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Heme Oxygenase-1 As a Target for the Design of Gene and Pharmaceutical Therapies for Autoimmune Diseases

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Abstract: One of the major goals in the research of autoimmune diseases is to develop specific therapies to regulate the expression and function of gene products that could contribute to restoring tolerance to self-constituents and replace conventional systemic immunosuppression, which is associated with important undesired side effects. Although significant progress has been made on the understanding of the pathogenesis of autoimmunity, therapies for these ailments have not seen a change. During the last decade, different strategies such as pharmacologic or gene therapy modulation of heme oxygenase-1 (HO-1) and the administration of its metabolic product, carbon monoxide (CO), have been shown to display beneficial immunoregulatory and cytoprotective properties. In different experimental autoimmune conditions, such as Experimental autoimmune encephalomyelitis, type-1 diabetes and systemic lupus erythematosus, genetic or pharmacological modulation of HO-1, as well as delivery of CO have shown to ameliorate disease progression. Furthermore, it has been demonstrated that dendritic cell and monocyte function can be modulated by HO-1 and/or CO. In this article, recent data related to the immunoregulatory properties of HO-1/CO will be discussed, focusing on their potential therapeutic use to treat autoimmune diseases.

Keywords: Heme oxygenase-1, carbon monoxide, autoimmunity, tolerance, therapy.

1. INTRODUCTION

 The development of autoimmunity is characterized by an auto-reactive immune response capable of injuring healthy host cells and tissues. The etiology of autoimmune diseases is unknown and associated with a confluence of genetic predisposition and environmental exposure [1]. During the last decade, important progress has been made in the characterization of the molecular mechanisms responsible for the onset and progression of autoimmunity, such as the involvement of Th17 related cytokines in multiple sclerosis [2, 3] and type I interferons in systemic lupus erythematosus (SLE) [4, 5]. Although this knowledge has promoted the development of anti-inflammatory drugs and biological agents to interfere with the immunepathogenesis, the current autoimmune therapies have not changed much in clinical practice with the advance of immunology. Thus, there is an urgent need for therapy improvement in terms of efficacy, adverse effects and specificity. Because cell-destruction and tissue damage during autoimmunity are mainly caused by immune cellmediated inflammation [6-8], new therapies should be

designed to target these inflammatory processes and suppress the damaging consequences of autoimmunity.

Heme oxygenase-1 (HO-1) is a powerful antiinflammatory enzyme, which plays a critical role in stress conditions such as organ transplantation and hypertension [9- 12]. We have previously reported that the restoration of HO-1 levels and administration of HO-1-derived metabolic product decreases inflammation in a mouse model of SLE [13]. Further, treatment with inducers of the HO-1/CO axis has also been shown to be effective at promoting tolerance in other pathologic conditions such as type 1 diabetes [14, 15] and neuroinflammation [16, 17]. We have recently described that patients with SLE, as well as lupus prone mice, show decreased levels of HO-1 in peripheral blood monocytes [13, 18]. In this context, HO-1 induction arises as a promising treatment to decrease inflammation in autoimmune diseases.

 In this review, we highlight and discuss current knowledge about HO-1 as a potential target for the treatment of autoimmune diseases. In addition, we address recent *in vitro* results showing the decrease of inflammation mediated by myeloid immune cells caused by metabolic products derived from heme degradation by HO-1.

2. HO-1 BIOLOGY AND TISSUE EXPRESSION

 Hemeoxygenases or haemoxygenases (HO) are enzymes that regulate cellular levels of the heme group [19]. A large

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proportion of heme transport and utilization is carried out by macrophages, either by defective erythrocyte phagocytosis or upon CD163-mediated uptake of hemoglobin–haptoglobin complexes [20]. These processes mainly occur in tissues enriched in macrophages, such as spleen and liver [19, 21]. The HO enzyme catabolyzes heme degradation into Fe^{2+} , biliverdin and carbon monoxide (CO) and is the rate-limiting enzyme in heme compounds degradation. Biliverdin is subsequently converted into bilirubin in a reaction catalyzed by the biliverdin reductase enzyme [22]. HO can modulate the function of some hemeproteins including nitric oxide synthase, soluble guanylatecyclase, cytochrome P450, peroxidase and catalase through the degradation of the heme group and CO release, which binds to this group, thereby inhibiting hemeproteins activity [19].

 There are three isoenzymes of HO: an inducible isoform designated HO-1 and two constitutively expressed isoforms, known as HO-2 and HO-3 [23]. There is great diversity in the tissue distribution of HO-1 and HO-2 isoforms [22]. HO-1 is a heat shock and a stress protein induced by oxidative agents [24, 25], which is constitutively expressed in testis, brain, placenta and the vascular system [26-30]. On the other hand, HO-2 is highly expressed in epidermal cells, germ cells and neural tissues, playing a relevant role in the normal function of these organs [31, 32]. Moreover, HO-2 may protect neural tissues from oxidative stress by reducing lipid peroxidation through the catabolism of free heme [33]. In addition, it has been recently shown in mouse models that HO-2 is capable of protecting from renovascular hypertension and cardiac hypertrophy [29]. Thus, HO-2 is an important component of cells that constitutively participate in the regulation of physiological processes. In contrast, studies focusing on HO-3 enzyme are still rather limited [34].

 The HO-1 isoform is mainly induced by heme, oxidative or free radical molecules during inflammatory conditions [9, 24, 25]. HO-1 is expressed in different cell types, such as hepatocytes [35], endothelial cells [36], myeloid cells [37, 38] and cells from the respiratory tract [39]. HO-1 expression can be induced through the activation of different signaling pathways, including cyclic adenosine-5 monophosphate (cAMP)-dependent mechanisms [40], protein kinase C (PKC), phospholipase A2 (PLA2) [41] and by mitogen-activated protein kinases (ERK and P38) [42, 43]. In addition, fine tuning of HO-1 mRNA and protein levels is achieved by the activity of transcription factors Nrf2 and Bach1 [44]. It is thought that a protein known as Keap1 keeps Nrf2 in the cytoplasm preventing nuclear translocation. In response to oxidative stress conditions, ROS can react with Keap1 sulfhydryl groups, which promotes Nrf2 release and translocation into the nucleus [45]. Nrf2 binds to small Maf proteins to form heterodimers and transactivates the antioxidant response elements (ARE), which induces the expression of antioxidative gene products, such as HO-1, NADPH-generating enzymes, glutathione biosynthesis enzymes, chaperone proteins and ubiquitin-proteasomes [45]. These ARE gene products react with ROS derived from environmental and intracellular sources, limiting cellular damage. On the contrary, Bach1/2 with small Maf proteins binds to Maf response elements (MARE), including ARE, to inhibit HO-1 gene transcription and allow HO-1 transcription when heme-Bach1 interaction occurs (Fig. **1**) [46, 47].

 Although the main function of HO-1 is to degrade free heme, several immunoregulatory features have been attributed to this enzyme. The anti-inflammatory role of HO-1 is thought to be mediated by the by-products generated after heme group degradation, which will be discussed in the following sections.

3. HO-1 PROMOTES CELL VIABILITY

Antioxidant Capacity of HO-1

 Heme group degradation plays an important role in cellular defense and survival. The HO-1 cytoprotective effects may reduce pathogenesis during several oxidation-mediated inflammatory diseases by limiting both, oxidative stress and apoptosis. HO-1 contributes to regulate intracellular prooxidant systems by degrading heme group and releasing CO from hemeproteins and enzymes, such as iNOS, peroxidase and catalase [19]. These reactions also promote the binding of ferritin to the free iron released from HO-1 catalysis over hemeproteins, which prevents free radical formation [48]. Moreover, it has been shown that cultured astrocytes from rat forebrain increase HO-1 expression after hydrogen peroxide exposure, preventing oxidative stress-mediated injury [33]. Further, in a mouse model of ischemic stroke with middle cerebral artery occlusion, HO-1 overexpression protects from injury, showing an increased expression of ferritin, decreased iron deposits and reduced lipid peroxidation [49]. Likewise, in diabetic rats, HO-1 induction increases superoxide dismutase and catalase activity, decreasing reactive oxygen species(ROS) levels in aortic tissue [50].

Anti-apoptotic Properties of HO-1

 The anti-apoptotic capacity of HO-1 has been demonstrated at different levels. In primary cultures of cerebrovascular endothelial cells from newborn pigs, the induction of HO-1 inhibited caspase 3 activity after glutamate-induced cell death [51]. After cerebral ischemia, transgenic mice that overexpress HO-1 showed neuroprotection by enhancing the expression of the antiapoptotic factor Bcl-2 [49]. Also, by using an *in vitro* model of anoxia-reoxygenation with rat pulmonary endothelial cells, it has been suggested that CO released by HO-1 catalysis activates p38 MAPK signaling pathway, limiting caspase-3 activity and inhibiting cell death [52]. In human neutrophils, it has been shown that heme decreases Bad degradation -a proapoptotic Bcl-2 member- and induces Bcl-xL expression, limiting the spontaneous mitochondrial apoptotic pathway [52, 53]. More importantly, Kahlo *et al.* showed that human trophoblasts from spontaneous abortions expressed low levels of HO-1 which were associated with a decreased expression of the anti-apoptotic molecule Bag-1 [54]. Furthermore, it has been shown that HO-1 overexpression by gene transfer increased Bcl-2 and Bag-1 levels in a model of liver transplantation [55]. In addition, HO-1 induction decreases cell death of PC12 epithelial cells by preventing the depolarization of mitochondrial transmembrane potential [56]. Furthermore, HO-1 induction and CO administration to mice improve kidney function following ischemia and protect against treatment with cytostatic drugs [57, 58]. Cisplatin is an anti-neoplastic drug, which leads to severe impairment of renal function by oxidative stress and caspase-mediated apoptosis. Interestingly, the

Fig. (1). *Bach1 and Nrf2 transcription factors modulate HO-1 mRNA transcription.* Under normal conditions, Keap1 sequesters Nrf2 in the cytoplasm and Bach1/2 with small Maf proteins bind to Maf response elements (MARE), including antioxidant response elements (ARE), repressing HO-1 gene transcription and allowing HO-1 transcription when heme-Bach1 interaction occurs. In human, a dinucleotide guanine-thymidine repeat $(GT)n_{15-40}$ polymorphism at HO-1 promoter region may impact in determining the level of the transcription induction. In stress conditions, such as oxidative stress, ROS may react to Keap1 releasing Nrf2 to translocate into the nucleus. Nrf2 forms heterodimers with small Maf proteins and binds to ARE to increase the expression of antioxidant enzymes such as HO-1 and glutathione biosynthesis enzymes. These antioxidant enzymes remove ROS derived from environmental and intracellular sources, limiting further oxidative damaged. Heme (and classical HO-1 inducer such as CoPP) binds to Bach1, thus allowing Nrf2 to induce HO-1 transcription. HO-1 protein degrades heme group producing CO, biliverdin and Fe^{2+} leading to cytoprotective, antiapoptotic and antiinflammatory effects. Mn-, Sn- and Zn-PP inhibit HO-1 activity by competitive blocker of heme substrate.

anti-apoptotic effect mediated by HO-1 improves transplant survival. In a recent study, it has been shown that HO-1 induction by a retroviral vector suppresses $TNF-\alpha$ -induced cell death [59]. Thus, HO-1 arises as a cell sentinel mediating anti-apoptotic effects associated with inflammatory stimuli that, if sustained are likely to cause release of intracellular components that can support a positive feedback for inflammation. HO-1 overexpression in the mitochondria or in the vicinity of mitochondria membranes is associated with a change in redox status of cells, which would prevent Cytochrome C release and the activation of caspases, which would contribute to prevent cell death [60, 61].

Cytoprotective Effects of HO-1 and its by-products

 HO-1 may also have anti-proliferative properties that are mediated by CO and biliverdin. While proliferation of vascular smooth muscle cells contributes to atherosclerosis pathogenesis, it has been shown that CO and biliverdin can reduce this process and induce cell cycle arrest by suppressing the expression of cdk2 activators and cyclin D1 [62-64].

 CO, in conjunction with bilirubin, may exert different functions, including the modulation of adiponectin levels, which prevents adipogenesis, inflammation and myocardial ischemia [65], the regulation of vascular function and stimulation of ion channels [66] leading to vasorelaxant effects [67]. Furthermore, CO inhibits cytochrome P450 activity, leading to vasoconstriction inhibition [68]. In addition, it has been suggested that CO could by itself contribute to cellular signaling, mainly as an activator of guanylatecyclase [69].

4. PHARMACOLOGICAL AND GENETIC TOOLS TO MODULATE HO-1 ACTIVITY

Pharmacologic Induction of HO-1 Activity

 Given that HO-1 has cytoprotective properties, this enzyme has been postulated as a potential therapeutic target for immune-mediated diseases where both, cell integrity and balance, can be drastically compromised. Thus, upregulation of HO-1 activity could be of clinical interest for these ailments. In experimental animal models, it has been demonstrated that HO-1 activity is increased in whole tissues after treatment with its natural substrate heme, as well as various metals, xenobiotics, endocrine factors, synthetic metalloporphyrins and stress factors [22]- Cobalt protoporphyrin IX (CoPP) has been reported as a classical inducer of the HO-1 expression in different tissues and capable of increasing significantly this enzyme activity (Fig. **1**) [22]. However, it has been shown that CoPP could also act as a competitive inhibitor of the activity of the HO system in *in*

vitro assays [70, 71]. In addition, hemin has also been shown to be a potent HO-1 inductor *in vivo* [22]. Interestingly, aspirin, one of the most common anti-inflammatory drugs, also induces HO-1 expression [72]. Moreover, there are some less common molecules that act as HO-1 inducers, such as AG1067, a probucol analog [73], probucol [22], paclitaxel [22], rapamycin [22] and curcumin [74].

 Biliverdin and bilirubin can be protective by preventing oxidation-mediated cell death and contribute to endothelial cell integrity by their antioxidant potential [75]. However, due to the potentially detrimental effects that accumulation of these molecules could cause in some tissues, their use in the clinic remains uncertain. For instance, high levels bilirubin are harmful to the central nervous system [76]. However, regulated biliverdin administration can be beneficial in several experimental models of disease, as this molecule can protect against lung injury due to lipopolysaccharide-induced shock [77], it can prevent liver damage caused by ischemia and reperfusion [78] and it can increase islet allograft due to its immune-suppressive and antioxidant properties [79, 80]. Furthermore, weekly administration of non-toxic amounts of CoPP in a mouse model for type 1 diabetes increased adiponectin levels, which promotes resistance to oxidative stress and reduces pathogenesis [81]. Moreover, it has been demonstrated that hemin causes a strong induction of HO-1 expression in leukocytes, after administration in a single intravenous dose, in a phase I clinical trial [82].

