., & Wang, Z. . (2003). Nucleus accumbens oxytocin and dopamine interact to regulate pair bond formation in female prairie voles. *Neuroscience*, 121(3), 537–544.
[https://doi.org/10.1016/S0306-4522\(03\)00555-4](https://doi.org/10.1016/S0306-4522(03)00555-4)
- Lobb, C. J., Wilson, C. J., & Paladini, C. A. (2010). A dynamic role for GABA receptors on the firing pattern of midbrain dopaminergic neurons. *Journal of Neurophysiology*, 104(1), 403–413. <https://doi.org/10.1152/jn.00204.2010>

- Lombardo, L. E., Bearden, C. E., Barrett, J., Brumbaugh, M. S., Pittman, B., Frangou, S., & Glahn, D. C. (2012). Trait impulsivity as an endophenotype for bipolar I disorder. *Bipolar Disorders*, 14(5), 565–570. <https://doi.org/10.1111/j.1399-5618.2012.01035.x>
- Loup, F., Tribollet, E., Dubois-Dauphin, M., & Dreifuss, J. J. (1991). *Localization of high-affinity binding sites for oxytocin and vasopressin in the human brain. An autoradiographic study. Brain Research* (Vol. 555). [https://doi.org/10.1016/0006-8993\(91\)90345-V](https://doi.org/10.1016/0006-8993(91)90345-V)
- Love, T. M. (2014). Oxytocin, motivation and the role of dopamine. *Pharmacology Biochemistry and Behavior*, 119, 49–60. <https://doi.org/10.1016/j.pbb.2013.06.011>
- Love, T. M., Enoch, M.-A., Hodgkinson, C. A., Peciña, M., Mickey, B., Koeppe, R. A., ... Zubieta, J.-K. (2012). Oxytocin gene polymorphisms influence human dopaminergic function in a sex-dependent manner. *Biological Psychiatry*, 72(3), 198–206. <https://doi.org/10.1016/j.biopsych.2012.01.033>
- Magnard, R., Vachez, Y., Carcenac, C., Boulet, S., Houeto, J.-L., Savasta, M., ... Carnicella, S. (2018). Nigrostriatal Dopaminergic Denervation Does Not Promote Impulsive Choice in the Rat: Implication for Impulse Control Disorders in Parkinson's Disease. *Frontiers in Behavioral Neuroscience*, 12, 312. <https://doi.org/10.3389/fnbeh.2018.00312>
- Mai, J. K., Berger, K., & Sofroniew, M. V. (1993). Morphometric evaluation of neurophysin-immunoreactivity in the human brain: pronounced inter-individual variability and evidence for altered staining patterns in schizophrenia. *Journal Für*

Hirnforschung, 34(2), 133–154.

Marcott, P. F., Mamaligas, A. A., & Ford, C. P. (2014). Phasic Dopamine Release Drives Rapid Activation of Striatal D2-Receptors. *Neuron*, 84(1), 164–176. <https://doi.org/10.1016/j.neuron.2014.08.058>

Martinez, E., Pasquereau, B., Saga, Y., Météreau, É., & Tremblay, L. (2019). The Anterior Caudate Nucleus Supports Impulsive Choices Triggered by Pramipexole. *Movement Disorders*, 35(2), 296–305. <https://doi.org/10.1002/mds.27898>

Mayer, A., Limberger, N., & Starke, K. (1988). Transmitter release patterns of noradrenergic, dopaminergic and cholinergic axons in rabbit brain slices during short pulse trains, and the operation of presynaptic autoreceptors. *Naunyn-Schmiedeberg's Archives of Pharmacology*, 338(6), 632–643. <https://doi.org/10.1007/BF00165627>

McGregor, I. S., & Bowen, M. T. (2012). Breaking the loop: Oxytocin as a potential treatment for drug addiction. *Hormones and Behavior*, 61(3), 331–339. <https://doi.org/10.1016/j.yhbeh.2011.12.001>

McHugh, M. J., Demers, C. H., Braud, J., Briggs, R., Adinoff, B., & Stein, E. A. (2013). Striatal-insula circuits in cocaine addiction: implications for impulsivity and relapse risk, 39(6). <https://doi.org/10.3109/00952990.2013.847446>