 Importantly and together with HO-1 induction, continuous administration of high amounts of CoPP as an HO-1 inducer may be toxic, mainly by consuming heme and therefore decreasing hemoproteins, such as cytochrome P450 [83- 85]. Thus, excessive or sustained administration of molecules that induce HO-1 could be deleterious for the patient.

 Although CO therapy may be beneficial in a wide spectrum of disorders such as multiple sclerosis, Type 1 Diabetes, SLE and nephrotoxicity, exposure to high levels of CO may be harmful [13, 17, 58, 86, 87]. In fact, CO binds hemeproteins and blocks mitochondrial electron transport, resulting in a release of detrimental superoxide anions, elevated titers of carboxyhemoglobin and reduction of oxygen transport to different tissues [87, 88]. Different molecules that can supply CO in solution and prolong its effects have been developed, such as CO-releasing molecules (CORMs) [89], which are effective *in vivo* and *in vitro* [90-92].

 Thus, rational administration of HO-1 inducers could increase the activity of enzyme with a sustained delivery of their products, such as CO and biliverdin/bilirubin that can be of potential use for clinical application in several diseases, such as hypertension, diabetes, cardiovascular disease and immune-mediated diseases.

 While most therapeutic approaches have focused on augmenting HO-1 activity, some pathologic conditions could be tackled by inhibiting HO-1 activity. In 1990s, HO-1 blockade using Sn-mesoporphyrin resolved the problem of progressive jaundice in newborns at risk of brain damage due to uncontrolled hyperbilirubinemia [22, 75, 76]. Blocking HO-1 activity in neonates was one of the first clues of HO-1 benefits in a clinical setting. It has also been described that treatment with zinc-protoporphyrin (ZnPP) improves mouse survival in a tumor-induced model mainly by making tumor cells more

sensible to reactive oxygen species derived from the antitumor response [93]. Besides ZnPP, metalloporphyrins such as Snand Mn- protoporphyrin are potent competitive inhibitors of the HO-1 enzymatic activity (Fig. **1**) [22]. Although ZnPP is a potent HO-1 inhibitor, it has also been reported *in vitro* that ZnPP, at low concentrations, can produce a synergism of HO-1 induction by heme [94]. In contrast to HO-1 induction, there is no gene therapy related to HO-1 inhibition.

HO-1 Transcriptional Regulation by a Dinucleotide Guanosine-thymine Repeat Polymorphism

 Transcriptional activity of HO-1 coding gene (HMOX1) can be regulated by a dinucleotide guanosine-thymine repeat (GT)n polymorphism at the promoter region, which determines the expression level of this enzyme [95]. Polymorphisms with short (GT)n repeats $(< 25$) promote higher levels of HO-1 expression as compared to long (GT)n repeats (n > 25) [95]. It has been suggested that (GT)n microsatellite length may influence the susceptibility to different diseases, including inflammatory ailments [95]. Previous studies have shown that short HMOX1 (GT)n polymorphisms associate with susceptibility to childhood-onset SLE but not with Juvenile Rheumatoid arthritis (JRA) [96]. Furthermore, RA patients carrying the short polymorphism showed a better clinical prognosis than did those with the long polymorphism [95]. Healthy individuals carrying the short polymorphism showed a higher percentage of monocytes expressing HO-1 than did individuals with the long polymorphism [97]. Thus, the short polymorphism displayed a significant protective effect in RA susceptibility [97]. Furthermore, these short and long polymorphisms have been associated with other pathologic conditions. For example, type 2 diabetes patients carrying the long polymorphism showed higher lipid peroxidation derived molecules in serum and higher frequencies of coronary artery disease than did patients with the short polymorphism [98, 99] (Fig. **1**). These observations suggest that HMOX1 polymorphism may impact on the HO-1 activity modulating pathologic conditions, such as RA, coronary artery disease and diabetes and may provide a novel target for immune mediated diseases treatment.

Gene Therapy to Modulate HO-1 Expression

 Genetic tools have recently been designed to promote HO-1 induction in different tissues to promote cytoprotection and cell survival. Viral vectors carrying HO-1 gene have been used as treatment for different pathologic conditions [10, 59, 100-103].

 While much work has been done to evaluate HO-1 gene therapy for transplant rejection and vascular diseases, there are scarce reports testing the effectiveness of HO-1 gene delivery in inflammatory diseases. In a muscular dystrophy model, it was shown that HO-1 lentiviral infection of porcine myogenic precursor cells can protect from staurosporine-induced apoptosis *in vitro* [104]*.* In addition, when these HO-1-transduced myogenic precursors were transplanted in pig skeletal muscle, a fivefold increase in cell survival was reported five days after transplantation [104]. Further, microinjection of adenovirus vector carrying human HO-1 gene (Ad-hHO-1) into the eye produced a local increase in the enzymatic activity, which was undetectable in peripheral tissues. This approach could be considered as a potential therapy for oxidative stress-mediated ocular disease [105]. In contrast, when Ad-hHO-1 was delivered intravenously it resulted in expression of human HO-1 mRNA in several tissues including liver, kidney, heart, lung, and brain, which may be considered as a potential therapy for different conditions, such as tissue toxicity and hypertension [106]. Further, transduction of bone marrow-derived macrophages with an adenovirus vector containing HO-1 gene results in overexpression of this enzyme with a consequent reduction of nitric oxide (NO) and TNF α release, augmenting IL10 production after LPS stimulation [107]. In a model for renal ischemia/reperfusion injury (IRI), Ad-hHO-1-transduced macrophages injected intravenously localized preferentially and rapidly at the injured kidney prevented local platelet and fibrin aggregation and improved renal function [107]. Similar results were obtained in a model of liver transplantation when Ad-hHO-1-transduced macrophages transferred simultaneously with transplanted hepatocytes led to an extension in animal survival due to a reduction in the expression of TLR-4, as well as in the production of proinflammatory cytokines and liver infiltration [108]. Intracardiac delivery of retroviral vectors carrying HO-1 gene reduces the production of the vasoconstrictor 20-hydroxyeicosatetraenoic acid (20-HETE) and ameliorates hypertension in spontaneously hypertensive rats [109]. In addition, it has been reported that the transfer of adenoviral vectors encoding for HO-1 to pregnant mice undergoing abortion improved pregnancy outcome [110]. Authors showed that HO-1 gene therapy reduced the pro-inflammatory cy tokine IFN γ and increased IL4 production in both spleen and decidual lymphocytes, which is important in placental tolerance [110, 111]. In a model of chronic allogeneic aorta rejection in rats, it was shown that Ad-hHO-1-transduced aorta overexpressed HO-1 after ten days of allogeneic transplantation, leading to a reduction of intima and adventitia layer infiltration by $CD45^+$ cells, macrophages and $CD4^+$ T cells, as well as reduced expression of co-stimulatory molecules, which results in a favorable graft outcome [102]. Further, it was shown that Ad-hHO-1 transduction of pancreatic islets improves allogenic transplantation survival by reducing leukocyte infiltration and apoptosis [112].

 Additionally, transgenic animals overexpressing HO-1 (tHO-1) have been generated showing similar results as obtained by pharmacologic induction or by the transduction of viral vectors encoding for HO-1 [113, 114]. tHO-1 mice expressing HO-1 under the control of the neuron-specific enolase promoter showed reduced stroke volumes and limited ischemic cerebral edema [114]. In a model of cardiac allograft transplantation, tHO-1 heart graft donor into WT recipient or WT heart graft donor into tHO-1 recipient improved allograft survival. Moreover, WT hearts transplanted into tHO-1 recipients displayed a decrease in $CD4^{\text{+}}$ T cell infiltration and activation measured by CD25 expression, prolonging graft survival [115]. Hearts transplanted into HO- $\frac{1}{1}$ Tg recipients showed decreased CD4⁺ lymphocyte infiltration and diminished immune activation, as determined by CD25 expression [115]. All these results suggest that HO-1 gene therapy could be considered as a potential strategy to improve therapies based on cell transfer or transplantation.

HO-1 Deficiency

 The first reported patient with HO-1 deficiency occurred in 1999 which showed growth retardation, hemolytic anemia, abnormal coagulation function, premature death, endothelial damage and atherosclerotic changes in the vasculature, linking HO-1 system with atherosclerosis [116]. Also, this patient showed major alterations in the immune system such as leukocytosis and lymph node swelling.

 Similarly to human HO-1 deficiency, HO-1 KO mice showed splenomegaly and lymph node swelling, leukocytosis, altered CD4⁺ T cells ratio, and an elevated amount of activated T cells [117]. Also, HO-1 KO mice showed increased vulnerability and a decreased survival in response to LPS-mediated challenge, a phenomenon that is thought to be associated to the modulation of mitochondrial function by HO-1 [117]. Furthermore, HO-1 KO mice showed an increase in cytokine responses, such as IL-1, IL-6 and TNF- α in LPS-stimulated splenocytes [118]. Similarly, when HO-1 KO splenocytes were stimulated with CD3/CD28 antibodies, the production of IL-2, IFN- γ and other cytokines was increased [118]. The exacerbated immune responses shown by HO-1 KO mice suggested that HO-1 activity may be closely related to a regulatory role in the activation of myeloid and lymphoid cells.

5. THE ROLE OF HO-1 IN IMMUNE CELLS

The Role of HO-1 in the Regulation of Dendritic Cells Immunogenicity

 Several inflammatory diseases, such as autoimmune diseases, are caused by alteration in the function of T cells [119-122]. The onset of this exacerbated response is triggered when antigen presenting cells (APCs) capture selfantigens, degrade and present them to self-reactive naïve T cells. The most important APCs with the capacity to prime resting T cell are the DCs [123]. These cells mainly reside in peripheral tissues where they constantly sense antigens and danger signals [124-128], to transfer these to lymphoid organs and present these antigens to naïve and memory T cells [123, 126], thus eliciting an antigen-specific adaptive immune response. In response to danger-inflammatory signals, DCs undergo a process known as maturation [126-128]. It has been reported that in the absence of pro-maturation stimuli, DCs remain unable to successfully process and present antigens to naïve T cells [129, 130]. DC maturation is important because it enhances the communication with T cells during antigen presentation, which involves TCR binding to MHC-peptide complexes, the expression of co-stimulatory molecules and the release of pro-inflammatory cytokines, such as IL-12 and IL-6 [123, 131-134]. In addition, DCs can also secrete anti-inflammatory cytokines such as IL-10, which regulate T cell priming by counteracting the inflammatory environment [135].

 Recently, it has been reported that HO-1 induction by CoPP in humans and rats impairs DC maturation during LPSmediated stimulus [38] (Fig. **2A**). Interestingly, HO-1 induction was not only associated with the inhibition of maturation markers, but also with the expression and secretion of antiinflammatory molecules. This was the case for IL-10, an antiinflammatory cytokine whose secretion by DCs was not altered by the induction of HO-1 expression by CoPP, supporting the tolerogenic profile promoted by the activity of this enzyme (Fig. **2A**). Moreover, HO-1 induction can impair the intracellular LPS-induced generation of ROS, which is a typical process of the innate immune that contributes to antigen processing [38, 136]. In this context, we have shown that maturation of murine and human DCs can also be inhibited by CO [92]. Thus, inhibition of DC maturation by HO-1 is attributable to CO with a reduced activation of IRF3 pathway, leading to a subsequent decrease in IL-12 levels [92] (Fig. **2A**). However, Mashreghi *et al.* reported that CoPP inhibition of DCs maturation by LPS stimulation is independent of HO-1 and dependent on STAT3 [137]. Authors showed that CoPP prevented maturation of DCs from both HO-1 deficient and wild type mice while STAT3 inhibition reversed the antiinflammatory effects of CoPP on DCs [137].

 As the main roles of DCs are the capture and presentation of antigens (including autoantigens) to T cells, HO-1 induction could promote the tolerogenic capacity of DCs and reduce the activation of autoreactive T cells. It has been demonstrated that plasmacytoid DCs from SLE patients release large amounts of IFN-alpha (IFN- α), which induces the maturation of conventional DCs, that in turn activates autoreactive T and B cells [138, 139]. In this context, it has been shown that treating HO-1 induction in DCs or treatment with CO reduces antigen presentation in both MHC-I and –II [140]. Interestingly, only the presentation of small soluble antigens (proteins), but not the presentation of antigens associated to large particles, such as 3 μ m latex beads in DCs was impaired by CO treatment. Thus, it seems that antigen size is important for antigen presentation inhibition by HO-1 activity. Along these lines, it has been described that small antigens, such as ovalbumin (OVA), can enter DCs by the mannose receptor (MR) or the pinocytic route [141, 142]. Once intracellular, MR-driven capture of OVA leads antigens to $Rab5⁺$ compartments where proteasomal degradation takes place. Then, class I MHC presentation is rapidly directed toward cells surface. OVA captured by pinocytosis follows the vesicular pathway, leading antigens to a sequential fusion of endosomal compartments that finally converge in the fusion of late endosome-containing OVA with lysosomes [129, 141, 143, 144]. Thus, interference with both MR and pinocytic routes can lead DCs to decrease their capacity to target soluble antigens to degradative compartments and present them on MHC-I and –II molecules. Consistent with this notion, we have recently shown that either HO-1 induction or treatment of murine DCs with CO at low doses impairs presentation of soluble antigens to T cells [140]. We observed that CO impairs the targeting of soluble OVA to lysosomes by stalling this antigen in late endosomes. Despite OVA was fully degraded in late endosomal compartment, fusion of these vesicles with lysosomes did not take place, preventing the loading of peptides on MHC-I and -II molecules. No alterations were observed for the proteasomal/degradative route, because both targeting of soluble antigens to early endosomes (Rab5⁺) and presentation of OVA as an endogenous antigen were not impaired by CO. Thus, CO produced by HO-1 decreased the capacity of DCs to present soluble antigens by reducing the destination of these molecules to lysosomal compartments and thus reducing the generation of pMHC complexes. Interestingly, we did not observe any alteration in the capacity of DCs to present bead-bound OVA. These large particles were captured by DCs trough phagocytosis. It has been recently proposed that large particles are captured by phagocytic cells by using, in part, the endoplasmic reticulum (ER) membrane [145]. The fusion of early

phagosomes with ER produces a large vesicle known as the Ergosome [145-147]. It has been shown that this compartment is able to produce pMHC-I complexes even before fusing with lysosomes [147]. Thus, despite CO impairs endosomal LPSdriven soluble OVA processing, the mechanism by which large phagosomes are processed within DCs renders them resistant to HO-1 activity, because it seems that they do not require fusion with lysosomes. However, recent findings have proposed that large particles, which are captured by ERmediated phagocytosis, acquired both MHC-I and -II in conjunction with other lysosomal and ERs proteins [148]. Thus, it is proposed that large cargo rapidly fuses with compartments that carry MHC-I and -II molecules and, after antigen degradation, emerging peptides can be loaded on MHCs. These peptides are derived from the phagosome-to-cytosol process with subsequent proteasomal degradation and reinternalization of fragments into the vesicle using TAPs proteins [146-148]. Also, antigens can be processed by phagoluminal lysosomal proteases such as Cathepsin D [146-148]. Interestingly, despite it still remains controversial, phagosome maturation inside APCs has been proposed as LPSindependent [149, 150]. Thus, despite HO-1 activity, by means of CO, is able to reduce LPS-dependent endosomal pathways by causing a stalling in the soluble antigen trafficking in DCs (Fig. **2B**), CO is unable to block Ergosome maturation and antigen presentation for large particles.