Meyer-Lindenberg, A., Domes, G., Kirsch, P., & Heinrichs, M. (2011). Oxytocin and vasopressin in the human brain: social neuropeptides for translational medicine. *Nature Reviews Neuroscience*, 12(9), 524–538. <https://doi.org/10.1038/nrn3044>

Milligan, G. (1993). Regional Distribution and Quantitative Measurement of the

- Phosphoinositidase C-Linked Guanine Nucleotide Binding Proteins G_{12} and G_{13} ?
in Rat Brain. *Journal of Neurochemistry*, 61(3), 845–851.
<https://doi.org/10.1111/j.1471-4159.1993.tb03595.x>
- Missale, C., Nash, S. R., Robinson, S. W., Jaber, M., & Caron, M. G. (1998). Dopamine receptors: from structure to function. *Physiological Reviews*, 78(1), 189–225.
- Moos, F., & Richard, P. (1982). Excitatory effect of dopamine on oxytocin and vasopressin reflex releases in the rat. *Brain Research*, 241(2), 249–260.
[https://doi.org/10.1016/0006-8993\(82\)91061-7](https://doi.org/10.1016/0006-8993(82)91061-7)
- Neumann, I. D., & Landgraf, R. (2012). Balance of brain oxytocin and vasopressin: implications for anxiety, depression, and social behaviors. *Trends in Neurosciences*, 35(11), 649–659. <https://doi.org/10.1016/j.tins.2012.08.004>
- Park, J., Willmott, M., Vetuz, G., Toye, C., Kirley, A., Hawi, Z., ... Kent, L. (2010). Evidence that genetic variation in the oxytocin receptor (OXTR) gene influences social cognition in ADHD. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 34(4), 697–702. <https://doi.org/10.1016/j.pnpbp.2010.03.029>
- Perry, J. C., & Körner, A. C. (2011). Impulsive phenomena, the impulsive character (der Triebhafte Charakter) and DSM personality disorders. *Journal of Personality Disorders*, 25(5), 586–606. <https://doi.org/10.1521/pedi.2011.25.5.586>
- Peters, S., Slattery, D. A., Flor, P. J., Neumann, I. D., & Reber, S. O. (2013). Differential effects of baclofen and oxytocin on the increased ethanol consumption following chronic psychosocial stress in mice. *Addiction Biology*, 18(1), 66–77.

<https://doi.org/10.1111/adb.12001>

- Rault, J.-L., Carter, C. S., Garner, J. P., Marchant-Forde, J. N., Richert, B. T., & Lay, D. C. J. (2013). Repeated intranasal oxytocin administration in early life dysregulates the HPA axis and alters social behavior. *Physiology & Behavior*, 112–113, 40–48. <https://doi.org/10.1016/j.physbeh.2013.02.007>
- Renda, C. R., Stein, J. S., & Madden, G. J. (2014). Impulsive Choice Predicts Poor Working Memory in Male Rats. *PLoS ONE*, 9(4), e93263. <https://doi.org/10.1371/journal.pone.0093263>
- Rice, M. E., & Patel, J. C. (2015). Somatodendritic dopamine release: recent mechanistic insights. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 370(1672), 20140185. <https://doi.org/10.1098/rstb.2014.0185>
- Robinson, E. S. J., Eagle, D. M., Economidou, D., Theobald, D. E. H., Mar, A. C., Murphy, E. R., ... Dalley, J. W. (2009). Behavioural characterisation of high impulsivity on the 5-choice serial reaction time task: Specific deficits in “waiting” versus “stopping.” *Behavioural Brain Research*, 196, 310–316. <https://doi.org/10.1016/j.bbr.2008.09.021>
- Roeper, J., Grace, A. A., & Bunney, B. S. (2013). Dissecting the diversity of midbrain dopamine neurons. *Trends in Neurosciences*, 36(6), 336–342. <https://doi.org/10.1016/j.tins.2013.03.003>
- Romero-Fernandez, W., Borroto-Escuela, D. O., Agnati, L. F., & Fuxe, K. (2013). Evidence for the existence of dopamine d2-oxytocin receptor heteromers in the

- ventral and dorsal striatum with facilitatory receptor-receptor interactions. *Molecular Psychiatry*. <https://doi.org/10.1038/mp.2012.103>
- Rosenfeld, A. J., Lieberman, J. A., & Jarskog, L. F. (2011). Oxytocin, dopamine, and the amygdala: a neurofunctional model of social cognitive deficits in schizophrenia. *Schizophrenia Bulletin*, 37(5), 1077–1087. <https://doi.org/10.1093/schbul/sbq015>
- Rung, J. M., Buhusi, C. V., & Madden, G. J. (2018). Reducing impulsive choice: V. The role of timing in delay-exposure training. *Behavioural Processes*. <https://doi.org/10.1016/j.beproc.2018.04.018>
- Schachar, R., Logan, G. D., Robaey, P., Chen, S., Ickowicz, A., & Barr, C. (2007). Restraint and cancellation: Multiple inhibition deficits in attention deficit hyperactivity disorder. *Journal of Abnormal Child Psychology*, 35(2), 229–238. <https://doi.org/10.1007/s10802-006-9075-2>
- Schott, B. H., Minuzzi, L., Krebs, R. M., Elmenhorst, D., Lang, M., Winz, O. H., ... Bauer, A. (2008). Mesolimbic Functional Magnetic Resonance Imaging Activations during Reward Anticipation Correlate with Reward-Related Ventral Striatal Dopamine Release. *Journal of Neuroscience*, 28(52), 14311–14319. <https://doi.org/10.1523/JNEUROSCI.2058-08.2008>
- Schultz, W., Carelli, R. M., & Wightman, R. M. (2015, October 1). Phasic dopamine signals: From subjective reward value to formal economic utility. *Current Opinion in Behavioral Sciences*. Elsevier Ltd. <https://doi.org/10.1016/j.cobeha.2015.09.006>
- Sellitto, M., Ciaramelli, E., & di Pellegrino, G. (2011). The neurobiology of intertemporal

- choice: insight from imaging and lesion studies. *Reviews in the Neurosciences*, 22(5), 565–574. <https://doi.org/10.1515/RNS.2011.046>
- Shahrokh, D. K., Zhang, T.-Y., Diorio, J., Gratton, A., & Meaney, M. J. (2010). Oxytocin-dopamine interactions mediate variations in maternal behavior in the rat. *Endocrinology*, 151(5), 2276–2286. <https://doi.org/10.1210/en.2009-1271>
- Siciliano, C. A., Calipari, E. S., Ferris, M. J., & Jones, S. R. (2015). Adaptations of presynaptic dopamine terminals induced by psychostimulant self-administration. *ACS Chemical Neuroscience*, 6(1), 27–36. <https://doi.org/10.1021/cn5002705>
- Simon, N. W., Beas, B. S., Montgomery, K. S., Haberman, R. P., Bizon, J. L., & Setlow, B. (2013). Prefrontal cortical-striatal dopamine receptor mRNA expression predicts distinct forms of impulsivity. *The European Journal of Neuroscience*, 37(11), 1779–1788. <https://doi.org/10.1111/ejn.12191>
- Skuse, D. H., Gallagher, L., & Frith, C. D. (2009). Dopaminergic-neuropeptide interactions in the social brain. *Trends in Cognitive Sciences*, 13(1), 27–35. <https://doi.org/10.1016/j.tics.2008.09.007>
- Smith, C. T., & Wallace, D. L. (2016). Modulation of impulsivity and reward sensitivity in intertemporal choice by striatal and midbrain dopamine synthesis in healthy adults. *Journal of Neurophysiology*, 115(3), 1146–1156. <https://doi.org/10.1152/jn.00261.2015>
- Sokoloff, P., Diaz, J., Le Foll, B., Guillin, O., Leriche, L., Bezard, E., & Gross, C. (2006). The dopamine D3 receptor: a therapeutic target for the treatment of neuropsychiatric