 Capture of external soluble antigens is a conserved mechanism by which DCs sense foreign and self-antigens to promote either activation or tolerance by T cells [151]. During graft rejection, DCs play an important role in directing the inflammatory response that finally leads to the destruction of the engrafted tissue [152, 153]. Disruption of new grafts has been associated with the activation of T cells that secrete elevated titers of pro-inflammatory cytokines that impair cell-to-cell junctions together with activated cytotoxic $T \overline{C}D8^+$ cells. Thus, reducing \overline{T} cell activation is a promising alternative to decrease graft rejection. However, as described above, T cells become activated after receiving stimulus from innate immune cells designed to present extracellular antigens, such as DCs. In this context, it has been recently proposed that targeting the function of these cells is a promising therapy to generate antigen-specific tolerance [153]. As inflammation associated to graft rejection is similar in various aspects to that observed for autoimmune diseases, it is possible that we could learn about how to handle selfinflammatory conditions by studying graft rejection models and the tools used in this field by conferring tolerance to certain antigens. It has been shown that transference of tolerogenic DCs significantly reduces graft rejection [152- 154] and that induction of HO-1 *in vivo* protects against graft rejection [101, 102, 155-157]. CO has been postulated as the candidate molecule that mediates the tolerogenic profile that supports graft acceptance in these studies [102]. Moreover, CO exposure or HO-1 induction in donor organs reduced the immunogenicity of DCs by decreasing MHCII and costimulatory molecules thus improving graft survival [158]. Also, Schumacher *et al.* state that HO-1 expression on DCs contributes to maintain pregnancy mainly by inducing an immature phenotype and avoiding presentation of paternal antigens to maternal T cells [159]. Consistent with this notion is the observation that the transfer of hemagglutinin

Fig. (2). *HO-1 produces CO and impairs both the production of Fig. (3) inflammatory cytokines and the presentation of soluble antigens in myeloid cells*. A) After TLR4/MD2 engagement by LPS, three independent pathways are triggered (NF_{KB}, p38/JNK1/2 and IRF3). Altogether, these pathways promote the production of either inflammatory or anti-inflammatory cytokines. After HO-1 induction, heme-group is degraded and CO, Fe^{2+} and biliverdin are produced. CO is able to inhibit the activation of (1) NFkB, (2) p38/JNK1/2 and (3) IRF3 signaling pathways. This leads to a decrease in the amount of inflammatory cytokines produced. Still remains unknown whether CO can block the traffic of inflammatory cytokines toward the extracellular milieu (4). In addition, CO has been reported as unable to affect IL-10 production and secretion (5). Inhibitory pathways are represented as gray lines. **B**) Dendritic cells capture soluble extracellular antigens and target these molecules to early endosomes (Rab5⁺). Then, after TLR4/MD2-mediated stimulation of Rab7⁺-vesicles (interlinear arrow), these compartments fuse with early endosomes forming late endosomes (Rab7⁺). Next, Lamp1⁺ lysosomes fuse with Rab7⁺-late endosomes in a TLR4/MD2-dependent manner (interlinear arrow) to produce antigen degradation and loading of peptides over class I and II MHCs (pMHCs). pMHCs are transported to the surface of DCs to further prime T cells. After heme degradation and CO production by HO-1, CO impairs the fusion of $Rab7^+$ -late endosomes with Lamp1⁺-lysosomes, thus reducing the target of the soluble antigen-derived peptides to MHCs. As a consequence, antigens are stalled and degraded in Rab7⁺-vesicles but without chances to be loaded on MHCs molecules (dotted arrow). Class I and II MHCs presentations are finally impaired by CO.

 (HA) -specific $CD8⁺$ T cells primed by CO-treated DCs loaded with an HA-derived peptide to Tg mice expressing HA in the pancreas did not induce diabetes. Instead diabetes was developed when these mice were transferred with HAspecific $CD8⁺$ T cells primed by untreated DCs loaded with HA. These observations support CO treatment as a promising new tool to induce tolerance in immune-mediated diseases [15]. HA-specific $CD8⁺$ T cells primed by CO-treated DCs showed reduced β 1 integrin and reduced ability to lyse isolated pancreatic islets [15]. The generation of tolerogenic DCs by pulsing them with CO gas, protected mice against an immune system-mediated disease, such as type1 diabetes.

 Although much advance has been made in understanding the effects of CO over DCs, the mechanism by which CO causes tolerance and reduces T cell activation by DCs remains unknown. As in autoimmune diseases, DCs may take up self-antigens and present them to self-reactive T cells, CO treatment could reduce intracellular antigen trafficking and modulate DC maturation to reduce T cell activation, as discussed above [140].

Suppression of Macrophage Function by HO-1 and CO

 Similar to DCs, HO-1 induction in macrophages might revert inflammatory processes. It has been shown that macrophages respond to TLR ligands, such as LPS, via the TLR4/MD2 complex [160, 161]. After LPS recognition, these cells secrete elevated amounts of inflammatory cytokines, including TNF- α , IL-6 and IL-12 [160, 161]. Also in acute phases of infection by Gram-negative bacteria, where LPS reaches abnormal levels in tissues, macrophages, supported by DCs activity, release inflammatory cytokines in an uncontrolled manner [160-162] leading even to death [163]. Thus, down regulation of the inflammatory capacity of macrophages is a promising approach to address bacteriadriven sepsis.

 It has been reported that in macrophages the response to LPS challenge can be suppressed by CO upon HO-1 induction [164]. Interestingly, the authors of this work described that CO has the capacity to decrease the secretion of TNF- α , IL-1 β and (MIP)-1 β by macrophages after LPS challenge (Fig. **2A**). In contrast, CO administration increased the secretion of the anti-inflammatory cytokine IL-10 by macrophages, which is similar to what was observed for DCs. Indeed, authors reported that the mechanism underlying this effect of CO involves the MAPK signaling pathway, because mice lacking these kinases were unable to respond to CO treatment [164]. Consistent with this notion is the observation that CO requires the activation of the JNK signaling

pathway to reduce inflammation in a mouse model for sepsis in mice [37]. Contrary to wild-type, mice lacking either *Jnk1* or *Jnk2* genes were unable to prevent inflammatory cytokine production in response to LPS after CO administration. Wild type animals drastically decreased the secretion of both IL-6 and IL-1 β in the presence of CO. However, *Jnk* knock-out animals failed to respond to CO and cytokine secretion remained unaltered. *In vitro*, this correlated with reduced secretion of these cytokines by LPS-pulsed CO-treated macrophages. Thus, CO exerts its immune-modulatory mechanism by interfering with the JNK signaling pathway [37].

 It has been shown that CO induces IL-10 secretion and that IL-10 has the capacity to induce HO-1 expression in macrophages by activating the p38 signaling pathway [165], which could be beneficial during inflammatory responses. Indeed, *in vivo* administration of IL-10 protects mice from LPS-mediated sepsis by inducing HO-1 expression. Although there is no direct experimental evidence showing that macrophages are the immune cells mediating this protective effect during sepsis, it is likely that innate cells, such as macrophages and DCs, could play an important role in these events. In fact, although it has been shown that DCs are more effective at secreting inflammatory mediators upon pathogen-associated molecular patterns (PAMPs) stimulation, the role of macrophages during this process cannot be ruled out [162, 166].

 The mechanisms responsible for the capacity of HO-1 and CO to reduce macrophage inflammatory response to LPS remain obscure. It has been recently shown that macrophages from mice with cystic fibrosis produce exacerbated amount of inflammatory cytokines when challenged with LPS [167]. In normal macrophages, LPS challenge induces a sustained activation of NF-KB/MAPK signaling pathway, which produces pro-inflammatory cytokines and ROS. This leads to the stress-induced expression of HO-1 and to the recruitment of HO-1 to cell surface with caveolin-1, suppressing TLR4 mediated signaling by CO production [167]. Interestingly, macrophages from cystic fibrosis patients showed a decreased expression of caveolin-1, which recruited less HO-1 to the cell surface leading to sustained NF-KB/MAPK signaling and production of ROS, altering macrophage homeostasis [167]. These data support the importance of HO-1 and CO as protective agents against inflammation and suggest that the proper cellular localization of HO-1 is required to ensure an effective anti-inflammatory response [167].

 Interestingly, in models that use inflammation mediated by both macrophages and pathogens in intestinal diseases, HO-1 induction or CO administration enhanced the killing of Gram negative bacteria, suggesting a different role for this enzyme [168]. Other studies have reported that CO-treated macrophages engulf elevated numbers of Gram-negative bacteria [169] and that CO administration enhances formation of the phagolysosomes [170]. Thus, it is likely that in macrophages, the capture, processing and presentation of antigens could be different from DCs when exposed to CO [140]. Further studies are required to clarify these apparent discrepancies. Studies about the role of HO-1 and CO as anti-inflammatory molecules need to be analyzed in depth for DCs and macrophages, because these mechanisms can be exploited to restrict inflammatory condition during pathogen-driven illness or autoimmune diseases.

Suppression of T Cell Function by HO-1 and CO

It has been reported that both $CD4^+CD25^-$ and $CD4^+$ $CD25⁺$ T cells express HO-1 and activation by $CD3/CD28$ antibodies induces the expression of this enzyme, suggesting a possible direct role for HO-1 in T cell function [171]. It has been shown that HO-1 induction as well as CO exposure could abolish IL-2 production and T cell proliferation after CD3/CD28 stimulation [172]. In activated T cells, CO blocks upstream signaling cascades of MEK1, thereby resulting in ERK inactivation and abolishing IL-2 secretion [172]. Thus, HO-1 activity could be associated with reduced T cell activation. Also, pharmacological overexpression of HO-1 in Jurkat T cells prevented Fas-mediated apoptosis triggered by anti-Fas antibodies [173]. Interestingly, it has been shown that HO-1 expression was dramatically increased in Foxp3 transfected Jurkat cells and pharmacological blockade of HO-1 activity abolished the suppressive function of $CD4+CD25+$ regulatory T cells, playing a critical role in maintaining peripheral tolerance and modulating immune responses [174].

 HO-1 would be clinically important as a focal point in the development of strategies to reverse the detrimental effects of T cell-mediated diseases. Also, it has been recently demonstrated that a repressor of HO-1, Bach2, is highly expressed in T cells. Bach2 is up-regulated during T cell development as naive T cells express high Bach2 levels [175]. The endogenous HO-1 substrate heme, promotes Bach2 degradation in B cells in conjunction with effector differentiation to plasma cells [176]. Consistent with this notion, naive murine Bach2^{-/-} (CD62LhiCD44lo) T cells in spleen and lymph node were decreased [175], suggesting that Bach2 is required for the homeostasis of peripheral T cells and might modulate effector memory-related genes and function in naive and effector-memory T cells. Most importantly, Bach2-

mice showed anti-nuclear and anti-double stranded DNA autoantibodies, extensive perivascular and alveolar infiltration by lymphocytes and macrophages in lungs and mild infiltration in gut and stomach $[177]$. Also, Bach 2^{-1} T cells evidenced a bias to Th2 related cytokines, such as IL-4 and IL-10 as well as Blimp-1 and Gata3 [175]. Other studies have shown that Bach2 cDNA transfection into effector memory T cells suppressed effector-memory T cell related genes, suggesting that Bach2 expression contributes to the maintenance of naive T cells. Also, previous work confirmed that Bach2 binds to Hmox1 (HO-1 encoding element) [175] and in Bach^{-/-} T cells, HO-1 mRNA was increased. Further, Bach2-/- mice showed to be defective in immune responses *in vivo* [175]. In addition, it has been reported that naive T cells from Bach2^{-/-} mice showed reduced numbers of $FOXP3^+$ cells and an impaired ability to form $FOXP3^+$ induced-Treg cells upon stimulation in the presence of TGF- β [177]. These studies suggest that Bach2 represses genes associated with effector cell differentiation. Bach2, which could be modulated by heme groups, may limit immune activation, playing a critical role in the maintenance of regulated immunity.

Suppression of B Cell Function by HO-1 and CO

Although there are few reports showing that HO-1 induction may modulate humoral immunity, the role of Bach2 in B/plasma cell modulation has been studied in more detail. HO-1 KO mice show higher serum IgM levels as compared to wild type mice, indicating that HO-1 ant its by- products, or CO/heme levels, would be involved in modulating antibody production [118]. The transcription factor B lymphocyte-induced maturation protein 1 (Blimp-1), a master regulator of plasma cells [178], was induced by heme stimulation in B cell cultures, then promoting plasma cell differentiation by inactivating Bach2, a repressor of Blimp-1 [176]. Also, authors showed that the administration of heme concomitantly with antigen, reduced the production of antigenspecific IgM *in vivo*, suggesting that heme also modulates the early phase of B-cell responses to antigen [176]. Furthermore, it has been shown that Bach2 is required for class switch recombination/somatic hypermutation of immunoglobulin genes [179]. Heme binds to Bach2, inducing its degradation and Bach1/Bach2 redundantly represses the expression of HO-1 in B cells [176], supporting the notion that HO-1/heme/CO are key modulators in humoral immunity. Bach 2^{-1} mice also have reduced numbers of naive T and B cells, which show a rapid differentiation to effector cells, suggesting a major role of this transcription factor in naive cells homeostasis [175].

6. INVOLVEMENT OF HO-1 IN AUTOIMMUNE DIS-EASES

Systemic Lupus Erythematous (SLE)

 Deregulated inflammation is a common feature of chronic autoimmune disease as SLE. The autoimmune response in SLE is mediated by both cells and humoral components of the immune system [180]. SLE is characterized by the presence of autoantibodies against dsDNA, nucleosomes, ribonucleoproteins and other nuclear components [181]. Systemic damage present in SLE is mainly due to immune complexes deposition in blood vessels, followed by granulocyte and complement activation, leading to a wide spread inflammation and tissue injury [119, 120]. Lupus symptoms are accompanied by alterations in the immune system such as the presence of pathogenic B- and T- cells that lead to autorreactive responses to nuclear self-antigens, as well as an increase in inflammatory cytokines producing a major inflammatory status [119, 120, 180]. Despite much progress has been made in developing novel immunotherapies, this has not been the case for the treatment of lupus, most probably due to its complexity, continuing with systemic immunosuppressant agents such as corticosteroids, azathioprine, mycophenolatemofetil and cyclophosphamide [182, 183].

 Recently, our group has reported that SLE patients showed a decreased HO-1 expression in peripheral blood monocytes [18]. Furthermore, mice lacking the $Fc\gamma RIIb$ receptor, a spontaneous lupus murine model, also showed decreased levels of HO-1 in DCs, T CD4⁺ cells and CD11b⁺ cells [13]. In this model, animals suffering lupus displayed elevated numbers of monocytes/granulocyte population in spleen. Here, when lupus prone mice were chronically treated with gaseous CO, monocyte/granulocyte expansion was limited [13]. Interestingly, $Fc\gamma RIIb^{-/-}$ mice in spleen contained less numbers of regulatory $TCD4$ ⁺ $Foxp3$ ⁺ cells than wild type mice, which was restored when animals received CO as an inhalator gas. In addition, histone specific autoantibodies were also decreased after CO-treatment. CO treatment, as well as HO-1 induction, were able to ameliorate the development of proteinuria in this model [13] (Fig. **3A** and **B**). Thus, the inflammatory environment mediated by elevated number of monocytes, neutrophils and self-antigenspecific T cells can be controlled by the expansion of regulatory/anti-inflammatory T cells after administration of CO. In addition, it has been shown that HO-1 induction by hemin administration in other lupus prone mice, MRL/lpr, was able to decrease autoantibodies (IgG anti DNA) production, ameliorate renal glomerulonephritis by decreasing immune complex deposition, and decrease the levels of the proinflammatory cytokine IFN- γ in serum [184].

Rheumatoid Arthritis

The role of HO-1 in rheumatic diseases could be further corroborated in the collagen induced arthritis model. In this experimental model of rheumatoid arthritis, the administration of CO decreased serum anti-collagen antibodies, ameliorated disease activity, and showed lower inflammation and cartilage erosion [185]. In other approach, when the role of endogenous HO-1 level was evaluated in arthritis development using HO-1 deficient mice, $HO-1^{-/-}$ and $HO-1^{+/-}$ mice developed accelerated clinical symptoms of arthritis compared with $HO-1^{+/+}$ animals after intraperitoneal injection of arthritic K/BxN mice serum (Fig. **3C**) [186]. In this work, the authors showed that $HO-1^{+/-}$ mice evidenced more synovial infiltration and accumulation of inflammatory cells in joint spaces and proteoglycan depletion than WT mice. Also, IL-6 and matrix metalloproteinase-3 (MMP-3) were increased in ankles of HO-1^{+/-} and HO-1^{-/-} arthritic mice compared to HO-1^{+/+} group [186]. Complementary, in mice treated with arthritogenic anti-type II collagen antibody plus LPS mixture, the CO exposure ameliorates clinical outcome of arthritis in footpads with a decrease in IL1- β and MCP-1 mRNA expression in the hind paws (Fig. **3D**) [187]. All this data in HO-1 deficient mice highlights HO-1 as a potential target for autoimmune disease therapy as well as a biological disease marker.

 In the human disease, serum HO-1 levels were increased in systemic juvenile idiopathic arthritis during the active and inactive phase of the disease and being higher than levels observed in polyarticular juvenile idiopathic arthritis (p-JIA), Kawasaki disease, SLE or mixed connective tissue disease, promoting serum HO-1 levels as a useful marker for the differential diagnosis of systemic juvenile idiopathic arthritis [188, 189]. These data suggest that HO-1 modulation as well as CO exposure would be a potential strategy for SLE treatment.

Multiple Sclerosis

 Multiple sclerosis (MS) is an autoimmune disease of the central nervous system (CNS) and is characterized by local inflammation, demyelization, and axonal loss. MS has remittent course leading to long-term disability. Although MS has been extensively studied, its etiology remains unknown and it is believed that immunological, environmental, genetic and infectious factors contribute to disease development [190, 191]. Most of current knowledge of immunomodulatory effects of HO-1 in MS was obtained from studies over

Fig. (3). *HO-1/CO treatment reduces immune disorders in systemic autoimmune diseases. Systemic Lupus Erythematosus*, **A**) Spleen $CD11c^+$, $CD11b^+$ and $CD4^+$ T cells express HO-1 in wild-type (WT) mice. Anti-histone IgG levels in serum are low. In the spleen of Fc γ RIIb-^{\prime} mice (suffering lupus, positive for anti-nuclear antibodies) there are decreased levels of HO-1 in CD11c⁺, CD11b⁺ and T CD4⁺ cells. In addition, serum levels of anti-histone IgG are increased. **B**) Lupic $Fc\gamma RIIb^{-/-}$ mice display in spleen an inflammatory syndrome associated to increased amount of Gr1⁺CD11b⁺ inflammatory monocytes, reduced amounts of regulatory T CD4⁺Foxp3⁺ cells and increased levels of antihistone IgG in serum. After chronic gaseous CO treatment, spleen levels of Gr1⁺CD11b⁺are reduced, regulatory CD4⁺Foxp3⁺T cells are increased and the levels of anti-histone IgG in serum are re-established to normal values. *Rheumatoid arthritis*, **C**) Role of HO-1 level in arthritis development using HO-1 deficient mice in the model of arthritic K/BxN mice serum transfer. HO-1^{-/-}and HO-1^{+/-}mice developed accelerated clinical symptoms such as synovial inflammation and proinflammatory cytokines production in compared with $HO-1^{+/+}$ animals after K/BxN mice serum transfer. **D**) Arthritic mice induced by the administration of anti-type II collagen antibody plus LPS mixture improve clinical outcome after CO exposure ameliorates inflammatory infiltrate in footpads with a decreased in IL1- β and MCP-1 expression.

experimental autoimmune encephalomyelitis (EAE) mice model. Both, MS and EAE are characterized by the presence of oxidative stress [192] and lipid oxidation [193]. During acute phases of EAE, HO-1 expression was markedly increased in microglia/macrophages and astrocytes. In CNS lesions from MS patients, HO-1 was mainly expressed in oligodendrocytes [194]. In contrast, it has been reported that peripheral blood mononuclear cells (PBMCs) from MS patients showed a reduced expression of HO-1during disease exacerbation, while corticosteroid treatment led to an increase in HO-1 expression [195] (Fig. **4A**). Interestingly, HO-1 induction by hemin was able to limit EAE development, while HO-1 inhibition by SnMP worsened the disease [196]. The protective effect of HO-1 in EAE was associated with inhibition of MHC class II expression by antigen presenting cells and inhibition of Th1 IFN- γ secretion, increasing Treg, decreasing IL-2, and $CD8⁺$ T cell accumulation, proliferation, and function within the CNS (Fig. **4B**). CO acts on APC to inhibit the expression of MHC-II thus limiting reactivation of pathogenic Th cells within the CNS. Exogenous CO mimicked these effects, suggesting that CO contributes to the protective action of HO-1 [16]. Confirming these results, pre-treatment with CORM-A1 also protects

from EAE development, showing amelioration of clinical score, disease incidence and reduction in the infiltration of polymorphonuclear cells [17, 197].

 In addition, HO-1 may have a partial role in maintaining the integrity of the blood–spinal cord barrier, ameliorating damage due to proinflammatory leukocytes infiltration [198]. Also, HO-1/CO treatment reduces neuroinflammation in murine cerebral malaria, a disease that showed immune mechanism similar to EAE, by maintaining blood brain barrier integrity thus limiting T cell infiltration [199]. Moreover, our group has shown that mice gestated under hypothyroidism, and that in the adulthood have been induced with EAE have higher EAE score, increased demyelination and greater $CD4^+$ and $CD8^+$ infiltrating cells in the spinal cord compared to mice gestated in euthyroidism [1]. Interestingly, the expression of HO-1 is reduced in these mice compared to mice gestated in euthyroidism (data not published). Although the mechanism of HO-1 protection from EAE remains to be unveiled, immunomodulation as well as cytoprotective properties on CNS may have a major role in preventing neuroinflammation disease. These data pose endogenous HO-1 as a major protective factor in EAE, and that HO-1 modulation could become a novel therapy for MS.

Type 1 Diabetes

In type 1 diabetes, β cell damage and vascular injury are partially caused by ROS generation, leading to apoptosis [200-203]. Most of the knowledge about HO-1 protective effects in diabetes is related to vascular and endothelial cell function attributed to heme group degradation and the concomitant biliverdin and CO production. The protective effect of HO-1 may be associated with a decreased infiltration of dendritic cells in the pancreas [202]. HO-1 has a protective effect in pancreatic cells and β -cells by increasing antiapoptotic proteins such as Akt and Bcl-xL, by reducing the pro-oxidant injury, and preventing the development of type 1 diabetes [81]. Hu *et al.* (2007) showed that HO-1 transduction ameliorated destructive insulitis and the incidence of diabetes in mice [204]. This would be associated with a lower T-helper cell (Th1)–mediated immune response, in which IL-2 and IFN- γ producing cells from spleen are decreased, as well as serum levels of these cytokines. Also, $HO-1$ transduction decreased the CD11 c^+ MHC-II⁺ cell population in spleen, suggesting that T cell priming is affected [204] (Fig. **4C**). Interestingly, Nikolic *et al.* showed that CORM-A1 treatment delayed the onset and reduced diabetes symptoms in NOD mice while reestablished insulin levels [86]. Also, this study showed that CORM-A1 treatment reduced both the expression of $IFN-\gamma$ and the proliferative capacity of lymph node cells. Most importantly, CORM-A1 treatment increased the proportion of Treg cells, reduced the numbers of IL-17 and IFN- γ producing cells while increased IL-10 cytokine production [86]. In addition to the anti-inflammatory functions, CORM-A1 prevented islet cells apoptosis by downregulation of cytochrome *c* expression and suppression of caspase 3 activity [86].

 Interestingly, HO-1 would be involved in different immuno-supressive signaling pathways. It has been reported that HO-1 expression would be regulated by PPARs agonists, which are known to improved diabetes in obese diabetic mice [205], consistent with the observed adiponectin increase [65]. Thus, HO-1 could be induced by other molecules that, in conjunction, can work together to decrease inflammation and produce a positive outcome for diabetic mice. The therapeutic utility of inducing HO-1 or CO exposure has yet to be tested in human diabetes.

Experimental Autoimmune Uveitis

 Experimental models of autoimmune uveitis (EAU) have been used to evaluate the therapeutic potential of HO-1 relevant to immune mediated inflammatory diseases of the eye such as Behcet's disease, sympathetic ophthalmia, ocular sarcoidosis and Vogt-Koyanagi-Harada (VKH) disease [206- 208]. Jang *et al.* demonstrated that HO-1 induction by hemin administration to rats immunized with interphotoreceptor retinoid-binding protein (IRBP) showed a lower grade of clinical inflammation than did the IRBP-control group examine with a biomicroscope [208]. Further, authors showed that HO-1 inhibition by the administration of Sn-PP to EAU rats accelerated the onset of clinical symptoms as well as worsening them [208]. Similarly, in an acute model of ocular inflammation, it was reported that hemin administration to LPS-treated rats significantly decreased the number of infiltrating polymorphonuclear cells in the anterior chamber of

the eye [209]. In addition, HO-1 induction by hemin administration reduced iNOS and IL-6 mRNA expression in the iris-ciliary body after LPS injection. Most importantly, authors demonstrated that hemin pretreatment reduced protein, NO, IL-6 and TNF- α levels in Aqueous humor from the anterior chamber after LPS injection [209]. Similarly, Rossi *et al.* showed that hemin administration reduced the amounts of IL-1 β and CXCL8 in ciliary bodies in the endotoxininduced uveitis model under both normal and hyperglycemia conditions highlighting the potential role of HO-1 induction in the treatment of ocular diseases secondary to diabetes [210].

 Clinically important is the fact that PBMCs from patients with active Behcet´s Disease (BD) expressed lower HO-1 mRNA levels than PBMCs from healthy controls [211]. TLR4 is involved in immune cells activation after ligation with specific DAMPS associated with Behcet´s Disease such as HSP60 [212]. Interestingly, Kirino *et al.* reported that PBMCs from BD patients showed an increase in TLR4 mRNA levels as compared to those from healthy controls [211]. Furthermore, the expression of TLR4 mRNA was inversely correlated with HO-1 mRNA levels in PBMCs from active BD patients [211].

7. INVOLVEMENT OF HO-1 IN INFLAMMATORY DISEASES

Inflammatory Bowel Disease

 Inflammatory bowel disease (IBD) is characterized by recurrent and severe gastrointestinal inflammation. As observed in EAE and type 1 diabetes models, CO-exposure improved the clinical score of mice with IBD. It has been demonstrated that CO administration or HO-1 induction expression by heme/CoPP ameliorates tissue destruction and inflammatory infiltrate in intestine of $IL10^{-1}$ spontaneous IBD mice model compared with air-exposed mice in model [213]. Furthermore, authors showed that CO administration and HO-1 induction in $IL-10^{-/-}$ mice showed less weight loss and a reduction in IL12p40 and TNF secretion by explants. Complementary to this studies, the same group demonstrated that both CO treatment and HO-1 induction in IBD TCRa^{-/-} mice reduced colitis, increased IL-10 and decreased IL-4 and IL-17 secretion by colonic cells compared with air-exposed TCRa^{-/-} mice [214]. Similar results with CO/HO-1 were observed in dextran sulfate sodiuminduced (DSS) colitis mice [215, 216]. Authors demonstrated that hemin significantly attenuated DSS induced colitis as evidenced by reducing inflammatory cell infiltration, IL-6 and IL-17 production, epithelia apoptosis and tissue destruction. In addition, HO-1 inhibition by SnPP pretreatment enhanced DSS-induced colitis inflammation, IL-6 production, mucosal bleeding and weight loss [215]. Also, hemin markedly increased Foxp3⁺ Treg cells population in mesenteric lymph node of DSS-induced colitis mice. Ulcerative colitis patients with active disease showed augmented HO-1 levels than patients with quiescent disease or healthy controls by immunohistochemical staining [217]. In contrast, Crohn´s disease patients showed no difference in HO-1 levels compared to healthy controls or patients during inactive phase. All these data promote CO/HO-1 axis as a potential tool for IBD treatment.

Fig. (4). *HO-1/CO treatment reduces immune disorders in organ-specific autoimmune diseases. Multiple Sclerosis*, **A**) MS patients showed a decreased HO-1 expression in PBMCs when disease is active. **B**) HO-1 induction as well as CO treatment in EAE mice ameliorates disease by decreasing MHC-II expression on CD11c⁺cells and modulating cytokine production by T cells such as IL-2. *Type 1 Diabetes*, **C**) HO-1/CO treatment in NOD Type 1 diabetes mice block disease development by decreasing pro-inflammatory cytokines by T cells such as IFN γ and IL-2, insulitis and decreasing CD11c⁺cells infiltration on β -islet areas.