- disorders. *CNS & Neurological Disorders Drug Targets*, 5(1), 25–43.
- St. Onge, J. R., Ahn, S., Phillips, A. G., & Floresco, S. B. (2012). Dynamic Fluctuations in Dopamine Efflux in the Prefrontal Cortex and Nucleus Accumbens during Risk-Based Decision Making. *Journal of Neuroscience*. <https://doi.org/10.1523/JNEUROSCI.3807-12.2012>
- St Onge, J. R., Stopper, C. M., Zahm, D. S., & Floresco, S. B. (2012b). Separate prefrontal-subcortical circuits mediate different components of risk-based decision making. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 32(8), 2886–2899. <https://doi.org/10.1523/JNEUROSCI.5625-11.2012>
- Stagkourakis, S., Kim, H., Lyons, D. J., & Broberger, C. (2016). Dopamine Autoreceptor Regulation of a Hypothalamic Dopaminergic Network. *Cell Reports*, 15(4), 735–747. <https://doi.org/10.1016/j.celrep.2016.03.062>
- Stoop, R. (2012). Neuromodulation by oxytocin and vasopressin. *Neuron*, 76(1), 142–159. <https://doi.org/10.1016/j.neuron.2012.09.025>
- Stopper, C. M., Tse, M. T. L., Montes, D. R., Wiedman, C. R., & Floresco, S. B. (2014). Overriding phasic dopamine signals redirects action selection during risk/reward decision making. *Neuron*, 84(1), 177–189. <https://doi.org/10.1016/j.neuron.2014.08.033>
- Strickland, J. C., & Johnson, M. W. (2020). Rejecting Impulsivity as a Psychological Construct: A Theoretical, Empirical, and Sociocultural Argument. *Psychological*

Review. <https://doi.org/10.1037/rev0000263>

Succu, S., Sanna, F., Melis, T., Boi, A., Argiolas, A., & Melis, M. R. (2007). Stimulation of dopamine receptors in the paraventricular nucleus of the hypothalamus of male rats induces penile erection and increases extra-cellular dopamine in the nucleus accumbens: Involvement of central oxytocin. *Neuropharmacology*, 52(3), 1034–1043. <https://doi.org/10.1016/j.neuropharm.2006.10.019>

Sulzer, D., Cragg, S. J., & Rice, M. E. (2016). Striatal dopamine neurotransmission: regulation of release and uptake. *Basal Ganglia*, 6(3), 123–148. <https://doi.org/10.1016/j.baga.2016.02.001>

Surmeier, D. J., Ding, J., Day, M., Wang, Z., & Shen, W. (2007). D1 and D2 dopamine-receptor modulation of striatal glutamatergic signaling in striatal medium spiny neurons. *Trends in Neurosciences*, 30(5), 228–235. <https://doi.org/10.1016/j.tins.2007.03.008>

Tapp, D. N., Singstock, M. D., Gottliebson, M. S., & McMurray, M. S. (2020). Central but not peripheral oxytocin administration reduces risk-based decision-making in male rats. *Hormones and Behavior*, 125, 104840. <https://doi.org/10.1016/j.yhbeh.2020.104840>

Tedford, S. E., Persons, A. L., & Napier, T. C. (2015). Dopaminergic lesions of the dorsolateral striatum in rats increase delay discounting in an impulsive choice task. *PloS One*, 10(4), e0122063. <https://doi.org/10.1371/journal.pone.0122063>

Thibonnier, M., & Schork, N. J. (1995). The genetics of hypertension. *Current Opinion*

- in Genetics & Development*, 5(3), 362–370. [https://doi.org/10.1016/0959-437X\(95\)80052-2](https://doi.org/10.1016/0959-437X(95)80052-2)
- Tops, M., Koole, S. L., Ijzerman, H., & Buisman-Pijlman, F. T. A. A. (2014). Why social attachment and oxytocin protect against addiction and stress: Insights from the dynamics between ventral and dorsal corticostriatal systems. *Pharmacology Biochemistry and Behavior*, 119, 39–48. <https://doi.org/10.1016/j.pbb.2013.07.015>
- Tricomi, E., Balleine, B. W., & O'Doherty, J. P. (2009). A specific role for posterior dorsolateral striatum in human habit learning. *European Journal of Neuroscience*, 29(11), 2225–2232. <https://doi.org/10.1111/j.1460-9568.2009.06796.x>
- Van Gaalen, M. M., van Koten, R., Schoffelmeer, A. N. M., & Vanderschuren, L. J. M. J. (2006). Critical Involvement of Dopaminergic Neurotransmission in Impulsive Decision Making. *Biological Psychiatry*, 60(1), 66–73. <https://doi.org/10.1016/j.biopsych.2005.06.005>
- Volkow, N. D., & Baler, R. D. (2015). NOW vs LATER brain circuits: Implications for obesity and addiction. *Trends in Neurosciences*. <https://doi.org/10.1016/j.tins.2015.04.002>
- Volkow, N. D., Tomasi, D., Wang, G.-J., Logan, J., Alexoff, D. L., Jayne, M., ... Du, C. (2014). Stimulant-induced dopamine increases are markedly blunted in active cocaine abusers. *Molecular Psychiatry*, 19(9), 1037–1043. <https://doi.org/10.1038/mp.2014.58>
- Waddington, J. L. (1980). GABAergic mechanisms in the substantia nigra. *Nature*,