Asthma and Allergy

 Asthma is a chronic disease characterized by airflow obstruction airways inflammation, hyper-reactivity and remodeling. As most of immune mediated diseases, the etiology of asthma is multifactorial, in which Th2 related cytokines play a predominant role. In OVA-sensitized/challenged mice, an allergic airway inflammation model, pretreatment with hemin reduced inflammatory infiltration, including eosinophil infiltration, in bronchoalveolar lavage fluid and decreased levels of serum OVA-specific IgE, compared to OVA mice [218]. Also, authors showed that the administration of hemin increased Foxp3 mRNA expression in the lung and increased the percentage of $CD4^+$ $CD25$ high regulatory T cells in blood compared with OVA mice, suggesting that regulatory T cells play a role in hemin protection of allergic airway inflammation [218, 219]. The protection of hemin in the OVA allergic airway inflammation model was specifically prevented by the administration of HO-1 enzymatic inhibitor SnPP [219]. Also, in the neutrophilic lung inflammation model, transgenic mice for TCR specific for MHC-II restricted OVA peptide exposed to OVA aerosols, hemin administration reduced the total number of neutrophils and lymphocytes with much less tissue injury [220]. In addition, hemin reduced numbers of Th17 cells in spleen, IL-17 levels in bronchoalveolar lavage fluid and Th17-related transcription factor retinoic acid-related orphan receptor γt (ROR γt) mRNA expression in lung tissues, mediators that were enhanced by OVA challenge. In contrast to the effect of hemin over Th17 profile, Th1 and Th2 profiles were not affected by HO-1 induction [220].

 A small study in nonsmoking adolescents with moderate to severe asthma showed that HO-1 nasal epithelial mRNA expression was inversely associated with upper and lower airway clinical outcomes promoting HO-1 as a potential disease marker of airway severity [221].

Hepatitis and Liver Injury

 A therapeutic potential for HO-1 induction in immunemediated liver injury models has also been suggested. Odaka *et al.* showed that hemin treatment prevented the infiltration of inflammatory cells around the liver central vein and hepatocyte necrosis in halothane–hypoxia model rats [222]. In addition, it has been reported that HO-1 inhibition by Sn-MP further increases the proinflammatory cytokine $TNF-\alpha$ and $NF-\alpha$ B activation in carbon tetrachloride-liver injury model rats [223]. Similar cytoprotective effects of HO-1 induction were observed in the acetaminophen-induced hepatotoxicity model [224]. Sass *et al.* showed that HO-1 induction by CoPP reduced hepatocyte apoptosis in various models for liver injury, such as anti-CD95 antibody treatment and LPS or TNF- α administration in D-galactosamine sensitized mice, mainly by inhibiting caspase-3 activity [225]. Further, overexpression of HO-1 by adenoviral gene transfer (injected intravenously) also reduced apoptosis and liver damage after anti-CD95 antibody treatment [225]. In addition, CO exposure prevented liver apoptosis induced by anti-CD95 antibody treatment [225] and CoPP administration reduced the expression of the proinflammatory cytokines $TNF-\alpha$ and $IFN-\gamma$ in LPS/Dgalactosamine treated mice [226]. Although several reports have evaluated the therapeutic potential of HO-1 induction during hepatic injury, the contribution of HO-1/CO for autoimmune hepatitis remains unknown [222-225].

 On the other hand, HO-1 inhibition has been shown to be beneficial during excessive iron accumulation after bile duct ligation in rats preventing fibrosis by decreasing $TGF- β 1,$ TIMP-1 and MMP expression while HO-1 induction increased liver fibrosis [227].

Most importantly, it has been shown that patients with chronic liver disease due to non-alcoholic steatohepatitis, chronic hepatitis C, and liver cirrhosis showed an increased HO-1 mRNA expression than healthy controls [228, 229]. Further, HO-1 expression correlated with lipid peroxidation end-product levels suggesting that HO-1 induction may be part of the response against the oxidative damage resulting critical in disease progression [228]. Thus, HO-1 induction could be a novel therapeutic option for chronic liver disease treatment.

8. ONGOING CLINICAL TRIALS

 Heme has been proved for the treatment of human porphyria [230] and hemin has been approved as a therapeutic agent by the U.S. Food and Drug Administration (FDA) for the treatment of acute porphyrias. Currently, some clinical trials and preclinical toxicology tests suggest that doses of CO gas or CORM taken from experimental mice model therapies, surrounding 20% of COHb levels, showed no side effects in healthy volunteers (ClinicalTrials.gov identifier: NCT00094406). Some clinical trials using CO exposure therapy are initiating and are mainly focused on preventing lung inflammation (ClinicalTrials.gov identifier: NCT000 94406; ClinicalTrials.gov Identifier: NCT00122694) and transplants rejection (ClinicalTrials.gov Identifier: NCT00 531856). In the next years, results of these studies will demonstrate if the CO therapy is a feasible tool for use in routine medical practice and more precisely in autoimmune disease treatment.

9. CONCLUDING REMARKS

 The multiple effects of HO-1 modulation, HO-1 gene therapy as well as CO gas/CORM exposure occur at different points of cellular and molecular biology such as cell activation, proliferation, apoptosis, cytokine response with a major role in redox balance. The efficiency of HO-1/CO axis in modulate proinflammatory response has been proven in different pathological conditions. Although further research must be done to evaluate the effectiveness of HO-1 gene therapy in immune mediated diseases, this strategy may have major clinical impact. The use of CORMs has significantly increased actual knowledge of the anti-inflammatory properties of HO-1 activity, neglecting side effects of hypoxia secondary to inhaled CO gas. Although, HO-1/CO is able to abrogate immune mediated disease in different experimental mice models, translating its efficacy in human autoimmune disease treatment such as SLE, MS and type 1 diabetes is a worthy challenge.

CONFLICT OF INTEREST

 The author(s) confirm that this article content has no conflicts of interest.

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REFERENCES

[1] Albornoz EA, Carreno LJ, Cortes CM, *et al.* Gestational Hypothyroidism Increases the Severity of Experimental Autoimmune Encephalomyelitis in Adult Offspring. Thyroid 2013; 23(12): 1627-37.

- [2] Cua DJ, Sherlock J, Chen Y, *et al.* Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. Nature 2003; 421(6924):744-8.
- [3] Rangachari M, Kuchroo VK. Using EAE to better understand principles of immune function and autoimmune pathology. J Autoimmun 2013; 45(0):31-9.
- [4] Sozzani S, Bosisio D, Scarsi M, Tincani A. Type I interferons in systemic autoimmunity. Autoimmunity 2010; 43(3):196-203.
- [5] Moisini I, Huang W, Bethunaickan R, *et al.* The Yaa Locus and IFN- α Fine-Tune Germinal Center B Cell Selection in Murine Systemic Lupus Erythematosus. J Immunol 2012; 189(9):4305-12.
- [6] Kleinau S. The impact of Fc receptors on the development of autoimmune diseases. Curr Pharm Des 2003;9(23):1861-70.
- [7] Santiago-Raber M-L, Amano H, Amano E, et al. Fcy receptordependent expansion of a hyperactive monocyte subset in lupusprone mice. Arthritis Rheum 2009; 60(8):2408-17.
- [8] Abram Clare L, Roberge Gray L, Pao Lily I, Neel Benjamin G, Lowell Clifford A. Distinct Roles for Neutrophils and Dendritic Cells in Inflammation and Autoimmunity in motheaten Mice. Immunity 2013; 38(3):489-501.
- [9] Ryter SW, Alam J, Choi AMK. Heme Oxygenase-1/Carbon Monoxide: From Basic Science to Therapeutic Applications. Physiol Rev 2006; 86(2):583-650.
- [10] Bouche D, Chauveau C, Roussel JC, et al. Inhibition of graft arteriosclerosis development in rat aortas following heme oxygenase-1 gene transfer. Transpl Immunol 2002; 9(2-4):235-8.
- [11] Monu SR, Pesce P, Sodhi K, *et al.* HO-1 Induction Improves The Type-1 Cardiorenal Syndrome in Mice With Impaired Angiotensin II–Induced Lymphocyte Activation. Hypertension 2013; 62(2):310- 6.
- [12] Soares MP, Lin Y, Anrather J, *et al.* Expression of heme oxygenase-1 can determine cardiac xenograft survival. Nat Med 1998; $4(9)$:1073-7.
- [13] Mackern-Oberti JP, Llanos C, Carreno LJ, *et al.* Carbon monoxide exposure improves immune function in lupus prone mice. Immunology 2013; 140(1):123-32.
- [14] Hu CM, Lin HH, Chiang MT, Chang PF, Chau LY. Systemic expression of heme oxygenase-1 ameliorates type 1 diabetes in NOD mice. Diabetes 2007; 56(5):1240-7.
- [15] Simon T, Pogu S, Tardif V, et al. Carbon monoxide-treated dendritic cells decrease beta1-integrin induction on CD8(+) T cells and protect from type 1 diabetes. Eur J Immunol 2013; 43(1):209-18.
- [16] Chora AA, Fontoura P, Cunha A, *et al.* Heme oxygenase-1 and carbon monoxide suppress autoimmune neuroinflammation. J Clin Invest 2007; 117(2):438-47.
- [17] Fagone P, Mangano K, Coco M, *et al.* Therapeutic potential of carbon monoxide in multiple sclerosis. Clin Exp Immunol 2012; 167(2):179-87.
- [18] Herrada AA, Llanos C, Mackern-Oberti JP, *et al.* Haem oxygenase 1 expression is altered in monocytes from patients with systemic lupus erythematosus. Immunology 2012; 136(4):414-24.
- [19] Maines MD. Heme oxygenase: function, multiplicity, regulatory mechanisms, and clinical applications. FASEB J 1988; 2(10):2557- 68.
- [20] Kristiansen M, Graversen JH, Jacobsen C, *et al.* Identification of the haemoglobin scavenger receptor. Nature 2001; 409(6817):198- 201.
- [21] Schacter B. Heme catabolism by heme oxygenase: physiology, regulation, and mechanism of action. Semin Hematol 1988; $25(4)$:21
- [22] Abraham NG, Kappas A. Pharmacological and Clinical Aspects of Heme Oxygenase. Pharmacol Rev 2008; 60(1):79-127.
- [23] Trakshel GM, Maines MD. Multiplicity of heme oxygenase isozymes. HO-1 and HO-2 are different molecular species in rat and rabbit. J Biol Chem 1989; 264(2):1323-8.
- [24] Mitani K, Fujita H, Sassa S, Kappas A. Heat shock induction of heme oxygenase mRNA in human Hep 3B hepatoma cells. Biochem Bioph Res Co 1989; 165(1):437-41.
- [25] Nascimento ALTO, Luscher P, Tyrrell RM. Ultraviolet A (320– 380 nm) radiation causes an alteration in the binding of a specific protein/protein complex to a short region of the promoter of the human heme oxygenase 1 gene. Nucleic Acids Res 1993; 21(5):1103-9.
- [26] Maines MD, Trakshel GM, Kutty RK. Characterization of two constitutive forms of rat liver microsomal heme oxygenase. Only

one molecular species of the enzyme is inducible. J Biol Chem 1986; 261(1):411-9.

- [27] Trakshel GM, Kutty RK, Maines MD. Purification and characterization of the major constitutive form of testicular heme oxygenase. The noninducible isoform. J Biol Chem 1986; 261(24):11131-7.
- [28] Kikuchi G, Yoshida T, Noguchi M. Heme oxygenase and heme degradation. Biochem Biophys Res Commun 2005; 338(1):558-67.
- [29] Stout JM, Gousset MU, Drummond HA, Gray W, 3rd, Pruett BE, Stec DE. Sex-specific effects of heme oxygenase-2 deficiency on renovascular hypertension. J Am Soc Hypertens 2012; 7(5):328-35.
- [30] George EM, Hosick PA, Stec DE, Granger JP. Heme Oxygenase Inhibition Increases Blood Pressure in Pregnant Rats. Am J Hypertens 2013; 26(7):924-30.
- [31] Applegate LA, Noël A, Vile G, Frenk E, Tyrrell RM. Two Genes Contribute To Different Extents To The Heme Oxygenase Enzyme Activity Measured In Cultured Human Skin Fibroblasts And Keratinocytes: Implications For Protection Against Oxidant Stress. Photochem Photobiol 1995; 61(3):285-91.
- [32] Kutty RK, Kutty G, Wiggert B, Chader GJ, Darrow RM, Organisciak DT. Induction of heme oxygenase 1 in the retina by intense visible light: suppression by the antioxidant dimethylthiourea. Proc Nat Acad Sci USA 1995; 92(4):1177-81.
- [33] Dwyer BE, Nishimura RN, Lu S-Y. Differential expression of heme oxygenase-1 in cultured cortical neurons and astrocytes determined by the aid of a new heme oxygenase antibody. Response to oxidative stress. Mol Brain Res 1995; 30(1):37-47.
- [34] Hayashi S, Omata Y, Sakamoto H, *et al.* Characterization of rat heme oxygenase-3 gene. Implication of processed pseudogenes derived from heme oxygenase-2 gene. Gene 2004; 336(2):241-50.
- [35] Bauer M, Bauer I. Heme oxygenase-1: redox regulation and role in the hepatic response to oxidative stress. Antioxid Redox Signal 2002; 4(5):749-58.
- [36] Zhang Y, Jiang G, Sauler M, Lee PJ. Lung endothelial HO-1 targeting *in vivo* using lentiviral miRNA regulates apoptosis and autophagy during oxidant injury. FASEB J 2013; 27(10):4041-58.
- [37] Morse D, Pischke SE, Zhou Z, *et al.* Suppression of inflammatory cytokine production by carbon monoxide involves the JNK pathway and AP-1. J Biol Chem 2003; 278(39):36993-8.
- [38] Chauveau C, Remy S, Royer PJ, *et al.* Heme oxygenase-1 expression inhibits dendritic cell maturation and proinflammatory function but conserves IL-10 expression. Blood 2005; 106(5):1694-702.
- [39] Morse D. The role of heme oxygenase-1 in pulmonary fibrosis. Am J Respir Cell Mol Biol 2003; 29(3 Suppl):S82-6.
- [40] Durante W, Christodoulides N, Cheng K, Peyton KJ, Sunahara RK, Schafer AI. cAMP induces heme oxygenase-1 gene expression and carbon monoxide production in vascular smooth muscle. Am J Physiol-Heart C 1997; 273(1):H317-H23.
- [41] Terry CM, Clikeman JA, Hoidal JR, Callahan KS. TNF-a and IL- 1α induce heme oxygenase-1 via protein kinase C, Ca2+, and phospholipase A2 in endothelial cells. Am J Physiol-Heart C 1999; 276(5):H1493-H501.
- [42] Elbirt KK, Whitmarsh AJ, Davis RJ, Bonkovsky HL. Mechanism of Sodium Arsenite-mediated Induction of Heme Oxygenase-1 in Hepatoma Cells: ROLE OF MITOGEN-ACTIVATED PROTEIN KINASES. J Biol Chem 1998; 273(15):8922-31.
- [43] Alam J, Wicks C, Stewart D, *et al.* Mechanism of Heme Oxygenase-1 Gene Activation by Cadmium in MCF-7 Mammary Epithelial Cells: ROLE OF p38 KINASE AND Nrf2 TRAN-SCRIPTION FACTOR. J Biol Chem 2000; 275(36):27694-702.
- [44] Shan Y, Lambrecht RW, Donohue SE, Bonkovsky HL. Role of Bach1 and Nrf2 in up-regulation of the heme oxygenase-1 gene by cobalt protoporphyrin. FASEB J 2006; 20(14):2651-3.
- [45] Kwak M-K, Wakabayashi N, Itoh K, Motohashi H, Yamamoto M, Kensler TW. Modulation of Gene Expression by Cancer Chemopreventive Dithiolethiones through the Keap1-Nrf2 Pathway: IDENTIFICATION OF NOVEL GENE CLUSTERS FOR CELL SURVIVAL. J Biol Chem 2003; 278(10):8135-45.
- [46] Igarashi K, Hoshino H, Muto A, *et al.* Multivalent DNA binding complex generated by small Maf and Bach1 as a possible biochemical basis for beta-globin locus control region complex. J Biol Chem 1998; 273(19):11783-90.
- [47] Ogawa K, Sun J, Taketani S, *et al.* Heme mediates derepression of Maf recognition element through direct binding to transcription repressor Bach1. EMBO J 2001; 20(11):2835-43.
- [48] Eisenstein RS, Garcia-Mayol D, Pettingell W, Munro HN. Regulation of ferritin and heme oxygenase synthesis in rat fibroblasts by