283(5748), 696–697. <https://doi.org/10.1038/283696a0>

Wilbertz, G., van Elst, L. T., Delgado, M. R., Maier, S., Feige, B., Philipsen, A., & Blechert, J. (2012). Orbitofrontal reward sensitivity and impulsivity in adult attention deficit hyperactivity disorder. *NeuroImage*, 60(1), 353–361. <https://doi.org/10.1016/j.neuroimage.2011.12.011>

Wilkie, T. M., Scherle, P. A., Strathmann, M. P., Slepak, V. Z., & Simon, M. I. (1991). Characterization of G-protein alpha subunits in the Gq class: expression in murine tissues and in stromal and hematopoietic cell lines., 88(22). <https://doi.org/10.1073/pnas.88.22.10049>

Winstanley, C. A., Olausson, P., Taylor, J. R., & Jentsch, J. D. (2010). Insight Into the Relationship Between Impulsivity and Substance Abuse From Studies Using Animal Models. *Alcoholism: Clinical and Experimental Research*, 34(8), no-no. <https://doi.org/10.1111/j.1530-0277.2010.01215.x>

Winstanley, C. A., Theobald, D. E. H., Cardinal, R. N., & Robbins, T. W. (2004). Contrasting roles of basolateral amygdala and orbitofrontal cortex in impulsive choice. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 24(20), 4718–4722. <https://doi.org/10.1523/JNEUROSCI.5606-03.2004>

Wittmann, M., Leland, D. S., & Paulus, M. P. (2007). Time and decision making: differential contribution of the posterior insular cortex and the striatum during a delay discounting task. *Exp Brain Res*, 179, 643–653. <https://doi.org/10.1007/s00221-006-0822-y>

- Wood, P. L. (1982). Actions of GABAergic agents on dopamine metabolism in the nigrostriatal pathway of the rat. *Journal of Pharmacology and Experimental Therapeutics*, 222(3).
- Xiao, L., Priest, M. F., Nasenbeny, J., Lu, T., & Kozorovitskiy, Y. (2017). Biased Oxytocinergic Modulation of Midbrain Dopamine Systems. *Neuron*, 95(2), 368-384.e5. <https://doi.org/10.1016/j.neuron.2017.06.003>
- Yates, J. R., Perry, J. L., Meyer, A. C., Gipson, C. D., Charnigo, R., & Bardo, M. T. (2014). Role of medial prefrontal and orbitofrontal monoamine transporters and receptors in performance in an adjusting delay discounting procedure. *Brain Research*, 1574, 26–36. <https://doi.org/10.1016/j.brainres.2014.06.004>
- Yawata, S., Yamaguchi, T., Danjo, T., Hikida, T., & Nakanishi, S. (2012). Pathway-specific control of reward learning and its flexibility via selective dopamine receptors in the nucleus accumbens. *Proceedings of the National Academy of Sciences of the United States of America*, 109(31), 12764–12769. <https://doi.org/10.1073/pnas.1210797109>
- Yoshida, M., Takayanagi, Y., Inoue, K., Kimura, T., Young, L. J., Onaka, T., & Nishimori, K. (2009). Evidence that oxytocin exerts anxiolytic effects via oxytocin receptor expressed in serotonergic neurons in mice. *The Journal of Neuroscience : The Official Journal of the Society for Neuroscience*, 29(7), 2259–2271. <https://doi.org/10.1523/JNEUROSCI.5593-08.2009>
- Young, L. J., & Wang, Z. (2004). The neurobiology of pair bonding. *Nature Neuroscience*, 7(10), 1048–1054. <https://doi.org/10.1038/nm1327>

- Zhang, H., Li, S., Wang, M., Vukusic, B., Pristupa, Z. B., & Liu, F. (2009). Regulation of dopamine transporter activity by carboxypeptidase E. *Molecular Brain*, 2(1), 10.
<https://doi.org/10.1186/1756-6606-2-10>
- Zink, C. F., & Meyer-Lindenberg, A. (2012). Human neuroimaging of oxytocin and vasopressin in social cognition. *Hormones and Behavior*, 61(3), 400–409.
<https://doi.org/10.1016/j.yhbeh.2012.01.016>