different forms of iron. Proc Nat Acad Sci USA 1991; 88(3):688- 92.

- [49] Panahian N, Yoshiura M, Maines MD. Overexpression of Heme Oxygenase-1 Is Neuroprotective in a Model of Permanent Middle Cerebral Artery Occlusion in Transgenic Mice. J Neurochem 1999; 72(3):1187-203.
- [50] Turkseven S, Kruger A, Mingone CJ, Kaminski P, Inaba M, Rodella LF, *et al.* Antioxidant mechanism of heme oxygenase-1 involves an increase in superoxide dismutase and catalase in experimental diabetes. Am J Physiol-Heart C 2005; 289(2):H701-H7.
- [51] Parfenova H, Basuroy S, Bhattacharya S, *et al.* Glutamate induces oxidative stress and apoptosis in cerebral vascular endothelial cells: contributions of HO-1 and HO-2 to cytoprotection. Am J Physiol-Heart C 2006; 290(5):C1399-C410.
- [52] Zhang X, Shan P, Alam J, Fu X-Y, Lee PJ. Carbon Monoxide Differentially Modulates STAT1 and STAT3 and Inhibits Apoptosis via a Phosphatidylinositol 3-Kinase/Akt and p38 Kinasedependent STAT3 Pathway during Anoxia-Reoxygenation Injury. J Biol Chem 2005; 280(10):8714-21.
- [53] Arruda MA, Rossi AG, de Freitas MS, Barja-Fidalgo C, Graça-Souza AV. Heme Inhibits Human Neutrophil Apoptosis: Involvement of Phosphoinositide 3-Kinase, MAPK, and NF-KB. J Immunol 2004; 173(3):2023-30.
- [54] Kahlo K, Fill Malfertheiner S, Ignatov T, *et al.* HO-1 As Modulator of the Innate Immune Response in Pregnancy. Am J Reprod Immunol 2013; 70(1):24-30.
- [55] Coito AJ, Buelow R, Shen X-D, *et al.* Heme oxygenase-1 gene transfer inhibits inducible nitric oxide synthase expression and protects genetically fat Zucker rat livers from ischemia-reperfusion injury1. Transplantation 2002; 74(1):96-102.
- [56] Li M-H, Cha Y-N, Surh Y-J. Carbon monoxide protects PC12 cells from peroxynitrite-induced apoptotic death by preventing the depolarization of mitochondrial transmembrane potential. Biochem Bioph Res Co 2006; 342(3):984-90.
- [57] Sandouka A, Fuller BJ, Mann BE, Green CJ, Foresti R, Motterlini R. Treatment with CO-RMs during cold storage improves renal function at reperfusion. Kidney Int 2006; 69(2):239-47.
- [58] Tayem Y, Johnson TR, Mann BE, Green CJ, Motterlini R. Protection against cisplatin-induced nephrotoxicity by a carbon monoxide-releasing molecule. Am J Physiol-Renal 2006; 290(4):F789- F94.
- [59] Kushida T, LiVolti G, Goodman AI, Abraham NG. TNF-amediated cell death is attenuated by retrovirus delivery of human heme oxygenase-1 gene into human microvessel endothelial cells. Transplantation Proc 2002;34(7):2973-8.
- [60] Di Noia MA, Van Driesche S, Palmieri F, *et al.* Heme Oxygenase-1 Enhances Renal Mitochondrial Transport Carriers and Cytochrome c Oxidase Activity in Experimental Diabetes. J Biol Chem 2006; 281(23):15687-93.
- [61] Turkseven S, Drummond G, Rezzani R, *et al.* Impact of silencing HO-2 on EC-SOD and the mitochondrial signaling pathway. J Biol Chem 2007; 100(4):815-23.
- [62] Zhao M, Liu Y, Bao M, Kato Y, Han J, Eaton JW. Vascular smooth muscle cell proliferation requires both p38 and BMK1 MAP kinases. Arch Biochem Biophys 2002; 400(2):199-207.
- [63] Öllinger R, Bilban M, Erat A, *et al.* Bilirubin: A Natural Inhibitor of Vascular Smooth Muscle Cell Proliferation. Circulation 2005; 112(7):1030-9.
- [64] Taillé C, El-Benna J, Lanone S, Boczkowski J, Motterlini R. Mitochondrial Respiratory Chain and NAD(P)H Oxidase Are Targets for the Antiproliferative Effect of Carbon Monoxide in Human Airway Smooth Muscle. J Biol Chem 2005; 280(27):25350-60.
- [65] L'Abbate A, Neglia D, Vecoli C, *et al.* Beneficial effect of heme oxygenase-1 expression on myocardial ischemia-reperfusion involves an increase in adiponectin in mildly diabetic rats. Am J Physiol-Heart C 2007; 293(6):H3532-H41.
- [66] Dong D-L, Zhang Y, Lin D-H, *et al.* Carbon Monoxide Stimulates the Ca2+–Activated Big Conductance K Channels in Cultured Human Endothelial Cells. Hypertension 2007; 50(4):643-51.
- [67] Wang R, Wang Z, Wu L. Carbon monoxide-induced vasorelaxation and the underlying mechanisms. Brit J Pharmacol 1997; 121(5):927-34.
- [68] Coceani F, Kelsey L, Seidlitz E. Carbon monoxide-induced relaxation of the ductus arteriosus in the lamb: evidence against the prime role of guanylyl cyclase. B Brit J Pharmacol 1996; 118(7):1689-96.
- [69] Verma A, Hirsch D, Glatt C, Ronnett G, Snyder S. Carbon monoxide: a putative neural messenger. Science 1993; 259(5093):381-4.
- [70] Yoshinaga T, Sassa S, Kappas A. Purification and properties of bovine spleen heme oxygenase. Amino acid composition and sites of action of inhibitors of heme oxidation. J Biol Chem 1982; 257(13):7778-85.
- [71] Sardana MK, Kappas A. Dual control mechanism for heme oxygenase: tin(IV)-protoporphyrin potently inhibits enzyme activity while markedly increasing content of enzyme protein in liver. Proc Nat Acad Sci 1987; 84(8):2464-8.
- [72] Grosser N, Abate A, Oberle S, *et al.* Heme oxygenase-1 induction may explain the antioxidant profile of aspirin. Biochem Biophy Res Commun 2003; 308(4):956-60.
- [73] Stocker R, Perrella MA. Heme Oxygenase-1: A Novel Drug Target for Atherosclerotic Diseases? Circulation 2006; 114(20):2178-89.
- [74] Scapagnini G, Foresti R, Calabrese V, Stella AMG, Green CJ, Motterlini R. Caffeic Acid Phenethyl Ester and Curcumin: A Novel Class of Heme Oxygenase-1 Inducers. Mol Pharmacol 2002; 61(3):554-61.
- [75] Kappas A, Drummond GS, Munson DP, Marshall JR. Sn-Mesoporphyrin Interdiction of Severe Hyperbilirubinemia in Jehovah's Witness Newborns as an Alternative to Exchange Transfusion. Pediatrics 2001; 108(6):1374-7.
- [76] Kappas A. A Method for Interdicting the Development of Severe Jaundice in Newborns by Inhibiting the Production of Bilirubin. Pediatrics 2004; 113(1):119-23.
- [77] Sarady-Andrews JK, Liu F, Gallo D, *et al.* Biliverdin administration protects against endotoxin-induced acute lung injury in rats. Am J Physiol-Lung C 2005; 289(6):L1131-L7.
- [78] Fondevila C, Shen X-D, Tsuchiyashi S, *et al.* Biliverdin therapy protects rat livers from ischemia and reperfusion injury. Hepatology 2004; 40(6):1333-41.
- [79] Wang H, Lee SS, Dell'Agnello C, *et al.* Bilirubin Can Induce Tolerance to Islet Allografts. Endocrinology 2006; 147(2):762-8.
- [80] Lee SS, Gao W, Mazzola S, *et al.* Heme oxygenase-1, carbon monoxide, and bilirubin induce tolerance in recipients toward islet allografts by modulating T regulatory cells. FASEB J 2007; $21(13):3450-7.$
- [81] Li M, Peterson S, Husney D, *et al.* Long-Lasting Expression of HO-1 Delays Progression of Type I Diabetes in NOD Mice. Cell Cycle 2007; 6(5):567-71.
- [82] Doberer D, Haschemi A, Andreas M, *et al.* Haem arginate infusion stimulates haem oxygenase-1 expression in healthy subjects. Brit J Pharmacol 2010; 161(8):1751-62.
- [83] Drummond GS, Kappas A. The cytochrome P-450-depleted animal: an experimental model for *in vivo* studies in chemical biology. Proc Nat Acad Sci USA 1982; 79(7):2384-8.
- [84] Lin JHC, Villalon P, Martasek P, Abraham NG. Regulation of heme oxygenase gene expression by cobalt in rat liver and kidney. Eur J Biochem 1990; 192(3):577-82.
- [85] Abraham NG, Lin JHC, Schwartzman ML, Levere RD, Shibahara S. The physiological significance of heme oxygenase. Int J Biochem 1988; 20(6):543-58.
- [86] Nikolic I, Saksida T, Mangano K, *et al.* Pharmacological application of carbon monoxide ameliorates islet-directed autoimmunity in mice via anti-inflammatory and anti-apoptotic effects. Diabetologia 2014; 57(5):980-90.
- [87] Gozzelino R, Jeney V, Soares MP. Mechanisms of Cell Protection by Heme Oxygenase-1. Ann Rev Pharmacol Toxicol 2010; 50(1):323-54.
- [88] Motterlini R, Otterbein LE. The therapeutic potential of carbon monoxide. Nat Rev Drug Discov 2010; 9(9):728-43.
- [89] Motterlini R, Clark JE, Foresti R, Sarathchandra P, Mann BE, Green CJ. Carbon Monoxide-Releasing Molecules: Characterization of Biochemical and Vascular Activities. Circulation Res 2002; 90(2):e17-e24.
- [90] Vera T, Henegar JR, Drummond HA, Rimoldi JM, Stec DE. Protective Effect of Carbon Monoxide–Releasing Compounds in Ischemia-Induced Acute Renal Failure. J Am Soc Nephrol 2005; 16(4):950-8.
- [91] Rodella L, Lamon BD, Rezzani R, *et al.* Carbon monoxide and biliverdin prevent endothelial cell sloughing in rats with type I diabetes. Free Radic Biol Med 2006; 40(12):2198-205.
- [92] Remy S, Blancou P, Tesson L, *et al.* Carbon monoxide inhibits TLR-induced dendritic cell immunogenicity. J Immunol 2009; 182(4):1877-84.
- [93] Fang J, Sawa T, Akaike T, Greish K, Maeda H. Enhancement of chemotherapeutic response of tumor cells by a heme oxygenase inhibitor, pegylated zinc protoporphyrin. Int J Cancer 2004; 109(1):8.
- [94] Shan Y, Pepe J, Lu TH, Elbirt KK, Lambrecht RW, Bonkovsky HL. Induction of the Heme Oxygenase-1 Gene by Metalloporphyrins. Arch Biochem Biophy 2000; 380(2):219-27.
- [95] Wagener FADTG, Toonen EJM, Wigman L, *et al.* HMOX1 promoter polymorphism modulates the relationship between disease activity and joint damage in rheumatoid arthritis. Arthritis Rheum 2008; 58(11):3388-93.
- [96] Cordova EJ, Martinez-Hernandez A, Ramirez-Bello J, *et al.* HMOX1 promoter (GT)n polymorphim is associated with childhood-onset systemic lupus erythematosus but not with juvenile rheumatoid arthritis in a Mexican population. Clin Exp Rheumatol 2012; 30(2):297-301.
- [97] Rueda B, Oliver J, Robledo G, *et al.* HO-1 promoter polymorphism associated with rheumatoid arthritis. Arthritis Rheum 2007; 56(12):3953-8.
- [98] Chen Y-H, Lin S-J, Lin M-W, *et al.* Microsatellite polymorphism in promoter of heme oxygenase-1 gene is associated with susceptibility to coronary artery disease in type 2 diabetic patients. Hum Genet 2002; 111(1):1-8.
- [99] Song F, Li X, Zhang M, *et al.* Association Between Heme Oxygenase-1 Gene Promoter Polymorphisms and Type 2 Diabetes in a Chinese Population. Am J Epidemiol 2009; 170(6):747-56.
- [100] Nakao A, Toyoda Y. Application of carbon monoxide for transplantation. Curr Pharm Biotechnol 2012; 13(6):827-36.
- [101] Braudeau C, Bouchet D, Tesson L, *et al.* Induction of long-term cardiac allograft survival by heme oxygenase-1 gene transfer. Gene Ther 2004; 11(8):701-10.
- [102] Chauveau C, Bouchet D, Roussel JC, et al. Gene transfer of heme oxygenase-1 and carbon monoxide delivery inhibit chronic rejection. Am J Transplant 2002; 2(7):581-92.
- [103] Abraham NG. Therapeutic applications of human heme oxygenase gene transfer and gene therapy. Curr Pharm Des 2003; 9(30):12.
- [104] Laumonier T, Yang S, Konig S, *et al.* Lentivirus Mediated HO-1 Gene Transfer Enhances Myogenic Precursor Cell Survival After Autologous Transplantation in Pig. Mol Ther 2007; 16(2):404-10.
- [105] Abraham NG, da Silva JL, Lavrovsky Y, *et al.* Adenovirusmediated heme oxygenase-1 gene transfer into rabbit ocular tissues. Inves Ophthalmol Vis Sci 1995; 36(11):2202-10.
- [106] Abraham NG, Jiang S, Yang L, *et al.* Adenoviral Vector-Mediated Transfer of Human Heme Oxygenase in Rats Decreases Renal Heme-Dependent Arachidonic Acid Epoxygenase Activity. J Pharmacol Exp Ther 2000; 293(2):494-500.
- [107] Ferenbach DA, Ramdas V, Spencer N, *et al.* Macrophages Expressing Heme Oxygenase-1 Improve Renal Function in Ischemia/Reperfusion Injury. Mol Ther 2010; 18(9):1706-13.
- [108] Shen X-D, Ke B, Uchida Y, et al. Native macrophages genetically modified to express heme oxygenase 1 protect rat liver transplants from ischemia/reperfusion injury. Liver Transpl 2011; 17(2):201- 10.
- [109] Sabaawy HE, Zhang F, Nguyen X, *et al.* Human Heme Oxygenase-1 Gene Transfer Lowers Blood Pressure and Promotes Growth in Spontaneously Hypertensive Rats. Hypertension 2001; 38(2):210- 5.
- [110] Zenclussen ML, Anegon I, Bertoja AZ, *et al.* Over-expression of heme oxygenase-1 by adenoviral gene transfer improves pregnancy outcome in a murine model of abortion. J Reproductive Immunol 2006; 69(1):35-52.
- [111] Zenclussen AC, Zenclussen ML, Ritter T, Volk HD. The use of gene therapy tools in reproductive immunology research. Curr Gene Ther 2005; 5(5):459-66.
- [112] Li Y, Li G, Dong W, *et al.* Transplantation of Rat Islets Transduced With Human Heme Oxygenase-1 Gene Using Adenovirus Vector. Pancreas 2006; 33(3):280-6.
- [113] Braudeau C, Bouchet D, Toquet C, *et al.* Generation of Heme Oxygenase-1-Transgenic Rats. Exp Biol Med 2003; 228(5):466-71.
- [114] Maines MD, Polevoda B, Coban T, *et al.* Neuronal Overexpression of Heme Oxygenase-1 Correlates with an Attenuated Exploratory Behavior and Causes an Increase in Neuronal NADPH Diaphorase Staining. J Neurochem 1998; 70(5):2057-69.
- [115] Araujo JA, Meng L, Tward AD, *et al.* Systemic Rather Than Local Heme Oxygenase-1 Overexpression Improves Cardiac Allograft Outcomes in a New Transgenic Mouse. J Immunol 2003; 171(3):1572-80.
- [116] Yachie A, Niida Y, Wada T, et al. Oxidative stress causes enhanced endothelial cell injury in human heme oxygenase-1 deficiency. J Clin Invest 1999;103(1):129-35.
- [117] Poss KD, Tonegawa S. Reduced stress defense in heme oxygenase 1-deficient cells. Proc Nat Acad Sci USA 1997; 94(20):10925-30.
- [118] Kapturczak MH, Wasserfall C, Brusko T, *et al.* Heme Oxygenase-1 Modulates Early Inflammatory Responses: Evidence from the Heme Oxygenase-1-Deficient Mouse. Am J Pathol 2004; 165(3):1045-53.
- [119] Brown EE, Edberg JC, Kimberly RP. Fc receptor genes and the systemic lupus erythematosus diathesis. Autoimmunity 2007; 40(8):567-81.
- [120] Cohen PL. T- and B-cell abnormalities in systemic lupus. J Invest Dermatol 1993; 100(1):69S-72S.
- [121] Inman RD. Immune complexes in SLE. Clin Rheum Dis 1982; 8(1):49-62.
- [122] Lorenz HM, Herrmann M, Kalden JR. The pathogenesis of autoimmune diseases. Scand J Clin Lab Invest Suppl 2001; 235:16-26.
- [123] Itano AA, Jenkins MK. Antigen presentation to naive CD4 T cells in the lymph node. Nat Immunol 2003; 4(8):733-9.
- [124] Banchereau J, Steinman RM. Dendritic cells and the control of immunity. Nature 1998; 392(6673):245-52.
- [125] Steinman RM, Adams JC, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. IV. Identification and distribution in mouse spleen. J Exp Med 1975; 141(4):804-20.
- [126] Benvenuti F, Lagaudriere-Gesbert C, Grandjean I, *et al.* Dendritic cell maturation controls adhesion, synapse formation, and the duration of the interactions with naive T lymphocytes. J Immunol 2004; 172(1):292-301.
- [127] Elluru SR, Vani J, Delignat S, *et al.* Modulation of human dendritic cell maturation and function by natural IgG antibodies. Autoimmun Rev 2008; 7(6):487-90.
- [128] Gil-Torregrosa BC, Lennon-Dumenil AM, Kessler B, *et al.* Control of cross-presentation during dendritic cell maturation. Eur J Immunol 2004; 34(2):398-407.
- [129] Chow A, Toomre D, Garrett W, Mellman I. Dendritic cell maturation triggers retrograde MHC class II transport from lysosomes to the plasma membrane. Nature 2002; 418(6901):988-94.
- [130] Trombetta ES, Ebersold M, Garrett W, Pypaert M, Mellman I. Activation of lysosomal function during dendritic cell maturation. Science 2003; 299(5611):1400-3.
- [131] Kalergis AM. Modulation of T cell immunity by TCR/pMHC dwell time and activating/inhibitory receptor pairs on the antigenpresenting cell. Curr Pharm Des 2003; 9(3):233-44.
- [132] Chirathaworn C, Kohlmeier JE, Tibbetts SA, Rumsey LM, Chan MA, Benedict SH. Stimulation through intercellular adhesion molecule-1 provides a second signal for T cell activation. J Immunol 2002; 168(11):5530-7.
- [133] Harris NL, Ronchese F. The role of B7 costimulation in T-cell immunity. Immunol Cell Biol 1999; 77(4):304-11.
- [134] Salgado FJ, Lojo J, Alonso-Lebrero JL, *et al.* A role for interleukin-12 in the regulation of T cell plasma membrane compartmentation. J Biol Chem 2003; 278(27):24849-57.
- [135] de Vries JE. Immunosuppressive and anti-inflammatory properties of interleukin 10. Ann Med 1995; 27(5):537-41.
- [136] Savina A, Jancic C, Hugues S, *et al.* NOX2 controls phagosomal pH to regulate antigen processing during crosspresentation by dendritic cells. Cell 2006; 126(1):205-18.
- [137] Mashreghi M-F, Klemz R, Knosalla IS, *et al.* Inhibition of Dendritic Cell Maturation and Function Is Independent of Heme Oxygenase 1 but Requires the Activation of STAT3. J Immunol 2008; 180(12):7919-30.
- [138] Jego G, Palucka AK, Blanck J-P, Chalouni C, Pascual V, Banchereau J. Plasmacytoid Dendritic Cells Induce Plasma Cell Differentiation through Type I Interferon and Interleukin 6. Immunity 2003; 19(2):225-34.
- [139] Ronnblom L, Alm G. Systemic lupus erythematosus and the type I interferon system. Arthritis Res Ther 2003; 5(2):68 - 75.
- [140] Tardif V, Riquelme SA, Remy S, *et al.* Carbon monoxide decreases endosome-lysosome fusion and inhibits soluble antigen presentation by dendritic cells to T cells. Eur J Immunol 2013; 43(11):2832-44.
- [141] Burgdorf S, Kautz A, Bohnert V, Knolle PA, Kurts C. Distinct pathways of antigen uptake and intracellular routing in CD4 and CD8 T cell activation. Science 2007; 316(5824):612-6.
- [142] Burgdorf S, Lukacs-Kornek V, Kurts C. The mannose receptor mediates uptake of soluble but not of cell-associated antigen for cross-presentation. J Immunol 2006; 176(11):6770-6.
- [143] Basha G, Omilusik K, Chavez-Steenbock A, *et al.* A CD74 dependent MHC class I endolysosomal cross-presentation pathway. Nat Immunol 2012; 13(3):237-45.
- [144] Dani A, Chaudhry A, Mukherjee P, *et al.* The pathway for MHCIImediated presentation of endogenous proteins involves peptide transport to the endo-lysosomal compartment. J Cell Sci 2004; 117(Pt 18):4219-30.
- [145] Desjardins M. ER-mediated phagocytosis: a new membrane for new functions. Nat Rev Immunol 2003; 3(4):280-91.
- [146] Guermonprez P, Amigorena S. Pathways for antigen cross presentation. Springer Semin Immunopathol 2005; 26(3):257-71.
- [147] Guermonprez P, Saveanu L, Kleijmeer M, Davoust J, Van Endert P, Amigorena S. ER-phagosome fusion defines an MHC class I cross-presentation compartment in dendritic cells. Nature 2003; 425(6956):397-402.
- [148] Wan Y, Wu Y, Zhou J, *et al.* Cross-presentation of phage particle antigen in MHC class II and endoplasmic reticulum markerpositive compartments. Eur J Immunol 2005; 35(7):2041-50.
- [149] Shiratsuchi A, Watanabe I, Takeuchi O, Akira S, Nakanishi Y. Inhibitory effect of Toll-like receptor 4 on fusion between phagosomes and endosomes/lysosomes in macrophages. J Immunol 2004; 172(4):2039-47.
- [150] Yates RM, Russell DG. Phagosome maturation proceeds independently of stimulation of toll-like receptors 2 and 4. Immunity 2005; 23(4):409-17.
- [151] Ganguly D, Haak S, Sisirak V, Reizis B. The role of dendritic cells in autoimmunity. Nat Rev Immunol 2013; 13(8):566-77.
- [152] Beriou G, Moreau A, Cuturi MC. Tolerogenic dendritic cells: applications for solid organ transplantation. Curr Opin Organ Transplant 2012; 17(1):42-7.
- [153] Murphy SP, Porrett PM, Turka LA. Innate immunity in transplant tolerance and rejection. Immunol Rev 2011; 241(1):39-48.
- [154] Kim IK, Bedi DS, Denecke C, Ge X, Tullius SG. Impact of innate and adaptive immunity on rejection and tolerance. Transplantation 2008; 86(7):889-94.
- [155] Clarke HM, Shrivastava S, Motterlini R, Sawle P, Chen D, Dorling A. Donor HO-1 expression inhibits intimal hyperplasia in unmanipulated graft recipients: a potential role for CD8+ T-cell modulation by carbon monoxide. Transplantation 2009; 88(5):653-61.
- [156] Sato K, Balla J, Otterbein L, *et al.* Carbon monoxide generated by heme oxygenase-1 suppresses the rejection of mouse-to-rat cardiac transplants. J Immunol 2001; 166(6):4185-94.
- [157] Hancock WW, Buelow R, Sayegh MH, Turka LA. Antibodyinduced transplant arteriosclerosis is prevented by graft expression of anti-oxidant and anti-apoptotic genes. Nat Med 1998; 4(12):1392-6.
- [158] Kotsch K, Martins PN, Klemz R, *et al.* Heme oxygenase-1 ameliorates ischemia/reperfusion injury by targeting dendritic cell maturation and migration. Antioxid Redox Signal 2007; 9(12):2049-63.
- [159] Schumacher A, Wafula PO, Teles A, *et al.* Blockage of Heme Oxygenase-1 Abrogates the Protective Effect of Regulatory T Cells on Murine Pregnancy and Promotes the Maturation of Dendritic Cells. PLoS One 2012; 7(8):e42301.
- [160] Hoshino K, Takeuchi O, Kawai T, *et al.* Cutting edge: Toll-like receptor 4 (TLR4)-deficient mice are hyporesponsive to lipopolysaccharide: evidence for TLR4 as the Lps gene product. J Immunol 1999; 162(7):3749-52.
- [161] Nomura F, Akashi S, Sakao Y, *et al.* Cutting edge: endotoxin tolerance in mouse peritoneal macrophages correlates with downregulation of surface toll-like receptor 4 expression. J Immunol 2000; 164(7):3476-9.
- [162] Siegemund S, Schutze N, Freudenberg MA, Lutz MB, Straubinger RK, Alber G. Production of IL-12, IL-23 and IL-27p28 by bone marrow-derived conventional dendritic cells rather than macrophages after LPS/TLR4-dependent induction by Salmonella Enteritidis. Immunobiology 2007; 212(9-10):739-50.
- [163] Jawad I, Luksic I, Rafnsson SB. Assessing available information on the burden of sepsis: global estimates of incidence, prevalence and mortality. J Glob Health 2012; 2(1):10404.
- [164] Otterbein LE, Bach FH, Alam J, Soares M, Tao Lu H, Wysk M, *et al.* Carbon monoxide has anti-inflammatory effects involving the mitogen-activated protein kinase pathway. Nat Med 2000; 6(4):422-8.
- [165] Lee TS, Chau LY. Heme oxygenase-1 mediates the antiinflammatory effect of interleukin-10 in mice. Nat Med 2002; 8(3):240-6.
- [166] Zhong J, Yang P, Muta K, *et al.* Loss of Jak2 selectively suppresses DC-mediated innate immune response and protects mice from lethal dose of LPS-induced septic shock. PLoS One 2010; 5(3):e9593.
- [167] Zhang PX, Murray TS, Villella VR, *et al.* Reduced caveolin-1 promotes hyperinflammation due to abnormal heme oxygenase-1 localization in lipopolysaccharide-challenged macrophages with dysfunctional cystic fibrosis transmembrane conductance regulator. J Immunol 2013; 190(10):5196-206.
- [168] Onyiah JC, Sheikh SZ, Maharshak N, *et al.* Carbon monoxide and heme oxygenase-1 prevent intestinal inflammation in mice by promoting bacterial clearance. Gastroenterology 2013; 144(4):789-98.
- [169] Otterbein LE, May A, Chin BY. Carbon monoxide increases macrophage bacterial clearance through Toll-like receptor (TLR)4 expression. Cell Mol Biol 2005; 51(5):433-40.
- [170] Chung SW, Liu X, Macias AA, Baron RM, Perrella MA. Heme oxygenase-1-derived carbon monoxide enhances the host defense response to microbial sepsis in mice. J Clin Invest 2008; 118(1):239-47.
- [171] Pae H-O, Oh G-S, Choi B-M, Chae S-C, Chung H-T. Differential expressions of heme oxygenase-1 gene in $CD25-$ and $CD25+$ subsets of human CD4+ T cells. Biochem Biophys Res Commun 2003; 306(3):701-5.
- [172] Pae H-O, Oh G-S, Choi B-M, et al. Carbon Monoxide Produced by Heme Oxygenase-1 Suppresses T Cell Proliferation via Inhibition of IL-2 Production. J Immunol 2004; 172(8):4744-51.
- [173] Choi B-M, Pae H-O, Jeong Y-R, *et al.* Overexpression of heme oxygenase (HO)-1 renders jurkat T cells resistant to Fas-mediated apoptosis: involvement of iron released by HO-1. Free Radic Biol Med 2004; 36(7):858-71.
- [174] Choi B-M, Pae H-O, Jeong Y-R, Kim Y-M, Chung H-T. Critical role of heme oxygenase-1 in Foxp3-mediated immune suppression. Bioch Biophys Res Comm 2005; 327(4):1066-71.
- [175] Tsukumo S-i, Unno M, Muto A, *et al.* Bach2 maintains T cells in a naive state by suppressing effector memory-related genes. Proc Nat Acad Sci 2013; 110(26):10735-40.
- [176] Watanabe-Matsui M, Muto A, Matsui T, *et al.* Heme regulates Bcell differentiation, antibody class switch, and heme oxygenase-1 expression in B cells as a ligand of Bach2. Blood 2011; 117(20):5438-48.
- [177] Roychoudhuri R, Hirahara K, Mousavi K, *et al.* BACH2 represses effector programs to stabilize Treg-mediated immune homeostasis. Nature 2013; 498(7455):506-10.
- [178] Turner Jr CA, Mack DH, Davis MM. Blimp-1, a novel zinc fingercontaining protein that can drive the maturation of B lymphocytes into immunoglobulin-secreting cells. Cell 1994; 77(2):297-306.
- [179] Muto A, Tashiro S, Nakajima O, et al. The transcriptional programme of antibody class switching involves the repressor Bach2. Nature 2004; 429(6991):566-71.
- [180] Ronnblom L, Elkon KB. Cytokines as therapeutic targets in SLE. Nat Rev Rheumatol 2010; 6(6):339-47.
- [181] Rekvig OP, Nossent JC. Anti-double-stranded DNA antibodies, nucleosomes, and systemic lupus erythematosus: A time for new paradigms? Arthritis Rheum 2003; 48(2):300-12.
- [182] Liu Z, Davidson A. Taming lupus[mdash]a new understanding of pathogenesis is leading to clinical advances. Nat Med 2012; 18(6):871-82.
- [183] Houssiau F, Ginzler E. Current treatment of lupus nephritis. Lupus 2008;17(5):426-30.
- [184] Takeda Y, Takeno M, Iwasaki M, *et al.* Chemical induction of HO-1 suppresses lupus nephritis by reducing local iNOS expression and synthesis of anti-dsDNA antibody. Clin Exp Immunol 2004; 138(2):237-44.
- [185] Bonelli M, Savitskaya A, Steiner C, *et al.* Heme oxygenase-1 endproducts carbon monoxide and biliverdin ameliorate murine collagen induced arthritis. Clin Exp Rheumatol 2012; 30:6.
- [186] Brines R, Maicas N, Ferrándiz ML, Loboda A, Jozkowicz A, Dulak J, *et al.* Heme Oxygenase-1 Regulates the Progression of K/BxN Serum Transfer Arthritis. PLoS One 2012; 7(12):e52435.
- [187] Takagi T, Naito Y, Inoue M, *et al.* Inhalation of Carbon Monoxide Ameliorates Collagen-induced Arthritis in Mice and Regulates the Articular Expression of IL-1 β and MCP-1. Inflammation 2009; 32(2):83-8.
- [188] Shimizu M, Yachie A. Compensated inflammation in systemic juvenile idiopathic arthritis: Role of alternatively activated macrophages. Cytokine 2012; 60(1):226-32.
- [189] Takahashi A, Mori M, Naruto T, *et al.* The role of heme oxygenase-1 in systemic-onset juvenile idiopathic arthritis. Mod Rheumatol 2009; 19(3):302-8.
- [190] Frohman EM, Racke MK, Raine CS. Multiple Sclerosis The Plaque and Its Pathogenesis. N Engl J Med 2006; 354(9):942-55.
- [191] Siffrin V, Vogt J, Radbruch H, Nitsch R, Zipp F. Multiple sclerosis – candidate mechanisms underlying CNS atrophy. Trends Neurosci 2010; 33(4):202-10.
- [192] Ruuls SR, Bauer J, Sontrop K, Huitinga I, t Hart BA, Dijkstra CD. Reactive oxygen species are involved in the pathogenesis of experimental allergic encephalomyelitis in Lewis rats. J Neuroimmunol 1995; 56(2):207-17.
- [193] Toshniwal P, Zarling E. Evidence for increased lipid peroxidation in multiple sclerosis. Neurochem Res 1992; 17(2):205-7.
- [194] Stahnke T, Richter-Landsberg C, Stadelmann C, Netzler A, Brück W. Differential upregulation of heme oxygenase-1 (HSP32) in glial cells after oxidative stress and in demyelinating disorders. J Mol Neurosci 2007; 32(1):25-37.
- [195] Fagone P, Patti F, Mangano K, *et al.* Heme oxygenase-1 expression in peripheral blood mononuclear cells correlates with disease activity in multiple sclerosis. J Neuroimmunol 2013; 261(1–2):82-6.
- [196] Liu Y, Zhu B, Luo L, Li P, Paty DW, Cynader MS. Heme oxygenase-1 plays an important protective role in experimental autoimmune encephalomyelitis. NeuroReport 2001; 12(9):1841-5.
- [197] Fagone P, Mangano K, Quattrocchi C, *et al.* Prevention of clinical and histological signs of proteolipid protein (PLP)-induced experimental allergic encephalomyelitis (EAE) in mice by the watersoluble carbon monoxide-releasing molecule (CORM)-A1. Clin Exp Immunol 2011; 163(3):368-74.
- [198] Lin Y, Vreman HJ, Wong RJ, Tjoa T, Yamauchi T, Noble-Haeusslein LJ. Heme oxygenase-1 stabilizes the blood-spinal cord barrier and limits oxidative stress and white matter damage in the acutely injured murine spinal cord. J Cereb Blood Flow Metab 2006; 27(5):1010-21.
- [199] Pamplona A, Ferreira A, Balla J, *et al.* Heme oxygenase-1 and carbon monoxide suppress the pathogenesis of experimental cerebral malaria. Nat Med 2007; 13(6):703-10.
- [200] Baynes JW. Role of Oxidative Stress in Development of Complications in Diabetes. Diabetes 1991; 40(4):405-12.
- [201] Wolff SP. Diabetes mellitus and free radicals: Free radicals, transition metals and oxidative stress in the aetiology of diabetes mellitus and complications. British Medical Bulletin. 1993 January 1, 1993;49(3):642-52.
- [202] Li M, Peterson S, Husney D, *et al.* Interdiction of the diabetic state in NOD mice by sustained induction of heme oxygenase: possible role of carbon monoxide and bilirubin. Antioxid Redox Signal 2007; 9(7):855-63.
- [203] Delmastro MM, Piganelli JD. Oxidative Stress and Redox Modulation Potential in Type 1 Diabetes. Clin Dev Immunol 2011; 2011: 593863.
- [204] Hu C-M, Lin H-H, Chiang M-T, Chang P-F, Chau L-Y. Systemic Expression of Heme Oxygenase-1 Ameliorates Type 1 Diabetes in NOD Mice. Diabetes 2007; 56(5):1240-7.
- [205] Krönke G, Kadl A, Ikonomu E, *et al.* Expression of Heme Oxygenase-1 in Human Vascular Cells Is Regulated by Peroxisome Proliferator-Activated Receptors. Arterioscler Thromb Vasc Biol 2007; 27(6):1276-82.
- [206] Gasparin F, Takahashi BS, Scolari MR, Gasparin F, Pedral LS, Damico FM. Experimental models of autoimmune inflammatory ocular diseases. Arquivos Brasileiros de Oftalmologia 2012; 75:143-7.
- [207] Kijlstra A, Hoekzema R, Lelij Avd, Doekes G, Rothova A. Humoral and cellular immune reactions against retinal antigens in clinical disease. Curr Eye Res 1990; 9(s1):85-9.
- [208] Jang JU, Lee SH, Choi CU, Bahk S-C, Chung HT, Yang YS. Effects of Heme Oxygenase-1 Inducer and Inhibitor on Experimental Autoimmune Uveoretinitis. Korean J Ophthalmol 2007; 21(4):238- 43.
- [209] Ohta K, Kikuchi T, Arai S, Yoshida N, Sato A, Yoshimura N. Protective role of heme oxygenase-1 against endotoxin-induced uveitis in rats. Exp Eye Res 2003; 77(6):665-73.
- [210] Rossi S, Amico M, Capuano A, *et al.* Hyperglycemia in Streptozotocin-Induced Diabetes Leads to Persistent Inflammation and Tis-

sue Damage Following Uveitis Due to Reduced Levels of Ciliary Body Heme Oxygenase-1. Mediators Inflamm 2006; 2006 (4): 60285.

- [211] Kirino Y, Takeno M, Watanabe R, et al. Association of reduced heme oxygenase-1 with excessive Toll-like receptor 4 expression in peripheral blood mononuclear cells in Behcet's disease. Arthritis Res Ther 2008; 10(1):R16.
- [212] Direskeneli H, Saruhan-Direskeneli G. The role of heat shock proteins in Behcet's disease. Clin Exp Rheumatol 2003; 21(4 Suppl 30):S44-8.
- [213] Hegazi RAF, Rao KN, Mayle A, Sepulveda AR, Otterbein LE, Plevy SE. Carbon monoxide ameliorates chronic murine colitis through a heme oxygenase 1–dependent pathway. J Exp Med 2005; 202(12):1703-13.
- [214] Sheikh SZ, Hegazi RA, Kobayashi T, *et al.* An Anti-Inflammatory Role for Carbon Monoxide and Heme Oxygenase-1 in Chronic Th2-Mediated Murine Colitis. J Immunol 2011; 186(9):5506-13.
- [215] Zhong W, Xia Z, Hinrichs D, *et al.* Hemin Exerts Multiple Protective Mechanisms and Attenuates Dextran Sulfate Sodium–induced Colitis. J Pediatr Gastroenterol Nutr 2010; 50(2):132-9.
- [216] Berberat PO, A-Rahim YI, Yamashita K, *et al.* Heme Oxygenase-1-Generated Biliverdin Ameliorates Experimental Murine Colitis. Inflamm Bowel Dis 2005; 11(4):350-9.
- [217] Barton S, Rampton D, Winrow V, Domizio P, Feakins R. Expression of heat shock protein 32 (hemoxygenase-1) in the normal and inflamed human stomach and colon: an immunohistochemical study. Cell Stress Chaperones 2003; 8(4):6.
- [218] Xia Z-W, Zhong W-W, Xu L-Q, *et al.* Heme Oxygenase-1- Mediated CD4+CD25high Regulatory T Cells Suppress Allergic Airway Inflammation. J Immunol 2006; 177(9):5936-45.
- [219] Xia Z-W, Xu L-Q, Zhong W-W, *et al.* Heme Oxygenase-1 Attenuates Ovalbumin-Induced Airway Inflammation by Up-Regulation of Foxp3 T-Regulatory Cells, Interleukin-10, and Membrane-Bound Transforming Growth Factor-β1. Am J Pathol 2007; 171(6):1904-14.
- [220] Zhang Y, Zhang L, Wu J, Di C, Xia Z. Heme Oxygenase-1 Exerts a Protective Role in Ovalbumin-induced Neutrophilic Airway In-

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flammation by Inhibiting Response in the Immune Cells Th17. J Biol Chem 2013; 288(48):34612-26.

- [221] Noel S, Saams J, Ong MJC, *et al.* Opposing effects of nasal epithelial NAD(P)H dehydrogenase quinine 1 and heme oxygenase 1 expression on upper and lower airway symptoms in adolescents with asthma. J Allergy Clin Immunol 2011; 128(2):422-4.e3.
- [222] Odaka Y, Takahashi T, Yamasaki A, *et al.* Prevention of halothaneinduced hepatotoxicity by hemin pretreatment: Protective role of heme oxygenase-1 induction. Biochem Pharmacol 2000; 59(7):871-80.
- [223] Nakahira K, Takahashi T, Shimizu H, *et al.* Protective role of heme oxygenase-1 induction in carbon tetrachloride-induced hepatotoxicity. Biochem Pharmacol 2003; 66(6):1091-105.
- [224] Chiu H, Brittingham JA, Laskin DL. Differential Induction of Heme Oxygenase-1 in Macrophages and Hepatocytes during Acetaminophen-Induced Hepatotoxicity in the Rat: Effects of Hemin and Biliverdin. Toxicol Appl Pharmacol 2002; 181(2):106-15.
- [225] Sass G, Soares MCP, Yamashita K, *et al.* Heme oxygenase-1 and its reaction product, carbon monoxide, prevent inflammationrelated apoptotic liver damage in mice. Hepatology 2003; 38(4):909-18.
- [226] Sass G, Seyfried S, Parreira Soares M, *et al.* Cooperative effect of biliverdin and carbon monoxide on survival of mice in immunemediated liver injury. Hepatology 2004; 40(5):1128-35.
- [227] Wang QM, Du JL, Duan ZJ, Guo SB, Sun XY, Liu Z. Inhibiting heme oxygenase-1 attenuates rat liver fibrosis by removing iron accumulation. World J Gastroenterol 2013; 19(19):2921-34.
- [228] Bessa SS, Mohamed Ali EM, Abd El-Wahab Ael S, Nor El-Din SA. Heme oxygenase-1 mRNA expression in egyptian patients with chronic liver disease. Hepat Mon 2012; 12(4):278-85.
- [229] Malaguarnera L, Madeddu R, Palio E, Arena N, Malaguarnera M. Heme oxygenase-1 levels and oxidative stress-related parameters in non-alcoholic fatty liver disease patients. J Hepatol 2005; 42(4):585-91.
- [230] Kordač V, Kozáková M, Martásek P. Changes of Myocardial Functions in Acute Hepatic Porphyrias. Role of Heme Arginate Administration. Ann Med 1989; 21(4):273-6.